

# Document made available under the Patent Cooperation Treaty (PCT)

International application number: PCT/SE04/001814

International filing date: 06 December 2004 (06.12.2004)

Document type: Certified copy of priority document

Document details: Country/Office: SE  
Number: 0303268-7  
Filing date: 05 December 2003 (05.12.2003)

Date of receipt at the International Bureau: 28 December 2004 (28.12.2004)

Remark: Priority document submitted or transmitted to the International Bureau in compliance with Rule 17.1(a) or (b)



World Intellectual Property Organization (WIPO) - Geneva, Switzerland  
Organisation Mondiale de la Propriété Intellectuelle (OMPI) - Genève, Suisse

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Patentavdelningen

PCT : SE 2004 / 0 0 1 8 1 4

21 -12- 2004

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(71) Sökande                   AngioGenetics Sweden AB, Göteborg SE  
Applicant (s)

(21) Patentansökningsnummer   0303268-7  
Patent application number

(86) Ingivningsdatum           2003-12-05  
Date of filing

Stockholm, 2004-12-09

För Patent- och registreringsverket  
For the Patent- and Registration Office

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Avgift  
Fee

**Title**

Angiogenesis affecting polypeptides, proteins, and compositions, and methods of use thereof.

**Field of the Invention**

The invention relates to polypeptides and proteins encoded thereby which are involved in vasculogenesis and/or angiogenesis. These agents may be targeted when producing materials and methods used in the diagnosis and therapy of angiogenesis-related conditions. The invention further relates to such diagnostic and therapeutic methods and agents.

**Background of the Invention**

Both vasculogenesis, the development of an interactive vascular system comprising arteries and veins, and angiogenesis, the generation of new blood vessels, play a role in embryonic development. In contrast, angiogenesis is limited in a normal adult to the placenta, ovary, endometrium, and sites of wound healing. Angiogenesis, or its absence, plays an important role in the maintenance of a variety of pathological states. Some of these states are characterized by neovascularization, e.g., cancer, diabetic retinopathy, glaucoma, and age related macular degeneration. Others, e.g., stroke, infertility, heart disease, ulcers, and scleroderma, are diseases of angiogenic insufficiency.

Angiogenesis has a number of stages (see, e.g., Zhu and Witte, *Invest New Drugs* 17:195-212, 1999). The early stages of angiogenesis include endothelial cell protease production, migration of cells, and proliferation. The early stages also appear to require some growth factors, with VEGF, TGF-A and selected chemokines all putatively playing a role. Later stages of angiogenesis include population of the vessels with mural cells (pericytes or smooth muscle cells), basement membrane production, and the induction of vessel bed specializations. The final stages of vessel formation include what is known as remodelling wherein a forming vasculature becomes a stable, mature vessel bed. Thus, the process is highly dynamic, often requiring coordinated spatial and temporal waves of gene expression.

The complex angiogenesis process is subject to disruption through interference with one or more critical steps, and numerous disease states can result from or be exacerbated by the disruption. Unregulated angiogenesis can cause or worsen disease, for example, ocular neovascularization has been implicated as the most common cause of blindness and underlies the pathology of approximately 20 eye diseases. In certain previously existing conditions such as arthritis, newly formed capillary blood vessels invade the joints and destroy cartilage. In diabetes, new capillaries formed in the retina invade the vitreous humour, causing bleeding and blindness.

In addition to pathologies linked to unregulated angiogenesis, insufficient angiogenesis can also lead to undesirable results. Dead or damaged tissue can lead to numerous pathologies, revascularization of damaged tissues through a healthy, normal angiogenic process is essential to preventing further complications.

Therefore, new targets and treatments that inhibit or enhance angiogenesis are needed. Identification of more key factors involved in any stage of angiogenesis could lead to new diagnostic methods for pathologic conditions related to angiogenesis. Further, elucidation and understanding of the key factors involved in angiogenesis could form the basis for new methods to investigate potential therapies for angiogenesis-related conditions.

#### **Brief Summary of the Invention**

In accordance with the objects outlined above, the present invention discloses eleven nucleic acid sequences and associated proteins which have key roles in vasculogenesis and/or angiogenesis. One object of the present invention is to present approaches for using the eleven novel factors as molecular targets for therapeutic intervention in angiogenesis-related disease states. It is a further object of the present invention to provide materials and methods that can be used to screen compounds for the ability to modulate angiogenesis or angiogenesis-related conditions.

Therapeutics specifically targeting the sequences and proteins identified herein are also provided as agents or compositions which modulate vasculogenesis or angiogenesis.

According to one embodiment of the invention, an isolated nucleic acid molecule according to any one of SEQ ID NO:s 2, 4, 7, 9, 12, 14, 17, 19, 21, 24, 26, 29, 31, 34, 36, 39, 41, 44, 46, 49, 51, 54, and 56 or a fragment or analogue thereof is provided which has the ability to stimulate or inhibit at least one biological activity selected from the group consisting of vasculogenesis, angiogenesis, vascular permeability, endothelial cell proliferation, endothelial cell differentiation, endothelial cell migration, and endothelial cell survival, or an isolated nucleic acid molecule which hybridizes to one of the foregoing sequences under stringent conditions. The invention is also directed to isolated nucleic acid molecules which hybridizes to a compliment of a nucleic acid molecule described above, and an isolated siRNA molecule of at least 19 base pairs targeted to an isolated nucleic acid molecule described above.

According to a further embodiment of the invention, an expression vector comprising one of the novel nucleic acids is provided. The nucleic acid may be operatively associated with a regulatory nucleic acid controlling the expression of the polypeptide encoded by the nucleic acid.

The invention further comprises host cells genetically engineered to contain a nucleic acid as described above, or transfected by an expression vector described above.

According to a further embodiment of the invention, a method of treating an angiogenesis-related condition in a cell, group of cells, or organism is provided, comprising administering an expression vector as described above to the cell, group of cells, or organism.

The invention further comprises antibodies with specific reactivity to the nucleic acid molecules described above. The antibodies may be polyclonal or monoclonal and may further comprise detectable labels, such as fluorescent labels.

According to a further embodiment of the invention, a transgenic, non-human animal is provided which has been genetically engineered to contain a transgene comprising a nucleic acid as described above, and animals which contain and express the transgene.

According to a further embodiment of the invention, a pharmaceutical composition is provided which comprises a nucleic acid sequence as described above. The compound may be administered to a cell, group of cells, or organism to affect vasculogenesis or angiogenesis. The effect may be to increase or decrease vasculogenesis or angiogenesis, and the method may be employed where the cells, group of cells, or organism has an angiogenesis-related disorder. Such angiogenesis-related disorders include cancer, retinopathy, macular degeneration, corneal ulceration, stroke, ischemic heart disease, infertility, ulcers, scleradoma, wound healing, ischemia, ischemic heart disease, myocardial infarction, myocardosis, angina pectoris, unstable angina, coronary arteriosclerosis, arteriosclerosis obliterans, Berger's disease, arterial embolism, arterial thrombosis, cerebrovascular occlusion, cerebral infarction, cerebral thrombosis, cerebral embolism, rubeosis proliferative vitreoretinopathy, chronic inflammation, inflammatory bowel disease, psoriasis, sarcoidosis, and rheumatoid arthritis.

According to a further embodiment of the present invention, an isolated polypeptide comprising a sequence of amino acids substantially corresponding to the amino acid sequence in any one of SEQ ID NO:s 3, 5, 8, 10, 13, 15, 18, 20, 22, 25, 27, 30, 32, 35, 37, 40, 42, 45, 47, 50, 52, 55, and 57 or a fragment or analogue thereof is provided which has the ability to affect angiogenesis in a cell, a group of cells, or an organism.

The invention further comprises host cells genetically engineered to express a polypeptide as described above, as well as antibodies specifically reactive with the polypeptides. The antibody may be polyclonal or monoclonal, and may further comprise a detectable label such as fluorescence.

According to a further embodiment of the invention, a transgenic, non-human animal is provided which has been genetically engineered to contain a transgene comprising a nucleic acid which encodes a polypeptide as described above, and animals that contain and express the transgene.

The invention further provides pharmaceutical compositions comprising an isolated polypeptide as described above. The pharmaceutical composition may be administered to a cell, group of cells, or organism in order to affect vasculogenesis or angiogenesis therein. Vasculogenesis or angiogenesis may be increased or decreased. The cell, group of cells, or organism may have an angiogenesis-related disorder. Representative angiogenesis-related disorders are noted above.

According to a further embodiment of the invention, a method of detecting an angiogenesis-related transcript in a cell in a patient is provided, the method

comprising contacting a biological sample from the patient with a polynucleotide that selectively hybridizes to a sequence at least 80% identical to a sequence according to any one of SEQ ID NO:s 2, 4, 7, 9, 12, 14, 17, 19, 21, 24, 26, 29, 31, 34, 36, 39, 41, 44, 46, 49, 51, 54, and 56, wherein an angiogenesis-related transcript is detected where hybridization is detected. The polynucleotide may comprise a sequence according to any one of SEQ ID NO:s 2, 4, 7, 9, 12, 14, 17, 19, 21, 24, 26, 29, 31, 34, 36, 39, 41, 44, 46, 49, 51, 54, and 56. The biological sample may be a tissue sample, or sample of isolated nucleic acids such as mRNA. According to this method, the nucleic acids may be amplified prior to contacting the biological sample with the polynucleotide. Further, the polynucleotide is immobilized on a solid surface.

According to a further embodiment of the present invention, a method of affecting at least one bioactivity selected from angiogenesis and vasculogenesis in a vertebrate organism is provided, where method comprises the step of administering an effective angiogenesis or vasculogenesis affecting amount of a nucleotide or polypeptide described herein to the organism. The organism may be mammal, such as mice, rats, rabbits, guinea pigs, cats, dogs, pigs, cows, monkeys, and humans. Vasculogenesis or angiogenesis may be enhanced, increased, inhibited, or decreased. This method may be used on organisms that have an angiogenesis-related disorder, such as those disorders described above.

According to a further embodiment of the invention, a transgenic increased angiogenesis laboratory animal is provided which comprises one or more cells in which the expression of a sequence according to any one of SEQ ID NO:s 2, 4, 7, 9, 12, 14, 17, 19, 21, 24, 26, 29, 31, 34, 36, 39, 41, 44, 46, 49, 51, 54, and 56 is upregulated. Transgenic decreased angiogenesis laboratory animals are also provided, which comprise one or more cells in which the expression of a sequence according to any one of SEQ ID NO:s 2, 4, 7, 9, 12, 14, 17, 19, 21, 24, 26, 29, 31, 34, 36, 39, 41, 44, 46, 49, 51, 54, and 56 is down regulated or absent.

The terms "angiogenesis-related condition" or "angiogenesis-related disease (state)" as used herein mean a condition which is marked by either an excess or a deficit of vessel development or which is improved by an increase or decrease in vessel development. Disorders associated with increased angiogenesis include, but are not limited to, cancer (including solid tumors, leukemias, and tumor metastases), benign tumors (including hemangiomas, acoustic neuromas, neurofibromas, trachomas, and pyogenic granulomas), retinopathy, macular degeneration, and corneal ulceration. Pathological states linked to decreased angiogenesis or states which can improve with increased angiogenesis include, but are not limited to, ischemic heart disease, infertility, ulcers, scleradoma, (insufficient) wound healing, ischemia, myocardial infarction, myocarditis, angina pectoris, unstable angina, coronary arteriosclerosis, arteriosclerosis obliterans (ASO), Berger's disease, arterial embolism, arterial thrombosis, cerebrovascular occlusion, cerebral infarction, cerebral thrombosis, cerebral embolism, and stroke. Other angiogenesis related diseases include, but are not limited to, diseases associated with rubeosis (neovascularization of the angle) and diseases caused by the abnormal proliferation of fibrovascular or fibrous tissue including all forms of proliferative vitreoretinopathy, whether or not associated with diabetes, diseases with symptoms of chronic inflammation, such as inflammatory bowel disease, psoriasis, sarcoidosis and rheumatoid arthritis.

A "host cell" is a naturally occurring cell or a transformed cell that contains an expression vector and supports the replication or expression of the expression vector. Host cells may be cultured cells, explants, cells *in vivo*, and the like. Host cells may be prokaryotic cells such as *E. coli*, or eukaryotic cells such as yeast, insect, amphibian, or mammalian cells such as CHO, HeLa, and the like (see, e.g., the American Type Culture Collection catalog or web site, [www.atcc.org](http://www.atcc.org)).

The terms "polypeptide," "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical mimetic of a corresponding naturally occurring amino acid, as well as to naturally occurring amino acid polymers and non-naturally occurring amino acid polymer.

The term "amino acid" refers to naturally occurring and synthetic amino acids, as well as amino acid analogs and amino acid mimetics that function in a manner similar to the naturally occurring amino acids. Naturally occurring amino acids are those encoded by the genetic code, as well as those amino acids that are later modified, e.g., hydroxyproline, gamma-carboxyglutamate, and O-phosphoserine. "Amino acid analogs" refers to compounds that have the same basic chemical structure as a naturally occurring amino acid. Such analogs have modified R groups (e.g., norleucine) or modified peptide backbones, but retain the same basic chemical structure as a naturally occurring amino acid. Amino acid mimetics refers to chemical compounds that have a structure that is different from the general chemical structure of an amino acid, but that functions in a manner similar to a naturally occurring amino acid. Amino acids may be referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUB Biochemical Nomenclature Commission. Nucleotides, likewise, may be referred to by their commonly accepted single-letter codes.

The term "conservative modifications" or "conservatively modified variants" as used herein applies to both amino acid and nucleic acid sequences. With respect to particular nucleic acid sequences, conservatively modified variants refers to those nucleic acids which encode identical or essentially identical amino acid sequences, or where the nucleic acid does not encode an amino acid sequence, to essentially identical sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given protein. For instance, the codons GCA, GCC, GCG and GCU all encode the amino acid alanine. Thus, at every position where an alanine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations are "silent variations," which are one species of conservatively modified variations. Every nucleic acid sequence herein which encodes a polypeptide also describes every possible silent variation of the nucleic acid. One of skill will recognize that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine, and TGG, which is ordinarily the only codon for tryptophan) can be modified to yield a functionally identical molecule. Accordingly, each silent variation of a nucleic acid which encodes a polypeptide is implicit in each described sequence with respect to the expression product, but not with respect to actual probe sequences.

As to amino acid sequences, one of skill will recognize that individual substitutions, deletions or additions to a nucleic acid, peptide, polypeptide, or protein sequence which alters, adds or deletes a single amino acid or a small percentage of amino acids in the encoded sequence is a "conservatively modified variant" where the alteration results in the substitution of an amino acid with a chemically similar amino acid. Conservative substitution tables providing functionally similar amino acids are well known in the art. Such conservatively modified variants are in addition to and do not exclude polymorphic variants, interspecies homologs, and alleles of the invention.

As used herein, "label" or "detectable moiety" refers to a composition detectable by spectroscopic, photochemical, biochemical, immunochemical, chemical, or other physical means. Examples of such labels include  $^{32}\text{P}$ , fluorescent dyes, electron-dense reagents, enzymes, biotin, digoxigenin, or haptens and proteins which can be made detectable, e.g., by incorporating a radiolabel into the peptide or used to detect antibodies specifically reactive with the peptide.

As used herein, "vector" or "expression vector" refers to a nucleic acid construct, generated recombinantly or synthetically, with a series of specified nucleic acid elements that permit transcription of a particular nucleic acid in a host cell. The expression vector can be part of a plasmid, virus, or nucleic acid fragment. Typically, the expression vector includes a nucleic acid to be transcribed operably linked to a promoter.

The phrase "stringent hybridization conditions" as used herein refers to conditions under which sequences will hybridize. Stringent conditions are sequence-dependent and will be different in different circumstances. Skilled workers have access to significant amounts of descriptive material detailing reaction conditions that are appropriate for a given sequence. One example is Innis et al. (1990) PCR Protocols, A Guide to Methods and Applications, Academic Press, Inc. N.Y.).

As used herein, the terms "inhibitors," "activators," and "modulators" of angiogenic polynucleotide and polypeptide sequences and angiogenic activity refer to inhibitory, activating, or modulating molecules. "Inhibitors" are compounds that, e.g., bind to, partially or totally block activity, decrease, prevent, delay activation, inactivate, desensitize, or down regulate the activity or expression of angiogenesis proteins, e.g., antagonists. "Activators" are compounds that increase, open, activate, facilitate, enhance activation, sensitize, agonize, or up regulate angiogenesis protein activity. Inhibitors, activators, or modulators include genetically modified versions of angiogenesis proteins, e.g., versions with altered activity, as well as naturally occurring and synthetic ligands, antagonists, agonists, antibodies, small chemical molecules and the like. Assays for inhibitors and activators include, e.g., expressing the angiogenic protein in vitro, in cells, or cell membranes, applying putative modulator compounds, and then determining the functional effects on activity, as described above.

### **Brief Description of the Figures**

Figure 1 shows a ratio-ratio plot with the  $\log_2$  expression ratio of genes in a cDNA library when compared to embryonic brain endothelial cell portion versus embryonic

brain left over portion and adult brain endothelial cell portion versus adult brain left over portion;

Figure 2 shows a ratio-intensity plot with average intensity versus  $\log_2$  expression ratio of genes in a cDNA library when compared to embryonic brain, heart, and skin endothelial cell versus left over embryonic portions and all adult endothelial cells and left over portions;

Figure 3 schematically depicts microarray data for gene HUP8001J6;

Figure 4 shows a wild type zebrafish embryo at 28 hpf;

Figure 5 shows a HUP8001J6 morphant embryo at 28 hpf;

Figure 6 shows a wild type zebrafish embryo at 56 hpf;

Figure 7 shows a HUP8001J6 morphant embryo at 56 hpf;

Figure 8 shows a wild type zebrafish at 48-56 hpf;

Figure 9 shows a HUP8001J6 morphant embryo at 48-56 hpf;

Figure 10 schematically depicts microarray data for gene HUP8001K17;

Figure 11 shows a HUP8001K17 morphant embryo at 28 hpf;

Figure 12 shows a HUP8001K17 morphant embryo at 56 hpf;

Figure 13 shows a HUP8001K17 morphant embryo at 48-56 hpf;

Figure 14 schematically depicts microarray data for gene HUP8001K21;

Figure 15 shows a HUP8001K21 morphant embryo at 28 hpf;

Figure 16 shows a HUP8001K21 morphant embryo at 56 hpf;

Figure 17 shows a HUP8001K21 morphant embryo at 48-56 hpf;

Figure 18 schematically depicts microarray data for gene HUP8003D24;

Figure 19 shows a HUP8003D24 morphant embryo at 48-56 hpf;

Figure 20 schematically depicts microarray data for gene HUP8004N1;

Figure 21 shows a HUP8004N1 morphant embryo at 28 hpf;

Figure 22 shows a HUP8004N1 morphant embryo at 56 hpf;

Figure 23 shows a HUP8004N1 morphant embryo at 48-56 hpf;

Figure 24 schematically depicts microarray data for gene HUP8010A10;

Figure 25 shows a HUP8010A10 morphant embryo at 28 hpf;

Figure 26 shows a HUP8010A10 morphant embryo at 56 hpf;

Figure 27 shows a HUP8010A10 morphant embryo at 48-56 hpf;

Figure 28 schematically depicts microarray data for gene NOC8003L17;

Figure 29 shows a NOC8003L17 morphant embryo at 28 hpf;

Figure 30 shows a NOC8003L17 morphant embryo at 56 hpf;

Figure 31 shows a NOC8003L17 morphant embryo at 48-56 hpf;

Figure 32 schematically depicts microarray data for gene NOC8009C9;

Figure 33 shows a NOC8009C9 morphant embryo at 28 hpf;

Figure 34 shows a NOC8009C9 morphant embryo at 56 hpf;

Figure 35 shows a NOC8009C9 morphant embryo at 48-56 hpf;

Figure 36 schematically depicts microarray data for gene NOC8009G23;

Figure 37 shows a NOC8009G23 morphant embryo at 28 hpf;

Figure 38 shows a NOC8009G23 morphant embryo at 56 hpf;

Figure 39 shows a NOC8009G23 morphant embryo at 48-56 hpf;

Figure 40 schematically depicts microarray data for gene OJC8003C9;

Figure 41 shows a OJC8003C9 morphant embryo at 28 hpf;

Figure 42 shows a OJC8003C9 morphant embryo at 56 hpf;

Figure 43 shows a OJC8003C9 morphant embryo at 48-56 hpf;

Figure 44 schematically depicts microarray data for gene OJC8009J7;

Figure 45 shows a OJC8009J7 morphant embryo at 28 hpf;

Figure 46 shows a OJC8009J7 morphant embryo at 56 hpf; and

Figure 47 shows a OJC8009J7 morphant embryo at 48-56 hpf.

#### **Detailed Description**

While particular embodiments of the present invention have been illustrated and described, it would be obvious to those skilled in the art that various other changes and modifications can be made without departing from the spirit and scope of the invention. It is therefore intended to cover in the appended claims all such changes and modifications that are within the scope of this invention. Materials, the synthesis of which are not specifically described, are either commercially available or can be prepared using methods well known to those of skill in the art. Except as otherwise noted, all amounts including quantities, percentages, portions, and proportions, are understood to be modified by the word "about", and amounts are not intended to indicate significant digits. Except as otherwise noted, the articles "a", "an", and "the" mean "one or more". All documents cited are, in relevant part, incorporated herein by reference; the citation of any document is not to be construed as an admission that it is prior art with respect to the present invention.

#### Identification of Candidate Genes

A cDNA Library was prepared by collecting the mRNA from purely isolated adult and embryonic mice vascular fragments. The collected mRNA was used to develop cDNA libraries with a broad coverage of genes expressed in the vasculature. Because of the variety in mouse age, the vascular genes represented those active at different times and in different situations in the vasculature.

After creation of the cDNA library, microarrays were created by printing DNA from the cDNA library onto a solid support as known in the art. The microarrays were used to reveal the gene candidates through gene expression profiling. Select tissues from adult and E 18.5 embryonic mice were collected. Tissue selection was based on the amount and purity of RNA available for extraction. After tissues were removed, they were separated into two portions using antibodies or lectins. The first portion, the endothelial cell fraction or EC, contained endothelial cells as well as pericytes and vascular smooth muscle cells which are tightly associated with the vascular fragments. The second portion, also referred to as the left over portion or LO, were those cells remaining after the EC was isolated.

From adult mice, brain and heart tissues were used, both the EC and LO of each. From embryonic mice, brain, heart, and skin, both EC and LO of each, were utilized. The RNA from each fraction was extracted. A common reference RNA (Universal Mouse Reference RNA; Stratagene, Inc.) was employed at this stage for reference purposes. The isolated RNA and the reference RNA were reverse transcribed then amplified twice through two rounds of antisense RNA amplification. The isolated RNA was labelled with the fluorophore cyanine-3 and the reference RNA with cyanine-5.

After labelling, the RNA was assayed through hybridization with the microarrays described above. The hybridized microarrays were scanned and image analysis used to process the experimental data. Normalizing the data through a signal intensity-based normalization algorithm allowed for statistical evaluation of differentially

expressed genes. Genes exhibiting differential expression were selected for further analysis.

#### Selection of Genes with Differential Expression

Using data collected as described above, certain genes were designated as selectively expressed in blood vessels. This was based on comparisons between adult and embryonic EC and LO values. Figure 1 shows a ratio-ratio plot of the data values obtained through comparisons between embryonic brain EC and LO genes and between adult brain EC and LO genes. A comparison of total adult EC with total embryonic EC was also conducted, data not shown. Data points represented with 'DT1 candidates' or 'DT2 candidates' were generally upregulated ( $>0 \log_2$  expression ratio).

Other genes were designated as selectively expressed during angiogenesis through a different comparison of data. The total embryonic EC portion (i.e., brain, heart, and skin EC portions) was compared to all remaining tissues, including the total embryonic LO portion and all adult RNA (EC and LO of both brain and heart). Figure 2 shows a ratio-intensity plot with the average intensity versus  $\log_2$  expression ratio of all genes. The data points marked DT3 candidates and DT4 candidates are those genes shown to be up regulated through this selective analysis. A comparison was also undertaken to analyze all EC portions versus all LO portions, data not shown.

A total of eleven genes of interest were selected for further analysis. Specific expression data for each gene follows throughout and includes a graph showing that gene's expression profile. Tables are used to show the intensity of the microarray signal, the  $\log_2$  expression ratio, p-value, and rank (rank given only for certain fields). The highest rank was awarded to the gene with the highest expression ratio value, the lowest rank was assigned to the gene with the lowest expression value, based on expression ratio values. P-values are given as a value from zero to one. Values close to one indicate a gene that is upregulated, whereas values close to zero indicate a statistically down regulated gene. A statistically significant p-value of 0.05 corresponds to a p-value of 0.05 or 0.95.

For graphs, tables, and text the abbreviation eec/r refers to embryonic EC portions versus all remaining portions, ec/lo refers to all EC portions versus all LO portions, abeclo refers to adult brain EC portion versus adult brain LO portion, abebebc refers to adult brain EC portion versus embryonic brain EC portion, aheclo refers to adult heart EC portion versus adult heart LO portion, ebeclo refers to embryonic brain EC portion versus embryonic brain LO portion, eheclo refers to embryonic heart EC portion versus embryonic heart LO portion, and eseclo refers to embryonic skin EC portion versus embryonic skin LO portion..

#### Evaluation of Selected Genes

Further analysis of selected genes was conducted through knockdown technology in zebrafish. The process involves the use of specific antisense oligonucleotides that block translation from targeted mRNA molecule(s). This allows for inhibition of the gene of interest and allows for a determination of gene function in the development and health of the zebrafish. Zebrafish share genes for vertebrate functions with mammalian vertebrates such as mice and humans. Studies have demonstrated that

organ and/or tissue development in zebrafish can reliably predict effects in humans (See, *inter alia*, Shin and Fishman, From zebrafish to humans: Modular medical models, Ann. Rev. Genomics and Human Genet. 2002; 3: 311-340; Clark *et al.*, An oligonucleotide fingerprint normalized and expressed sequence tag characterized zebrafish library, Genome Res 2001 Sep;11(9):1594-602. Because of their rapid external development, zebrafish embryo development can be easily monitored and analyzed. The presence of a yolk sac helps provide data from the development of a critically deficient embryo further than that possible with other research organisms, such as mice.

To prepare the embryos, the zebrafish homolog of the target gene was identified. Then, a specific morpholino phosphorodiamidate oligonucleotide was designed to match the AUG initiation codon or splice acceptor/donor site of the target gene. To create a stock solution of morpholino, pellets containing 100 nmoles of the phosphorodiamidate oligonucleotides were dissolved in 33.3  $\mu$ l milli-Q water, giving a concentration of 25 mg/ml, and stored at -20°C. To create injection solution, 8  $\mu$ l of the stock solution was added to 92  $\mu$ l of sterile-filtered 1X Danieu buffer (58 mM NaCl, 0.7 mM KCl, 0.4 mM MgSO<sub>4</sub>, 0.6 mM Ca(NO<sub>3</sub>)<sub>2</sub>, 5 mM HEPES, pH 7.6) supplemented with 15mM Tris-Cl, pH 8.0. The 2 mg/ml injection solution was also stored at -20°C.

During injection, the materials and embryos were maintained at approximately 28°C. Injection needles were calibrated so that injection times could optimally be within a range of 100-600 msec. Embryos from the one cell stage to the early eight cell stage were used. The morpholinos were microinjected into the yolk sac. Specific injection volumes, or effective dose of the morpholino, are described below. Typical initial doses included 3, 6, and 12 ng (1.5, 3, and 6 nl, respectively). Toxicity at the 3 nl dose resulted in subsequent doses of 0.5, 1, and 2 ng (1, 2, and 4 nl, respectively). Approximately 40 embryos were injected at each dose level, and approximately 40 embryos were retained as non-injected controls.

After the morpholinos were injected into fertilized egg cells, the embryos engineered to have a knockdown of the specific gene were allowed to develop (See Nasevicius and Ekker, Effective targeted gene 'knockdown' in zebrafish, Nature Genetics vol 26, October 2000.). The embryos were monitored throughout development, both by examining morphology and undertaking specific analysis and assays of developing tissues.

In addition to single morpholino injections, double morpholino injections were performed as well. Specific injection volumes for double injections are described below. At the end of the first post-injection day, with embryos at the blastula or gastrula stage, propyl thioracil (PTU) 2x solution was added to the embryos, doubling their suspension volume. 48 hours post fertilization (hpf) the double injected embryos were fixed with cadherin 5 (cdh5) for *in situ* hybridization.

When 20% or more of the double injected embryos displayed low effect defects in the vasculature observed with cdh5, or when 10% or more of the embryos displayed medium or high effect defects, then microangiopathy and *in situ* hybridization with fli-1, flk-1, flt-4, tie-1, tie-2, and cdh5 were conducted. At least 120 embryos were administered the double morpholino dose, of which at least 100 were harvested at 24

hpf for *in situ* hybridization with the above-noted molecular markers. Remaining embryos were used for microangiopathy.

Data specific to the evaluation of each of the eleven targets are described below. In general, the morphology observations conducted at 24-28 hpf included an indication of whether the embryos exhibited general delay relative to control embryos. Further, cell death type and degree were recorded, general embryo shape and brain morphology were recorded as well. Finally, yolk sac edema, if present, was evaluated and recorded, as was heart morphology.

Also, at approximately 24 hpf, double morpholino embryos were evaluated for *in situ* hybridization of fli-1, flk-1, flt-4, tie-1, tie-2, and cdh5. Overall morphology and the degree of reduction of staining in the intersegmental vessels as compared to control embryos, correlating to a percentage of lost expression, were noted. Those embryos showing a loss of 1-35% of intersegmental expression were considered to have a low effect, those embryos showing a loss of 36-70% of intersegmental expression were considered to have a medium effect, and those embryos showing a loss of 71-100% of intersegmental expression were considered to have a high effect.

At 48-56 hpf various parameters were reviewed and recorded, such as general embryo shape, degree of cell death, blood circulation, and heart morphology. For the embryos fixed with cdh5, staining was evaluated throughout the vasculature as described immediately above.

Microangiopathy was also evaluated at 48hpf in double morpholino embryos. In order to observe the blood vessels, the embryos were transferred into a tricaine solution and the sinus venosa/common cardinal vein was injected with 10  $\mu$ l FITC-Dextran solution (2,000,000 Da, 20 mg/ml).

#### Gene HUP8001J6

The gene having the sequence shown in SEQ ID NO:1 was identified as selectively expressed during angiogenesis based on microarray data, see Figure 3. Specific data are given below in Table 1. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:2) and the human homolog (SEQ ID NO:4) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 3 and 5, respectively.

	intensity	log2 exp ratio	p-value	rank
secf	8,2	0,88		600
secf	8,1	0,80		449
abeclo	7,6	0,60	1,00	
abecelobec	8,3	0,27	0,76	
abeclo	7,3	1,17	1,00	
ebeclo	9,1	-0,29	0,23	
ebeclo	8,1	1,21	1,00	
eseclo	8,1	-0,13	0,25	

Table 1: Expression profile data for gene HUP8001J6.

Based on this expression profile, the gene was further analyzed in zebrafish embryos. Two corresponding zebrafish genes were identified for targeting. Two morpholinos were prepared, sz265 and

sz266, each targeted to one of the zebrafish genes. Six (6) ng of sz265 morpholino and 2 ng of sz266 morpholino were administered to each fertilized egg. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression was analyzed in the assay and results differed somewhat based on the probe used. The probe *fli-1* revealed that 13% of the 16 morphant embryos assayed had high effects, that is, loss of 71-100% of intersegmental expression. The probe *flk-1*, VEGF receptor 2, indicated that 7% of the 15 morphants reviewed had high effects. When analyzed through the probe *tie-1*, all 15 morphants observed were normal. The probe *cdh5*, VE cadherin, indicated that all 23 morphants observed were normal. The probe *flt-4*, VEGF receptor 3, indicated that all 15 morphants observed were normal. And the probe *tie-2* showed all 15 observed morphants as normal.

The following table, Table 2, summarizes this data.

Probe	Number analyzed	Results
<i>fli-1</i>	16	13%H
<i>flk-1</i>	15	7%H
<i>tie-1</i>	15	normal
<i>cdh5</i>	23	normal
<i>flt-4</i>	15	normal
<i>tie-2</i>	15	normal

Table 2: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Both the wild type embryos, used as control, and the morphant embryos observed showed normal morphology. Figures 4 and 5 show the wild type and morphant embryos at 28 hpf, respectively. At 56 hpf the embryos were again observed, a wild type embryo is shown in Figure 6 and a morphant embryo in Figure 7. As highlighted by an arrow in the representative embryo of Figure 7, 82% of the 56 morphant embryos showed severe yolk sac edema. Further, 70% showed severely reduced blood flow and 14% showed reduced intersegmental blood flow.

Additional analyses were conducted on 48-56 hpf morphant embryos. A primary *in situ* hybridization screen with *cdh5* on 22 morphants revealed that 18% had reduced intersegmental expression, at a low effect level, whereas the remaining 82% were normal. Microangiography on 22 morphants was used to locate the presence of FITC-Dextran in various regions of the embryo. No FITC-Dextran was observed in the heart, head, nor was there any leaky vasculature. 14% of the morphants showed high effects of reduced intersegmental vasculature, and another 14% showed low effects of the same. Thus, 28% were affected by some changes to the intersegmental vasculature and 72% appeared normal. A wild type embryo at this time stage is shown in Figure 8, whereas Figure 9 shows a representative of the 48-56 hpf embryos analyzed. The arrow points to an area of reduced intersegmental vasculature.

#### Gene HUP8001K17

The gene having the sequence shown in SEQ ID NO:6 was identified as selectively expressed in blood vessels based on microarray data, see Figure 10. Specific data are given below in Table 3. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:7) and the human homolog (SEQ ID NO:9) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 8 and 10, respectively.

	Intensity	502 exp. ratio	p-value	rank
cecl	7,8	-0,11		6732
cecl	7,8	0,59		804
abcl	7,9	3,52	1,00	
abcebec	8,4	2,10	1,00	
ancl	7,5	-1,05	0,00	
escl	8,8	2,15	1,00	
shcl	7,5	-0,07	0,42	
Escl	7,5	-0,56	0,01	

Table 3: Expression profile data for gene HUP8001K17.

Based on this expression profile, the gene was further analyzed in zebrafish embryos. One corresponding zebrafish gene was identified for targeting. Two different morpholinos were prepared, sz143 and sz144, each targeted to the zebrafish gene. Different amounts of morpholinos were administered as described below. The predetermined amount of each morpholino was administered to each fertilized egg. The embryos were allowed to develop. At 24 hpf secondary *in situ* hybridization screens with six different probes were conducted.

One screen was performed on embryos that received 1 ng of sz143 morpholino and 4 ng of sz144 morpholino. Four probes specifically selected to analyze axial and intersegmental vessel expression revealed the following: using the *fli-1* probe, 6% of the 17 embryos analyzed had medium intersegmental expression effects. Another 18% had high effects. The probe *flk-1*, VEGF receptor 2, indicated that 20% of the 15 morphants reviewed had medium and 13% had high effects. When analyzed through the probe *tie-1*, 63% of the 16 morphants observed had high effects. The probe *cdh5*, VE cadherin, indicated that 4% of the 24 morphants observed had medium effects, and another 4% had high effects. The probe *flt-4*, VEGF receptor 3, indicated that all 14 morphants observed were normal, and the probe *tie-2* showed all 18 observed morphants as normal.

The other screen was performed on embryos that received 1.5 ng of sz143 morpholino and 6 ng of sz144 morpholino. The probe *fli-1* indicated that 42% of the 12 morphants analyzed had high effects. The *flk-1* probe demonstrated that 23% of the 13 morphants observed had high effects. The *tie-1* probe revealed 54% of 13 morphants had high effects. The probe *cdh5* indicated that 15% of 27 morphants had medium effects and another 11% had high effects. The probe *flt-4* indicated that all 14 morphants observed were normal. And the probe *tie-2*, showed all 18 observed morphants as normal.

The following Table 4 summarizes the foregoing data.

Probe	Morphants with 1 ng sz143, 4 ng sz144		Morphants with 1.5 ng sz143, 6 ng sz144	
	Number analyzed	Results	Number analyzed	Results
<i>fli-1</i>	17	6%M, 18%H	12	42%H
<i>flk-1</i>	15	20%M, 13%H	13	23%H
<i>tie-1</i>	16	63%H	13	54%H
<i>cdh5</i>	24	4%M, 4%H	27	15%M, 11%H
<i>flt-4</i>	13	normal	14	normal
<i>tie-2</i>	19	normal	18	normal

Table 4: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. The wild type embryos, used as control, showed normal morphology as expected. As indicated previously, Figure 4 shows a wild type embryo at 28 hpf. The morphant embryos received a 1.5 ng dose of sz143 and a 6 ng dose of sz144, all did not exhibit normal morphology. A representative embryo is shown in Figure 11. Twenty embryos were observed, 50% of them showed a curly down body, indicated by the arrowhead in Figure 11, with yolk tube extension, indicated with a short arrow. Mild cell death was observed in 60% of the embryos, as shown by the long arrow in Figure 11. Finally, 50% of the embryos had yolk cell edema.

At 56 hpf embryos were again observed, for reference a wild type embryo is shown in Figure 6. A morphant at the corresponding stage is shown in Figure 12. Twenty (20) embryos were observed, 90% had a curly down body as shown by the long arrow in Figure 12, with reduced head as indicated by the short arrow. Pericardial edema, shown by the arrowhead, was observed in 90% of the embryos and reduced blood flow was also seen in 90% of the embryos.

Additional analyses were conducted on 48-56 hpf morphant embryos. A primary *in situ* hybridization screen with *cdh5* on 18 morphants which had received 1 ng sz143 and 3 ng sz144 revealed that 28% had reduced intersegmental expression, at a low effect level, with short and curly tails. The remaining 72% were normal. The same *in situ* hybridization screen was conducted using 11 morphants which had received 2 ng sz143 and 6 ng sz144. This revealed that 9% had reduced intersegmental expression, at a low effect level, with very short tails. The remaining 91% were normal.

Microangiography on 26 morphants which had received 1.5 ng of sz143 and 6 ng of sz144 was used to locate the presence of FITC-Dextran in various regions of the embryo. No FITC-Dextran was observed in the heart, but 31% of the embryos had FITC-Dextran in the head and heart. A total of 27% of the morphants had reduced intersegmental vasculature, and leaky vasculature was observed in 35% of the embryos. Only 42% of the embryos appeared normal. The combined percentages are greater than 100% since some embryos exhibited more than one non-normal feature. Figure 13 shows a representative of the 48-56 hpf embryos analyzed. The arrow points to an area of reduced intersegmental vasculature, and the arrowhead indicates a point of leaky vasculature. For reference, a wild type embryo at this time stage is shown in Figure 8. The experimental data reveal that the gene is expressed by scattered cells in many organs, but most clearly seen in the CNS.

#### Gene HUP8001K21

The gene having the sequence shown in SEQ ID NO:11 was identified as selectively expressed during angiogenesis based on microarray data, see Figure 14. Specific data are given below in Table 5. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:12) and the human homolog (SEQ ID NO:14) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 13 and 15, respectively.

	Intensity	log2 exp ratio	p-value	rank
sec11	8,5	0,88		594
ec10	8,4	0,80		442
ape10	8,5	0,71	1,00	
apeo10	8,8	-0,02	0,48	
ape10	8,0	1,15	1,00	
apeo10	9,5	-0,03	0,47	
ape10	8,3	0,72	0,98	
apeo10	8,3	0,49	0,99	

Table 5: Expression profile data for gene HUP8001K21.

Based on this expression profile, the gene was further analyzed in zebrafish embryos. Two corresponding zebrafish genes were identified for targeting. Two morpholinos were prepared, sz257 and sz258, each targeted to one of the zebrafish genes.

Twelve (12) ng of sz257 morpholino and 12 ng of sz258 morpholino were administered to each fertilized egg. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression was analyzed in the assay and results differed somewhat based on the probe used. The probe *fli-1* revealed that all of the 15 morphant embryos assayed were normal. The probe *flk-1* indicated that 13% of the 16 morphants reviewed had low effects and 6% had high effects. When analyzed through the probe *tie-1*, all 15 morphants observed were normal. The probe *cdh5* indicated that all 26 morphants observed were normal. The probe *flt-4* indicated that all 17 morphants observed were normal, the probe *tie-2* showed all 20 observed morphants as normal.

The following table, Table 6, summarizes this data.

Probe	Number analyzed	Results
<i>fli-1</i>	15	normal
<i>flk-1</i>	16	13%L, 6%H
<i>tie-1</i>	15	normal
<i>cdh5</i>	26	normal
<i>flt-4</i>	17	normal
<i>tie-2</i>	20	normal

Table 6: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Figure 15 shows a representative morphant embryo at 28 hpf. As indicated by the arrow, yolk sac edema was observed in 47% of the 55 morphants analyzed. At 56 hpf a total of 53 embryos were observed, a representative morphant embryo is shown in Figure 16. As highlighted by the long arrow, expanded hindbrain was found in 34% of embryos. Yolk sac edema, shown by a short arrow, was also observed in 58% of embryos. An arrowhead points out the location checked for pericardial edema; it was not observed.

Additional analyses were conducted on 48-56 hpf morphant embryos. A primary *in situ* hybridization screen with *cdh5* on 20 morphants showed all as normal. Microangiography on 31 morphants was used to locate the presence of FITC-Dextran in various regions of the embryo, see Figure 17. Of the 31 embryos, 19% had FITC-Dextran in the heart but none had it in the heart and the head. Reduced intersegmental vasculature was seen as indicated by the arrow in Figure 17. High effects were observed in 19% of the embryos, medium effects in 13% and low

effects in 29%. No leaky vasculature was observed. Normal embryos accounted for 19% of the sample.

#### HUP8003D24

The gene having the sequence shown in SEQ ID NO:16 was identified as selectively expressed during angiogenesis based on microarray data, see Figure 18. Specific data are given below in Table 7. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:17) and two human homologs (SEQ ID NO:s19 and 21) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 18, 20, and 22, respectively.

	intensity	log2 exp ratio	p-value	rank
eech	9.1	0,48		2061
eecler	9.0	1,30		149
abecia	8.1	3,14	1,00	
abecobec	8.7	2,26	1,00	
abecia	8.9	0,23	0,85	
ebecia	9.1	0,51	0,90	
enecko	9.5	1,32	1,00	
esecia	9.0	1,84	1,00	

Table 7: Expression profile data for gene HUP8003D24

Based on this expression profile, the gene was further analyzed in zebrafish embryos. Three corresponding zebrafish genes were identified for targeting. Two morpholinos were prepared, sz185 and sz186, which were

targeted to the three zebrafish genes. For the lower dose group, 3ng of sz185 morpholino and 6ng of sz186 morpholino were administered to each fertilized egg. For the double dose group, 6ng of sz185 morpholino and 12ng of sz186 morpholino were administered. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression was analyzed in the assay and results differed somewhat based on the probe used. In the lower dose group, the probe *fli-1* revealed that 18% of the 11 morphant embryos assayed had medium effects, i.e., 36-70% loss of intersegmental expression, and 18% had high effects. The probe *flk-1* indicated that 36% of the 11 morphants reviewed had high effects. The probe *tie-1* indicated that 29% of the 14 morphants observed had high effects. The probe *cdh5* indicated 31% of the 16 morphants observed had high effects, and 31% had medium effects. The probe *flt-4* indicated that all 16 morphants observed were normal, and the probe *tie-2* indicated that all 15 morphants observed were normal.

In the higher dose group, the probe *fli-1* revealed that 33% of the 3 morphant embryos assayed had low effects and 33% had medium effects. The probe *flk-1* indicated that 100%, or both of the 2 morphants reviewed, had medium effects. The probe *cdh5* indicated 100%, all 7 of the morphants observed had high effects. The probe *flt-4* indicated that all 3 morphants observed were normal, and the probe *tie-2* indicated that all 7 morphants observed were normal.

The following Table 8 summarizes the foregoing data.

Probe	Morphants with 3 ng sz185, 6 ng sz186		Morphants with 6 ng sz185, 12 ng sz186	
	Number analyzed	Results	Number analyzed	Results
<i>fli-1</i>	11	18%M, 36%H	3	33%L, 33%M

<i>flk-1</i>	11	36%H	2	100%M
<i>tie-1</i>	14	29%H	n/a	no data
<i>cdh5</i>	16	31%M, 31%H	7	100%H
<i>flt-4</i>	16	normal	3	normal
<i>tie-2</i>	15	normal	7	normal

Table 8: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Cell death was observed in 70% of the 66 embryos observed. Yolk sac edema was observed in 29% of the morphants. At 56 hpf a total of 66 embryos were observed for phenotypic characteristics. Yolk sac edema was observed in 42% of embryos, 35% showed reduced IS blood flow and 26% showed reduced blood flow.

Additional analyses were conducted on 48-56 hpf morphant embryos.

A primary *in situ* hybridization screen with *cdh5* on 21 morphants receiving the lower doses noted above (3ng sz185, 6ng sz86) showed 52% as normal. Low effects of reduced intersegmental expression were seen in 43% of the embryos, and medium effects in 5%. Embryos receiving the double doses (6ng sz185, 12ng sz186), when viewed at the 48-56 hpf stage revealed 74% of the that the 23 embryos observed were normal. Low effects of reduced intersegmental expression were observed in 22% and medium effects in 4% of the embryos.

Microangiography on morphants given the lower dose of morpholinos (3ng sz185, 6ng sz186) was used to locate the presence of FITC-Dextran in various regions of the embryo, see Figure 19. Of the 33 embryos, none had FITC-Dextran in the heart, or the heart and head. Reduced intersegmental vasculature was seen as indicated by the arrow in Figure 19. High effects were observed in 6% of the embryos, medium effects in 15% and low effects in 36%. No leaky vasculature was observed. Normal embryos accounted for 43% of the sample.

The data reveal that the gene is expressed in many locations, such as vessels and epithelial structures in the kidneys as well as in large vessels, megakaryocytes, in heart valves and in the skin epithelium.

#### Gene HUP8004N1

The gene having the sequence shown in SEQ ID NO:23 was identified as selectively expressed during angiogenesis based on microarray data, see Figure 20. Specific data are given below in Table 9. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:24) and the human homolog (SEQ ID NO:26) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 25 and 27, respectively.

	intensity	log2 exp ratio	p-value	rank
eeqr	9.0	1.12		262
eeqr	9.0	0.75		516
abeddo	8.0	0.44	0.79	
abeddo	8.6	-0.80	0.03	
shedo	9.0	-0.47	0.02	
shedo	8.8	1.68	1.00	
eneso	9.3	1.18	1.00	
eneso	9.0	1.22	1.00	

Table 9: Expression profile data for gene HUP8004N1

Based on this expression profile, the gene was further analyzed in zebrafish embryos. Two corresponding zebrafish genes were identified for targeting. Two morpholinos were prepared, sz223 and sz224, each targeted to one of the zebrafish genes. Two dosing strategies were employed. The first dose group received 2ng of sz223 morpholino and 1ng of sz224 morpholino in each fertilized egg. The second dose group received 1ng of sz223 morpholino and 0.5ng of sz224 morpholino in each fertilized egg. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression in embryos from the second dose group (1ng sz223, 0.5ng sz224) was analyzed in the assay and results differed somewhat based on the probe used. The probe *fli-1* revealed that 7% of the 15 morphant embryos assayed had low effects, and 7% had high effects. The probe *flk-1* indicated that 7% of the 14 morphants reviewed had high effects. The probe *tie-1* indicated that 7% of the 14 morphants observed had low effects, and 7% had high effects. The probe *cdh5* indicated that 8% of the 26 morphants observed had medium effects. The probe *flt-4* indicated that all 15 morphants observed were normal, and the probe *tie-2* indicated that all 15 morphants observed were normal.

The following table, Table 10, summarizes this data.

Probe	Number analyzed	Results
<i>fli-1</i>	15	7%L, 7%H
<i>flk-1</i>	14	7%H
<i>tie-1</i>	14	7%L, 7%H
<i>cdh5</i>	26	8%M
<i>flt-4</i>	15	normal
<i>tie-2</i>	15	normal

Table 10: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Figure 21 shows a representative morphant embryo at 28 hpf. As indicated by the arrow, yolk sac edema was observed in 56% of the 59 morphants studied. At 56 hpf a total of 20 embryos were observed for phenotypic characteristics, a representative morphant embryo is shown in Figure 22. As indicated by the arrow, pericardial edema was observed in 35% of embryos, 65% had a blood pool in the yolk, also indicated by the arrow, and 30% showed reduced IS blood flow.

Additional analyses were conducted on 48-56 hpf morphant embryos from the second dose group. A primary *in situ* hybridization screen with *cdh5* on 22 morphants showed all as normal. Microangiography on 30 second dose group morphants was used to locate the presence of FITC-Dextran in various regions of the embryo, see Figure 23. Of the 30 embryos, 13% had FITC-Dextran in the heart, and 3% in the heart and head. Reduced intersegmental vasculature was seen as indicated

by the arrow in Figure 23. High effects were observed in 7% of the embryos and low effects in 20%. No leaky vasculature was observed. Normal embryos accounted for 57% of the sample.

The data reveal that the gene is expressed in specific endothelium. In kidneys, it is expressed by certain vessels and some other epithelial structures. There is also some expression in the liver.

#### Gene HUP8010A10

The gene having the sequence shown in SEQ ID NO:28 was identified as selectively expressed during angiogenesis based on microarray data, see Figure 24. Specific data are given below in Table 11. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:29) and the human homolog (SEQ ID NO:31) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 30 and 32, respectively.

	intensity	log2 exp ratio	p-value	rank
cead	8,8	0,98		424
ecdo	8,7	0,36		1747
abdo	8,4	0,26	0,63	
anccedec	8,2	-2,20	0,00	
shedo	8,4	-0,06	0,38	
stedo	8,7	0,72	0,95	
enecl	9,4	0,14	0,67	
esedo	8,9	-0,37	0,03	

Table 11: Expression profile data for gene HUP8010A10

Based on this expression profile, the gene was further analyzed in zebrafish embryos. A corresponding zebrafish genes was identified for targeting. Two morpholinos were prepared, sz267 and sz268, each targeted to one of the zebrafish genes. In a first dosage group, 4ng of sz267 morpholino and 2ng of sz268 morpholino were administered to each fertilized egg. In a second dosage group, 6ng of sz267 morpholino and 3ng of sz268 morpholino were administered. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression was analyzed in the assay and results differed somewhat based on the probe used. In the first dose group (4ng sz267, 2ng sz268), the probe *fli-1* revealed that 13% of the 15 morphant embryos assayed had high effects. The probe *flk-1* indicated that 33% of the 15 morphants reviewed had low effects and 20% had high effects. The probe *tie-1* indicated that all 17 morphants observed were normal. The probe *cdh5* indicated that 8% of the 25 morphants observed had high effects. The probe *flt-4* indicated that all 13 morphants observed were normal, and the probe *tie-2* indicated that all 16 morphants observed were normal.

In the second dose group (6ng sz267, 3ng sz268), the probe *fli-1* revealed that 25% of the 16 morphant embryos assayed had low effects, and 19% had high effects. The probe *flk-1* indicated that 33% of the 6 morphants reviewed had low effects. The probe *tie-1* indicated that 67% of the 15 morphants observed had high effects. The probe *cdh5* indicated that 21% of the 24 morphants observed had low effects, 13% had medium effects, and 29% had high effects. The probe *flt-4* indicated that all 15 morphants observed were normal, and the probe *tie-2* indicated that all 15 morphants observed were normal.

The following Table 12 summarizes the foregoing data.

Probe	Morphants with 4 ng sz267, 2 ng sz268		Morphants with 6 ng sz267, 3 ng sz268	
	Number analyzed	Results	Number analyzed	Results
<i>fli-1</i>	15	13%H	16	25%L, 19%H
<i>flk-1</i>	15	33%L, 20%H	6	33%L
<i>tie-1</i>	17	normal	15	67%H
<i>cdh5</i>	25	8%H	24	21%L, 13%M, 29%H
<i>flt-4</i>	13	normal	15	normal
<i>tie-2</i>	16	normal	15	normal

Table 12: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Figure 25 shows a representative morphant embryo at 28 hpf. Cell death in the head was observed in 47% of the 61 embryos observed, as indicated by the arrow in Figure 25. The arrowhead indicates expanded hindbrain, which was seen in 51% of embryos. Mild yolk sac edema was observed in 21% of the morphants. At 56 hpf a total of 59 embryos were observed for phenotypic characteristics, a representative morphant embryo is shown in Figure 26. The arrow indicates expanded hindbrain, which was seen in 44% of embryos. Mild yolk sac edema was observed in 29% of the morphants and is indicated by the arrowhead. Reduced IS blood flow was noted in 14% of the embryos, and reduced blood flow was found in 17% of the embryos.

Additional analyses were conducted on 48-56 hpf morphant embryos from the second dose group. A primary *in situ* hybridization screen with *cdh5* on 19 morphants showed 68% as normal. Low effects of reduced intersegmental expression were seen in 32% of the embryos. Microangiography on morphants was used to locate the presence of FITC-Dextran in various regions of the embryo, see Figure 27. Of the 32 embryos, none had FITC-Dextran in the heart, or the heart and head. Reduced intersegmental vasculature was seen as indicated by the arrow in Figure 27. High effects were observed in 6% of the embryos, medium effects in 3% and low effects in 34%. No leaky vasculature was observed. Normal embryos accounted for 56% of the sample.

#### Gene NOC8003L17

The gene having the sequence shown in SEQ ID NO:33 was identified as selectively expressed in blood vessels based on microarray data, see Figure 28. Specific data are given below in Table 13. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:34) and the human homolog (SEQ ID NO:36) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 35 and 37, respectively.

	Intensity	log2 expression	p value	rank
<i>eecl1</i>	7,7	-0,06		6087
<i>eecl6</i>	7,6	0,40		1547
<i>eecl9</i>	7,5	1,47	0,97	
<i>eecl10</i>	8,3	0,30	0,77	
<i>eecl11</i>	7,2	-0,04	0,41	
<i>eecl12</i>	8,3	1,87	1,00	
<i>eecl13</i>	7,8	0,60	0,99	
<i>eecl14</i>	7,5	-0,13	0,24	

Table 13: Expression profile data for gene NOC8003L17

Based on this expression profile, the gene was further analyzed in zebrafish embryos. One corresponding zebrafish gene was identified for targeting. Two morpholinos were prepared, sz180 and sz181, each targeted to the zebrafish

gene. In a first dose group, 12ng of sz180 and 1ng of sz181 were administered to each fertilized egg. In a second dose group, 12ng of sz180 and 2ng of sz181 were administered to each fertilized egg. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression in embryos from the first dose group was analyzed in the assay and results differed somewhat based on the probe used. The probe *fli-1* revealed that 40 of the 15 morphant embryos assayed had high effects. The probe *flk-1* indicated that all 7 of the morphants reviewed were normal. The probe *cdh5* indicated that 15% of the 20 morphants observed had low effects, as well as 5% with medium effects and 30% with high effects. The probe *flt-4* indicated that all 11 morphants observed were normal, and the probe *tie-2* indicated that all 16 morphants observed were normal.

The following table, Table 14, summarizes this data.

Probe	Number analyzed	Results
<i>fli-1</i>	15	40%H
<i>flk-1</i>	7	normal
<i>tie-1</i>	15	normal
<i>cdh5</i>	20	15%L, 5%M, 30%H
<i>flt-4</i>	11	normal
<i>tie-2</i>	16	normal

Table 14: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Figure 29 shows a representative morphant embryo. As indicated by the arrow, yolk sac edema was observed in 67% of the 48 morphants studied. Embryos observed at 56 hpf demonstrated normal morphology, a representative embryo is shown in Figure 30.

Additional analyses were conducted on 48-56 hpf morphant embryos. A primary *in situ* hybridization screen with *cdh5* on 14 morphants from the second dose group showed 79% as normal, the remaining 29% showing medium effects of reduced intersegmental expression. Microangiography on 29 first dose group morphants was used to locate the presence of FITC-Dextran in various regions of the embryo, see Figure 31. Of the 29 embryos, none had FITC-Dextran in the heart, 3% had FITC-Dextran in the heart and head. Reduced intersegmental vasculature was seen as indicated by the arrow in Figure 31. High effects were observed in 7% of the embryos, medium effects in 3% and low effects in 38%. No leaky vasculature was observed. Normal embryos accounted for 45% of the sample.

GeneNOC8009C9

The gene having the sequence shown in SEQ ID NO:38 was identified as selectively expressed during angiogenesis based on microarray data, see Figure 32. Specific data are given below in Table 15. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:39) and the human homolog (SEQ ID NO:41) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 40 and 42, respectively.

	Intensity	log2 expression	p-value	rank
eechr	8,1	0,84		685
ecto	8,0	1,19		184
apecio	7,4	1,57	1,00	
apecebec	8,1	1,44	1,00	
anecid	8,4	0,12	0,74	
efecio	8,0	1,52	1,00	
ehedlo	8,3	0,53	0,98	
esecio	7,9	2,41	1,00	

Table 15: Expression profile data for gene NOC8009C9

Based on this expression profile, the gene was further analyzed in zebrafish embryos. One corresponding zebrafish genes was identified for targeting. Two morpholinos were prepared, sz241 and sz242, targeted to the zebrafish gene. Three (3)ng of sz241 morpholino and 1ng of sz242 morpholino were administered to each fertilized egg. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression was analyzed in the assay and results differed somewhat based on the probe used. The probe *fli-1* revealed that 7% of the 15 morphant embryos assayed had medium effects. The probe *flk-1* indicated that all 15 morphants observed were normal. The probe *tie-1* indicated that 7% of the 15 morphants observed had high effects. The probe *cdh5* indicated that 15% of the 20 morphants observed had low effects. Medium effects were seen in 5% and high effects in 15%. The probe *flt-4* indicated that all 15 morphants observed were normal, and the probe *tie-2* indicated that all 16 morphants observed were normal.

The following table, Table 16, summarizes this data.

Probe	Number analyzed	Results
<i>fli-1</i>	15	7%M
<i>flk-1</i>	15	normal
<i>tie-1</i>	15	7%H
<i>cdh5</i>	20	15%L, 5%M, 15%H
<i>flt-4</i>	15	normal
<i>tie-2</i>	16	normal

Table 16: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Figure 33 shows a representative morphant embryo at 28 hpf. Mild cell death was observed in 25% of the 53 embryos observed. At 56 hpf 52 embryos were observed for phenotypic characteristics, a representative morphant embryo is shown in Figure 34. As indicated by the arrow, pericardial edema was seen in 13% of embryos. The arrowhead points toward a region of yolk sac edema, seen in 25% of embryos.

Reduced IS blood flow was observed in 13% and 15% showed reduced axial blood flow.

Additional analyses were conducted on 48-56 hpf morphant embryos. A primary *in situ* hybridization screen with *cdh5* on 31 morphants showed all as normal. Microangiography on morphants was used to locate the presence of FITC-Dextran in various regions of the embryo, see Figure 35. Of the 29 embryos, 7% had FITC-Dextran in the heart, and 14% in the heart and head. Reduced intersegmental vasculature was seen as indicated by the arrows in Figure 35. High effects were observed in 3% of the embryos, medium effects in 3% and low effects in 21%. No leaky vasculature was observed. Normal embryos accounted for 52% of the sample. The data reveal that the gene is expressed in and around the heart and around organs, including some expression in select organs.

#### Gene NOC8009G23

The gene having the sequence shown in SEQ ID NO:43 was identified as selectively expressed during angiogenesis based on microarray data, see Figure 36. Specific data are given below in Table 17. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:44) and the human homolog (SEQ ID NO:46) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 45 and 47, respectively.

	intensity	log2 exp ratio	p-value	rank
<i>eecl1</i>	8,3	0,23		3262
<i>eecl0</i>	8,2	1,19		185
<i>apecl0</i>	7,2	2,94	1,00	
<i>apecebe</i>	8,1	2,09	1,00	
<i>aneclo</i>	8,0	1,15	1,00	
<i>eebcl0</i>	8,2	0,75	0,94	
<i>enecl0</i>	9,0	-0,02	0,47	
<i>eecl0</i>	8,1	2,51	1,00	

Table 17: Expression profile data for gene NOC8009G23

Based on this expression profile, the gene was further analyzed in zebrafish embryos. One corresponding zebrafish gene was identified for targeting. Two morpholinos were prepared, sz149 and sz150, each targeted to the zebrafish

gene. In a first dose group, 1.5ng of sz149 morpholino and 1.5ng of sz150 morpholino were administered to fertilized eggs. In a second dose group, 2ng of sz149 morpholino and 2ng of sz150 morpholino were administered to fertilized eggs. In a third dose group, 3ng of sz149 morpholino and 3ng of sz150 morpholino were administered to fertilized eggs. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression was analyzed in the assay and results differed somewhat based on the probe used. In studies with embryos from the second dose group, the probe *fli-1* revealed that 21% of the 14 morphant embryos assayed had high effects. The probe *flik-1* indicated that all 16 morphants reviewed were normal. The probe *tie-1* indicated that 36% of the 22 morphants observed had high effects. The probe *cdh5* indicated that 13% of the 15 morphants observed had low effects with breaks in axial expression. The probe *flt-4* indicated that all 16 morphants observed were normal, and the probe *tie-2* indicated that all 13 morphants observed were normal.

In studies from the second dose group, the probe *fli-1* revealed that 13% of the 15 morphant embryos assayed had medium effects, 27% had high effects. The probe

*flk-1* indicated that 7% of the 15 morphants reviewed had low effects, 7% had medium effects and 20% had high effects. The probe *tie-1* indicated that 62% of the 13 morphants observed had high effects. The probe *cdh5* indicated that 25% of the 12 morphants observed had high effects, some with breaks in axial expression. The probe *flt-4* indicated that 13% of the 15 morphants observed had low effects in the axial vessels, including breaks in axial expression and severely malformed tails. The probe *tie-2* indicated that all 8 morphants observed were normal.

The following Table 18 summarizes the foregoing data.

Probe	Morphants with 2 ng sz149, 2 ng sz150		Morphants with 3 ng sz149, 3 ng sz150	
	Number analyzed	Results	Number analyzed	Results
<i>fli-1</i>	14	21%H	15	13%M, 27%H
<i>flk-1</i>	16	normal	15	7%L, 7%M, 20%H
<i>tie-1</i>	22	36%H	13	62%H
<i>cdh5</i>	15	13%L***	12	25%H***
<i>flt-4</i>	16	normal	15	13% low axial effects
<i>tie-2</i>	13	normal	8	normal

Table 18: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Figure 37 shows a representative morphant embryo at 28 hpf. As indicated by the arrow, cell death was observed in 40% of the 20 embryos observed. As indicated by the arrowhead, yolk sac edema was observed in 55% of the morphants. Curly down body was seen in 40% of morphants. At 56 hpf a total of 20 embryos were observed for phenotypic characteristics, a representative morphant embryo is shown in Figure 38. As shown by the arrow, pericardial edema was observed in 55% of morphants. The arrowhead points toward yolk sac edema, observed in 55% of morphants. Curly down body was reported in 30% of embryos.

Additional analyses were conducted on 48-56 hpf morphant embryos. A primary *in situ* hybridization screen with *cdh5* on 22 morphants from the first dose group showed 90% as normal. Low effects of reduced intersegmental expression and curly down embryos were seen in 5% of the embryos, and high effects with very short tails were seen in 5% of embryos. A primary *in situ* hybridization screen with *cdh5* was also performed on 20 morphants from the third dose group, showing 80% as normal. Low effects of reduced intersegmental expression were seen in 5% of the embryos, and medium effects in 15% of embryos.

Microangiography on morphants was used to locate the presence of FITC-Dextran in various regions of the embryo, see Figure 39. Of the 25 embryos, none had FITC-Dextran in the heart, or the heart and head. Reduced intersegmental vasculature was seen in 24% of the embryos as indicated by the arrow in Figure 39. No leaky vasculature was observed. Normal embryos accounted for 76% of the sample, an example of normal intersegmental vessels is indicated by the arrowhead.

Gene QJC8003C9

The gene having the sequence shown in SEQ ID NO:48 was identified as selectively expressed in blood vessels based on microarray data, see Figure 40. Specific data are given below in Table 19. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:49) and the human homolog (SEQ ID NO:51) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 50 and 52, respectively.

	intensity	log2 exp ratio	p-value	rank
eccl	8,1	0,67		1246
eclo	8,0	1,37		123
abeclo	7,1	2,90	1,00	
abeceloc	8,1	0,59	0,92	
aneclo	7,6	0,19	0,85	
ebeclo	8,7	1,41	1,00	
ebeclo	8,4	1,89	1,00	
eseclo	7,7	1,28	1,00	

Table 19: Expression profile data for gene OJC8003C9

Based on this expression profile, the gene was further analyzed in zebrafish embryos. One corresponding zebrafish gene was identified for targeting. Two morpholinos were prepared, sz129

and sz130, each targeted to the zebrafish gene. In a first dose group, 3ng of sz129 morpholino and 4.5ng of sz130 morpholino were administered to each fertilized egg. In a second dose group, 4ng of sz129 morpholino and 6ng of sz130 morpholino were administered to each fertilized egg. In a third dose group, 6ng of sz129 morpholino and 8ng of sz130 morpholino were administered to each fertilized egg. In a fourth dose group, 6ng of sz129 morpholino and 9ng of sz130 morpholino were administered to each fertilized egg. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression was analyzed in the assay and results differed somewhat based on the probe used. In embryos from the second dose group, the probe *fli-1* revealed that 20% of the 10 morphant embryos assayed had low effects, 10% had medium effects and 10% had high effects. The probe *flk-1* indicated that 11% of the 9 morphants reviewed had low effects, 33% had medium effects and 11% had high effects. The probe *tie-1* indicated that 22% of the 9 morphants observed had high effects. The probe *cdh5* indicated that 14% of the 7 morphants observed had medium effects and 14% had high effects. The probe *flt-4* indicated that all 9 morphants observed were normal, and the probe *tie-2* indicated that all 7 morphants observed were normal.

In embryos from the third dose group, the probe *fli-1* revealed that 10% of the 10 morphant embryos assayed had medium effects, and 50% had high effects. The probe *flk-1* indicated that 25% of the 12 morphants reviewed had medium effects and 25% had high effects. The probe *tie-1* indicated that 17% of the 6 morphants observed had low effects, and 50% had high effects. The probe *cdh5* indicated that 40% of 5 morphants observed had medium effects. The probe *flt-4* indicated that all 9 morphants observed were normal, and the probe *tie-2* indicated that all 6morphants observed were normal.

The following Table 20 summarizes the foregoing data.

Probe	Morphants with 3 ng sz185, 6 ng sz186		Morphants with 6 ng sz185, 12 ng sz186	
	Number	Results	Number	Results

	analyzed		analyzed	
<i>fli-1</i>	10	20%L, 10%M, 10%H	10	10%M, 50%H
<i>flk-1</i>	9	11%L, 33%M, 11%H	12	25%M, 25%H
<i>tie-1</i>	9	22%H	6	17%L, 50%H
<i>cdh5</i>	7	14%M, 14%H	5	40%M
<i>fli-4</i>	9	normal	9	normal
<i>tie-2</i>	7	normal	6	normal

Table 20: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Figure 41 shows a representative morphant embryo at 28 hpf. As indicated by the arrow, curly down body was found in 35% of the 20 morphants observed. At 56 hpf a total of 20 embryos were observed for phenotypic characteristics, a representative morphant embryo is shown in Figure 42. As indicated by the long arrow, 60% of the embryos had cell death with an associated expanded hindbrain ventricle. Yolk sac edema was observed in 25% of embryos, as indicated by the short arrow. The arrowhead points out the lack of pericardial edema associated with the yolk sac edema.

Additional analyses were conducted on 48-56 hpf morphant embryos. A primary *in situ* hybridization screen with *cdh5* on 19 morphants from the first dose group showed all were normal. The primary *in situ* hybridization screen with *cdh5* on 10 morphants from the fourth dose group showed only 90% normal, the remaining 10% exhibiting low effects with curly tails. Microangiography on 19 morphants from the third dose group was used to locate the presence of FITC-Dextran in various regions of the embryo, see Figure 43. Of the 19 embryos, none had FITC-Dextran in the heart and head but 16% had it in the heart alone. Reduced intersegmental vasculature was seen in 37% of the embryos. No leaky vasculature was observed. Normal embryos accounted for 47% of the sample.

#### Gene OJC8009J7

The gene having the sequence shown in SEQ ID NO:53 was identified as selectively expressed in blood vessels based on microarray data, see Figure 44. Specific data are given below in Table 21. Using sequence and annotation databases the equivalent gene in mice (SEQ ID NO:54) and the human homolog (SEQ ID NO:56) was also deduced. Proteins encoded by these sequences are given at SEQ ID NO:s 55 and 57, respectively.

	intensity	log2 exo ratio	p-value	rank
<i>ecdr</i>	8.5	0.62		1445
<i>ecdo</i>	8.3	1.03		257
<i>abecdo</i>	8.3	2.37	1.00	
<i>abceced</i>	9.2	0.38	0.84	
<i>anecdo</i>	7.9	0.93	1.00	
<i>abecdo</i>	8.8	1.53	1.00	
<i>enecdo</i>	8.8	1.03	1.00	
<i>esecdo</i>	8.5	0.48	0.99	

Table 21: Expression profile data for gene OJC8009J7

Based on this expression profile, the gene was further analyzed in zebrafish embryos. One corresponding zebrafish gene was identified for targeting. Two morpholinos were prepared, sz175 and sz176, each targeted to the zebrafish gene. Two (2) ng of sz175 morpholino and 12ng of sz176 morpholino were

administered to each fertilized egg. The embryos were allowed to develop. At 24 hpf a secondary *in situ* hybridization screen with six different probes was conducted.

Intersegmental expression was analyzed in the assay and results differed somewhat based on the probe used. The probe *fli-1* revealed that 7% of the 14 morphant embryos assayed had low effects, and 29% had high effects. The probe *flk-1* indicated that 25% of the 12 morphants reviewed had low effects and 8% had high effects. The probe *tie-1* indicated that 11% of the 9 morphants observed had medium effects and 11% had high effects. The probe *cdh5* indicated that 19% of the 16 morphants observed had low effects, 6% had medium effects, and 19% had high effects. The probe *flt-4* indicated that all 10 morphants observed were normal, and the probe *tie-2* indicated that all 6 morphants observed were normal.

The following table, Table 22, summarizes this data.

Probe	Number analyzed	Results
<i>fli-1</i>	14	7%L, 29%M
<i>flk-1</i>	12	25%L, 8%H
<i>tie-1</i>	9	11%M, 11%H
<i>cdh5</i>	16	19%L, 6%M, 19%H
<i>flt-4</i>	10	normal
<i>tie-2</i>	6	normal

Table 22: secondary *in situ* hybridization data

At 28 hpf embryos were observed morphologically. Figure 45 shows a representative morphant embryo at 28 hpf. As evidenced from the figure, particularly when viewed in light of the 28 hpf wild type embryo of Figure 4, the morphants exhibited normal morphology. At 56 hpf embryos again were observed for phenotypic characteristics, a representative morphant embryo is shown in Figure 46. Again, the normal morphology observed in the embryos can be easily understood when Figure 46 is viewed in light of the 56 hpf wild type embryo of Figure 6.

Additional analyses were conducted on 48-56 hpf morphant embryos. A primary *in situ* hybridization screen with *cdh5* on 17 morphants showed 88% as normal. Medium effects of reduced intersegmental expression were seen in the other 12% of embryos. Microangiography on morphants was used to locate the presence of FITC-Dextran in various regions of the embryo, see Figure 47. Of the 33 embryos, none had FITC-Dextran in the heart and head combined, but 4% had it in the heart alone. Reduced intersegmental vasculature was seen in 15% of embryos. No leaky vasculature was observed. Normal embryos accounted for 81% of the sample. The leaks observed came from blood vessels in the posterior head as indicated by the arrowhead.

#### Novel Applications Ascertained from *in vivo* Data

The present invention relates to the eleven gene targets, and proteins related thereto, which were originally identified as upregulated during vasculogenesis or angiogenesis through microarray evaluation and subsequently proven to play a critical role *in vivo* with zebrafish embryo experimentation. These genes and proteins can form the basis of novel methods and treatments directed to

angiogenesis-related conditions. For example, biological samples from a patient suspected of suffering from an angiogenesis-related condition can be screened to ascertain if genes or proteins of the present invention are expressed at the correct time, location, and intensity in the patient. Such screening methods form part of the claimed invention. If a gene and/or protein is identified as improperly expressed, therapies to correct the condition such as gene therapy or medicament can be initiated according to methods and procedures described herein or known in the art. With such specific data as is now possible using tools described herein, rapid diagnosis and specific, targeted treatment is possible.

One type of screening method envisioned relies on gene amplification for detecting patients with conditions related to vasculogenesis or angiogenesis. Such methods could employ PCR, *in situ* hybridisation, and/or Southern blotting techniques to elucidate the condition. Another type of screening method could be based on evaluations of gene expression, using known techniques such as quantitative PCR, microarrays, Northern blotting, or *in situ* hybridisation. Yet another type of screening method that could be used would measure or monitor protein expression and could be effected with techniques such as immunohistochemistry, Western blotting, ELISA, or FACS.

If it is determined, through methods of the present invention or other methods, that an angiogenesis-related condition could be improved through administration of compounds containing genes and/or proteins according to the invention, one or more of the genes and/or proteins could be administered together or sequentially by methods known in the art.

Isolated nucleic acid molecules or proteins of the present invention can be obtained, for example, by synthesis using standard direct peptide synthesizing techniques or recombinant methods. Proteins may be isolated or purified in a variety of ways known to those skilled in the art, such as electrophoretic purification or chromatographic techniques.

Administration of the compounds of the present invention can be effected by any method that enables delivery of the compounds to the site of desired action. These methods include oral routes, intraduodenal routes, parenteral injection (including intravenous, subcutaneous, intramuscular, intravascular or infusion), and topical administration.

Gene therapy approaches may be used to introduce nucleotides of the present invention into a cell, group of cells, or organism. Both *in vivo* and *ex vivo* methods can be utilized. Vectors typically are used in this procedure. Non-virus or virus vectors could be employed, for example recombinant adenovirus or retrovirus. According to this use, the desired gene is introduced into a DNA virus or RNA virus, such as avirulent retrovirus, adenovirus, adeno-associated virus, herpes virus, vaccinia virus, poxvirus, poliovirus, Sindbis virus, Sendai virus, SV40, and immunodeficiency virus (HIV). The recombinant virus is then infected into the target cell(s). Multiple genes could be incorporated in a single vector, alternatively, they could be introduced to the target cell(s) in separate vectors simultaneously or sequentially. These methods are known in the art and are described in numerous patents and publications.

Another means to interfere with gene expression or protein production contemplated by the present invention is to employ small interfering RNA (siRNA). siRNA comprises a sense and antisense strand of RNA corresponding to the gene of interest, for example, SEQ ID NO:2. A siRNA molecule consists of approximately 19 nucleotides plus an overhang of approximately 2 nucleotides at the 3' end. Some preferred methods include between 19-23 nucleotides plus 3' overhang. The siRNA is introduced to the cell or cells of interest through known methods. Following introduction, the cell or cells destroy ssRNA having the same sequence. This results in a reduction or prevention in translation of a targeted gene and a corresponding reduction or prevention in protein production.

The amount of active compound administered can be determined after assessing the subject being treated, the severity of the disorder or condition, the rate of administration, and the disposition of the compound. Doses may be administered all at once, or spread out over a discrete time period.

Compounds of the present invention may be applied as a sole therapy or may involve one or more other active medicinal or pharmaceutical agent. Compositions may include carriers, adjuvants, buffers, or excipients as known in the art. If desired, the compositions may further contain ingredients such as flavorings, sweeteners, binders, dyes, lubricating agents, perfume, thickening agents, stabilizers, emulsifiers, dispersants, suspending agents, preservatives, and pH regulating agents. Compositions may be in any suitable form, for example, tablet, capsule, pill, powder, sustained release formulation, solution, suspension, emulsion, ointment or cream. The compositions may be sterile. Methods of preparing various pharmaceutical compositions with a specific amount of active compound are known or apparent to those skilled in this art. The pharmaceutical compositions of the present invention that have been described can be applied to all diseases that require vasculogenic or angiogenic therapy.

For example, one method for the treatment of an angiogenesis-related disorder involves a composition according to the present invention used to vascularize ischemic tissue. There are many ways to determine if a tissue is at risk of suffering ischemic damage from undesirable vascular occlusion. Such methods are well known in the art and include, for example, imaging techniques such as MRI to evaluate myocardial disease. After determining where and when to apply compositions of the present invention, the compositions can be administered to increase angiogenesis in tissue affected by or at risk of being affected by a vascular occlusion. This could be an effective means of preventing and/or attenuating ischemia in such tissue. Methods are known in the art to evaluate and measure the degree to which ischemia has been attenuated.

Further treatment methods according to the present invention include the use of any known technique that permits visualization, measurement, and/or evaluation of the functionality and degree of ischemia of the patient's heart. Such evaluations could be made prior to initiating treatment, during the course of treatment, after treatment has been completed, or at some or all stages. Examples of such techniques include echocardiography, cardiovascular nuclear imaging, magnetic resonance imaging, and contrast angiography.

Although the present invention takes a step forward in the understanding of vasculogenesis and angiogenesis, and treatments for conditions related to the same, there is still a need in the art to further understand these conditions. Therefore, the present invention further contemplates the creation and use of non-human transgenic animals which could be used for analysis and experimentation. Transgenic animals containing mutant, knock-out or modified genes corresponding to those disclosed herein are therefore also included in the invention. Transgenic animals are genetically modified animals into which recombinant, exogenous or cloned genetic material has been experimentally transferred. Such genetic material is often referred to as a transgene. The nucleic acid sequence of the transgene may be integrated either at a locus of a genome where that particular nucleic acid sequence is not otherwise normally found or at the normal locus for the transgene. The transgene may consist of nucleic acid sequences derived from the genome of the same species or of a different species than the species of the target animal.

Transgenic animals can be produced by a variety of different methods including transfection, electroporation, microinjection, gene targeting in embryonic stem cells and recombinant viral and retroviral infection as known in the art. The method of introduction of nucleic acid fragments into recombination competent mammalian cells can be by any method that favors co-transformation of multiple nucleic acid molecules. Detailed procedures for producing transgenic animals are available to one skilled in the art, for example, U.S. Pat. Nos. 5,489,743 and 5,602,307.

Transgenic technology can be used to produce animals which lack one or more of the eleven genes described above. Such knockout animals can be used, especially when their growth and development is measured against data from a wild type or control animal, to elucidate timing and function of the deleted gene(s). Further, these animals could also be engineered to exhibit angiogenesis-related disease states, thus furthering the understanding of the role of the particular gene(s) in the progression of the selected disease. This knowledge would be an advance in the state of the art and could lead to promising new therapies for the prevention, management, and cure of disease.

Further uses of transgenic animals according to the present invention include replacement of one or more of the above-identified gene(s) in the research organism with the human homolog of the gene. For example, a transgenic mouse whose gene corresponding to SEQ ID NO:7 has been replaced with the human homolog, SEQ ID NO:9. While it is accepted that research into effective drug therapies can be conducted in animal models, such a transgenic mouse could be a more effective screening tool into potential drug candidates for human use.

**We Claim:**

1. An isolated nucleic acid molecule according to any one of SEQ ID NO:s 2, 4, 7, 9, 12, 14, 17, 19, 21, 24, 26, 29, 31, 34, 36, 39, 41, 44, 46, 49, 51, 54, and 56 or a fragment or analogue thereof which has the ability to stimulate or inhibit at least one biological activity selected from the group consisting of vasculogenesis, angiogenesis, vascular permeability, endothelial cell proliferation, endothelial cell differentiation, endothelial cell migration, and endothelial cell survival, or an isolated nucleic acid molecule which hybridizes to one of the foregoing sequences under stringent conditions.
2. An isolated nucleic acid molecule which hybridizes to the compliment of a nucleic acid molecule according to Claim 1 under stringent conditions.
3. An isolated siRNA molecule targeted to an isolated nucleic acid molecule according to Claims 1 or 2, wherein the isolated siRNA molecule is at least 19 base pairs long.
4. An expression vector comprising the nucleic acid according to Claims 1 or 2, optionally the nucleic acid may be operatively associated with a regulatory nucleic acid controlling the expression of the polypeptide encoded by said nucleic acid.
5. A host cell genetically engineered to contain a nucleic acid according to Claims 1 or 2.
6. A host cell transfected with an expression vector according to Claim 4.
7. A method of treating an angiogenesis-related condition in a cell, group of cells, or organism, comprising the step of administering an expression vector according to Claim 4 to the cell, group of cells, or organism.
8. An antibody with specific reactivity to a nucleic acid according to Claims 1 or 2, wherein the antibody may preferably be polyclonal or monoclonal and wherein the antibody may further comprise a detectable label such as a fluorescent label.
9. A transgenic, non-human animal which has been genetically engineered to contain a transgene comprising a nucleic acid according to Claims 1 or 2, preferably, the transgene may be expressed.
10. A pharmaceutical composition comprising a nucleic acid sequence according to Claims 1 or 2.
11. A method of affecting vasculogenesis or angiogenesis in a cell, group of cells, or organism, comprising the step of administering a pharmaceutical composition according to Claim 16 to the cell, group of cells, or organism, the affecting may preferably cause an increase or decrease, more preferably, the cell, group of cells, or organism has an angiogenesis-related disorder such as cancer, retinopathy, macular degeneration, corneal ulceration, stroke, ischemic heart disease, infertility, ulcers, scleradoma, wound healing, ischemia, ischemic heart disease, myocardial infarction, myocardiosis, angina pectoris, unstable angina, coronary arteriosclerosis,

arteriosclerosis obliterans, Berger's disease, arterial embolism, arterial thrombosis, cerebrovascular occlusion, cerebral infarction, cerebral thrombosis, cerebral embolism, rubeosis proliferative vitreoretinopathy, chronic inflammation, inflammatory bowel disease, psoriasis, sarcoidosis and rheumatoid arthritis.

12. An isolated polypeptide comprising a sequence of amino acids substantially corresponding to the amino acid sequence in any one of SEQ ID NO:s 3, 5, 8, 10, 13, 15, 18, 20, 22, 25, 27, 30, 32, 35, 37, 40, 42, 45, 47, 50, 52, 55, and 57 or a fragment or analogue thereof, said polypeptide having the ability to affect angiogenesis in a cell, a group of cells, or an organism.

13. A host cell genetically engineered to express a polypeptide according to Claim 12.

14. An antibody specifically reactive with a polypeptide according to Claim 12, wherein the antibody may preferably be polyclonal or monoclonal and wherein the antibody may further comprise a detectable label such as a fluorescent label.

15. A transgenic, non-human animal which has been genetically engineered to contain a transgene comprising a nucleic acid which encodes a polypeptide according to Claim 12, preferably, the transgene may be expressed.

16. A pharmaceutical composition comprising an isolated polypeptide according to Claim 12.

17. A method of affecting vasculogenesis or angiogenesis in a cell, group of cells, or organism, comprising the step of administering a pharmaceutical composition according to Claim 16 to the cell, group of cells, or organism, the affecting may preferably cause an increase or decrease, more preferably, the cell, group of cells, or organism has an angiogenesis-related disorder such as cancer, retinopathy, macular degeneration, corneal ulceration, stroke, ischemic heart disease, infertility, ulcers, scleradoma, wound healing, ischemia, ischemic heart disease, myocardial infarction, myocardosis, angina pectoris, unstable angina, coronary arteriosclerosis, arteriosclerosis obliterans, Berger's disease, arterial embolism, arterial thrombosis, cerebrovascular occlusion, cerebral infarction, cerebral thrombosis, cerebral embolism, rubeosis proliferative vitreoretinopathy, chronic inflammation, inflammatory bowel disease, psoriasis, sarcoidosis and rheumatoid arthritis.

18. A method of detecting an angiogenesis-related transcript in a cell in a patient, the method comprising contacting a biological sample from the patient with a polynucleotide that selectively hybridizes to a sequence at least 80% identical to a sequence according to any one of SEQ ID NO:s 2, 4, 7, 9, 12, 14, 17, 19, 21, 24, 26, 29, 31, 34, 36, 39, 41, 44, 46, 49, 51, 54, and 56, wherein an angiogenesis-related transcript is detected where hybridization is detected, preferably the polynucleotide comprises a sequence according to any one of SEQ ID NO:s 2, 4, 7, 9, 12, 14, 17, 19, 21, 24, 26, 29, 31, 34, 36, 39, 41, 44, 46, 49, 51, 54, and 56, preferably the biological sample is a tissue sample or is comprised of isolated nucleic acids such as mRNA, preferably the nucleic acids are amplified prior to the step of contacting the biological sample with the polynucleotide, preferably the polynucleotide is immobilized on a solid surface.

19. A method of affecting at least one bioactivity selected from angiogenesis and vasculogenesis in a vertebrate organism, said method comprising the step of administering to said organism an effective angiogenesis or vasculogenesis affecting amount of a nucleotide or polypeptide according to Claims 1 or 12, wherein the organism is preferably a mammal such as mice, rats, rabbits, guinea pigs, cats, dogs, pigs, cows, monkeys, and humans, wherein vasculogenesis or angiogenesis is preferably enhanced, increased, inhibited, or decreased, wherein the organism preferably has an angiogenesis-related disorder such as cancer, retinopathy, macular degeneration, corneal ulceration, stroke, ischemic heart disease, infertility, ulcers, scleradoma, wound healing, ischemia, ischemic heart disease, myocardial infarction, myocardosis, angina pectoris, unstable angina, coronary arteriosclerosis, arteriosclerosis obliterans, Berger's disease, arterial embolism, arterial thrombosis, cerebrovascular occlusion, cerebral infarction, cerebral thrombosis, cerebral embolism, rubeosis proliferative vitreoretinopathy, chronic inflammation, inflammatory bowel disease, psoriasis, sarcoidosis and rheumatoid arthritis.

20. A transgenic increased or decreased angiogenesis laboratory animal comprising one or more cells in which the expression of a sequence according to any one of SEQ ID NO:s 2, 4, 7, 9, 12, 14, 17, 19, 21, 24, 26, 29, 31, 34, 36, 39, 41, 44, 46, 49, 51, 54, and 56 is upregulated, downregulated, or absent.

**Abstract**

The present invention relates to polynucleotides and proteins associated with vasculogenesis- and angiogenesis-related disorders. The invention further relates to methods for the identification of compounds that modulate the expression of angiogenesis-related genes and gene products and to using such compounds as therapeutic agents in the treatment of angiogenesis-related disorders. The invention also relates to methods for the diagnostic evaluation, genetic testing and prognosis of angiogenesis-related disorders, and to methods and compositions for the treatment of these disorders.

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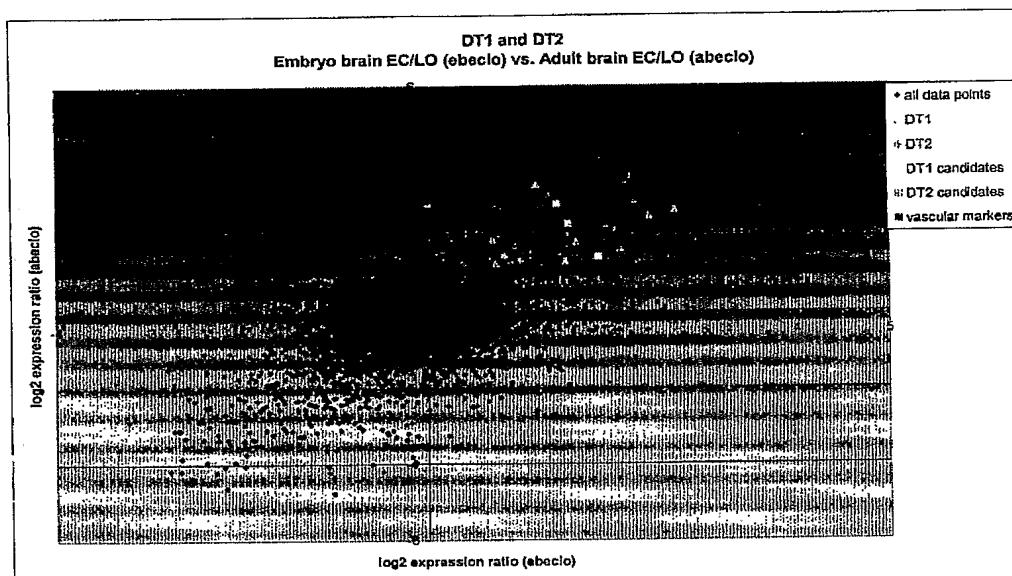


Figure 1

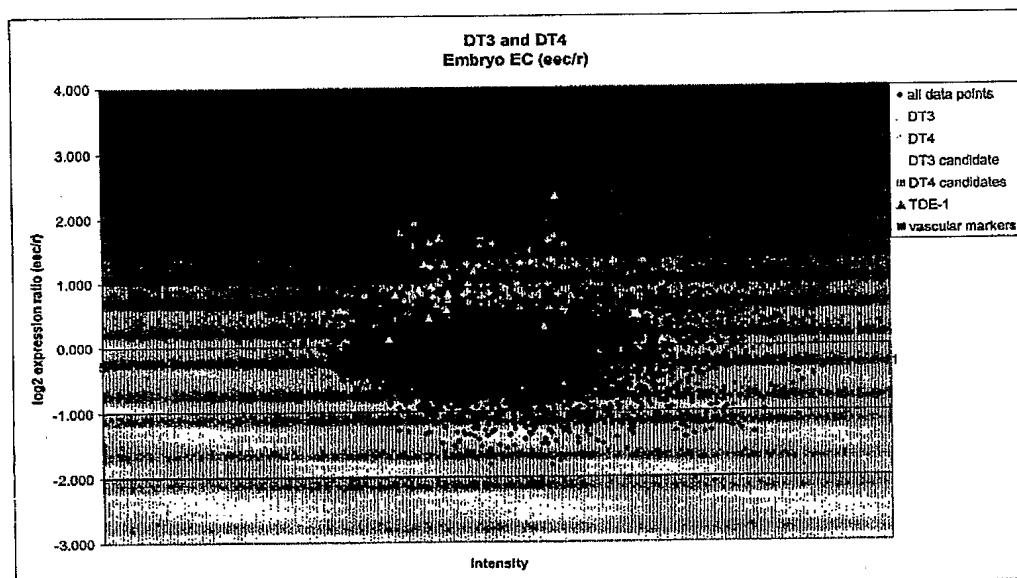


Figure 2

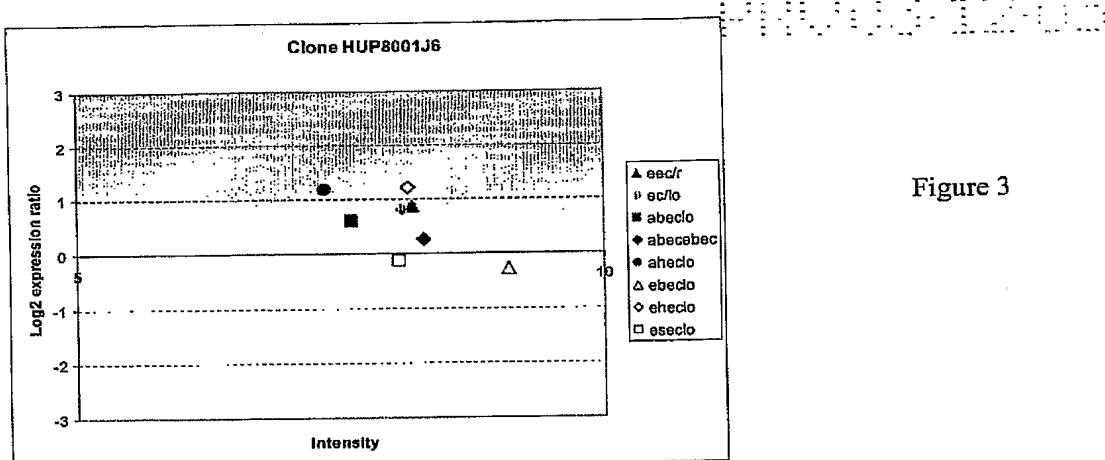


Figure 3

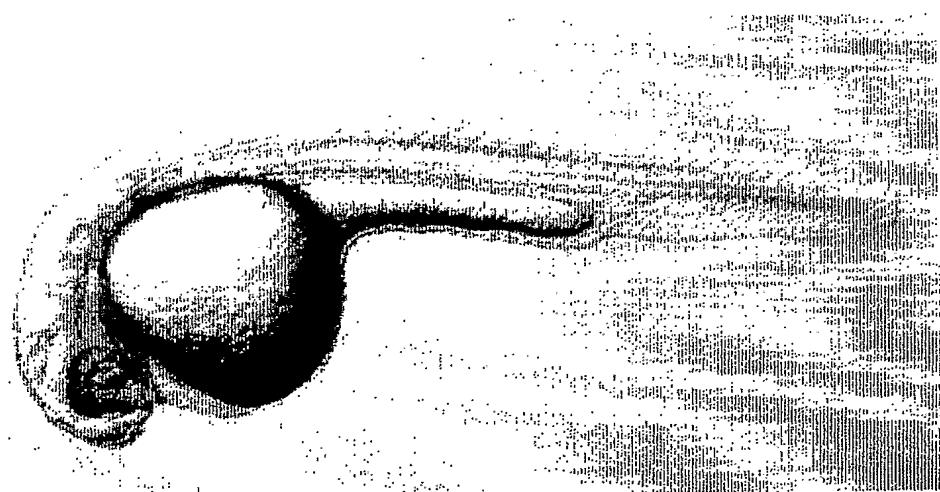


Figure 4

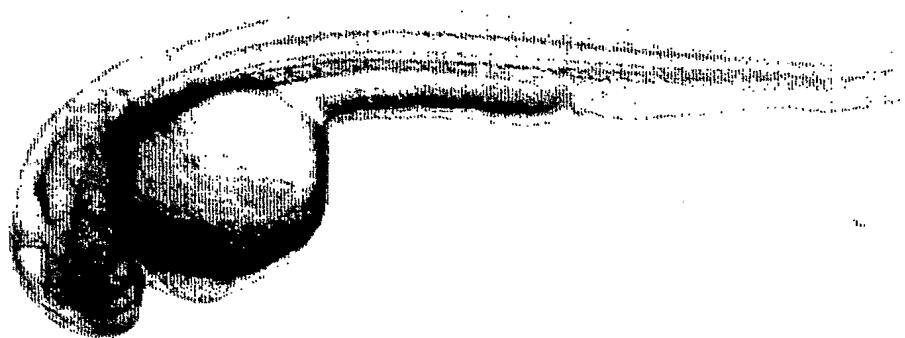


Figure 5

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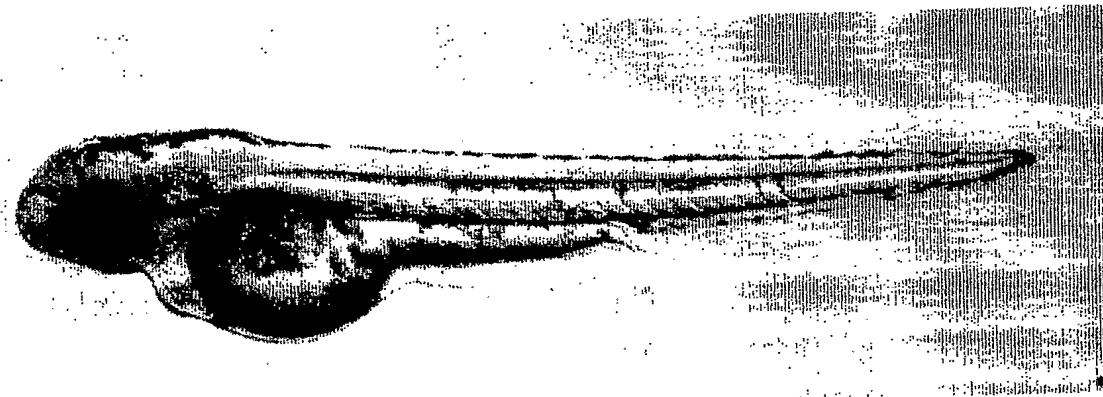


Figure 6

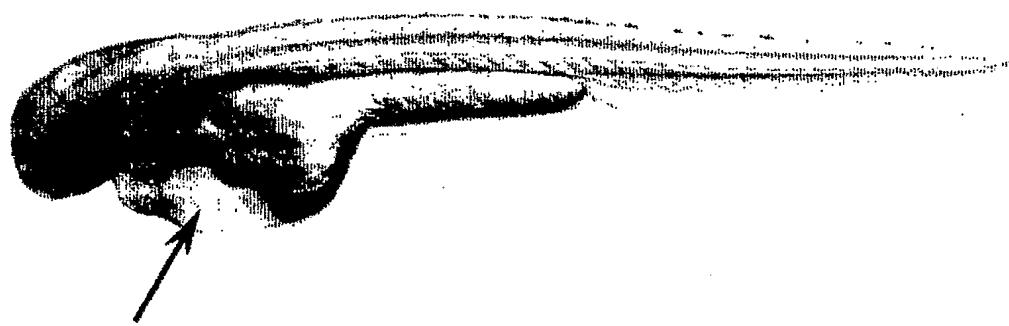


Figure 7

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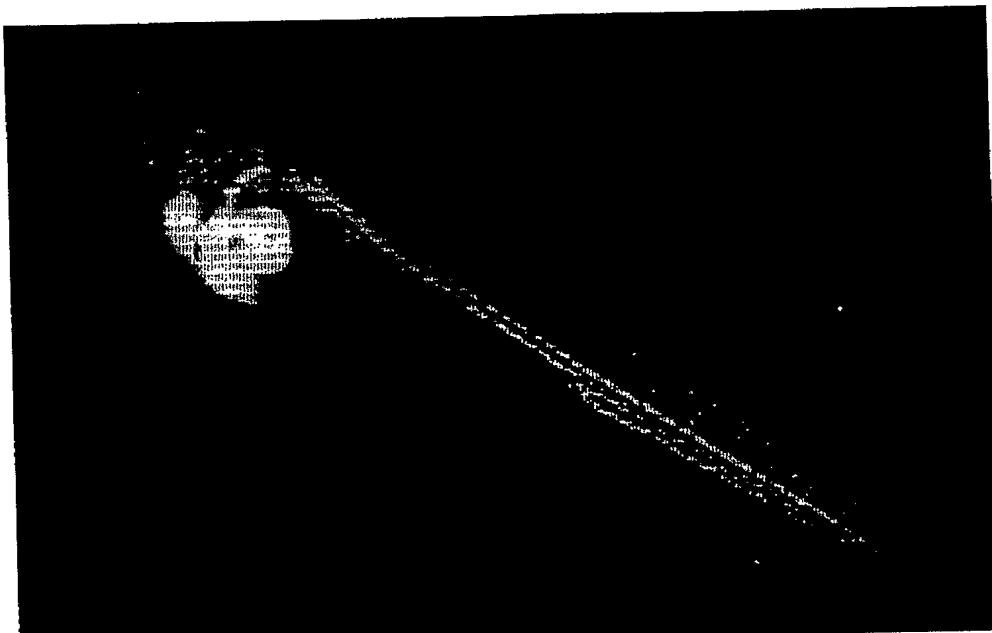


Figure 8

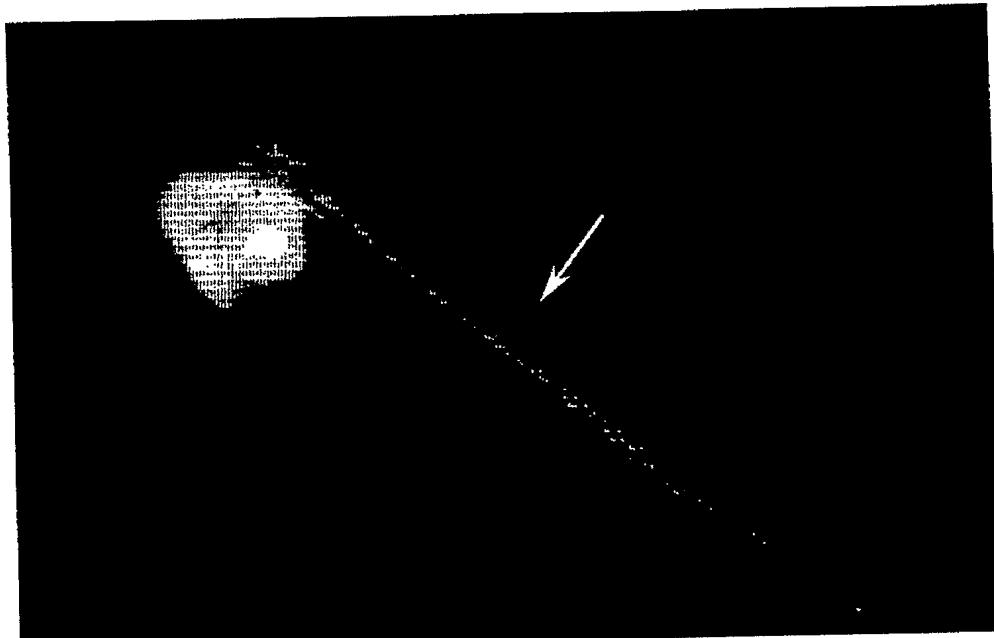


Figure 9

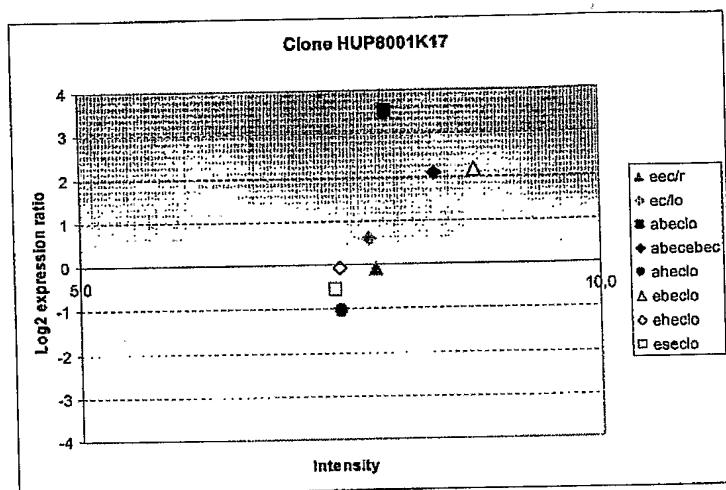


Figure 10

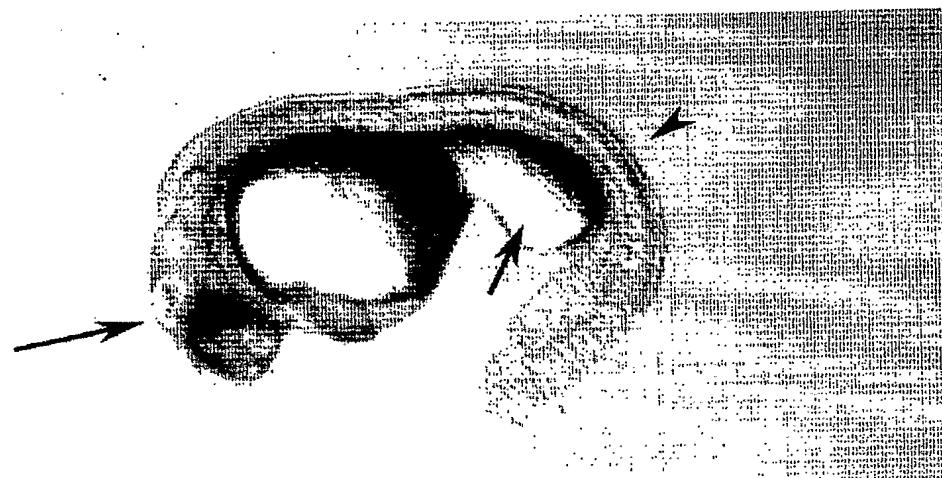


Figure 11

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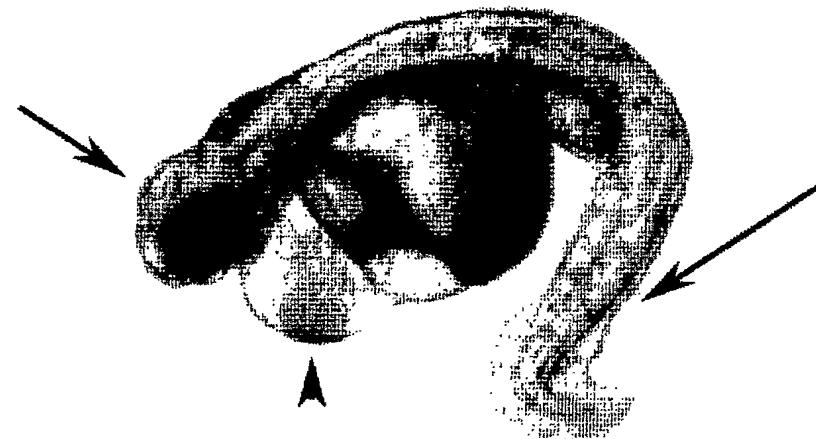


Figure 12

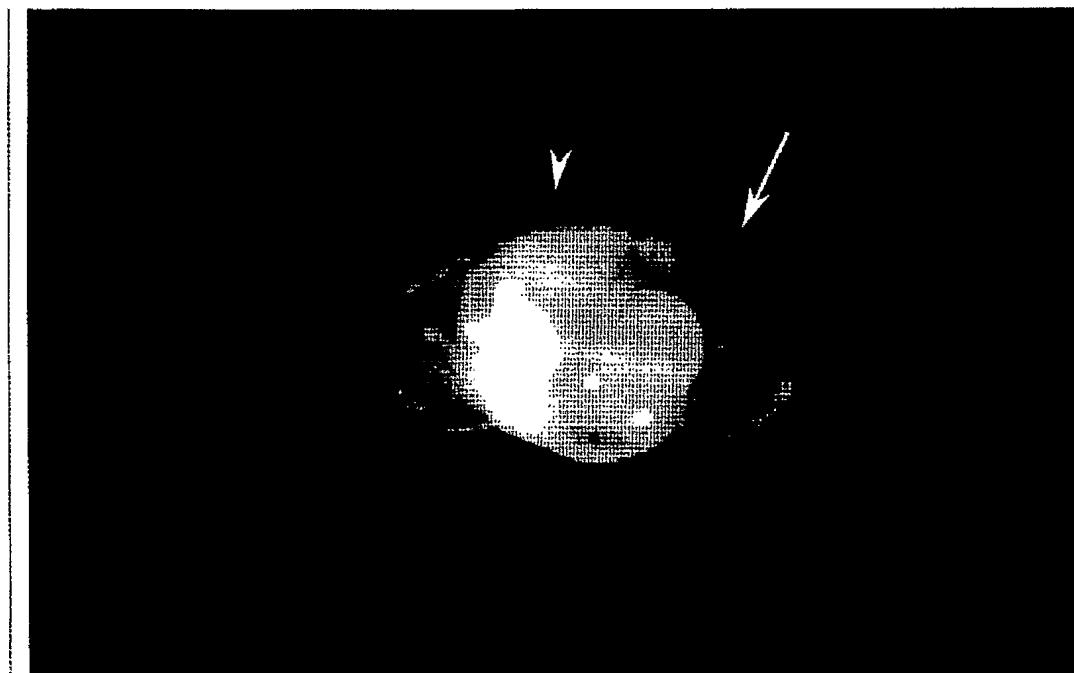


Figure 13

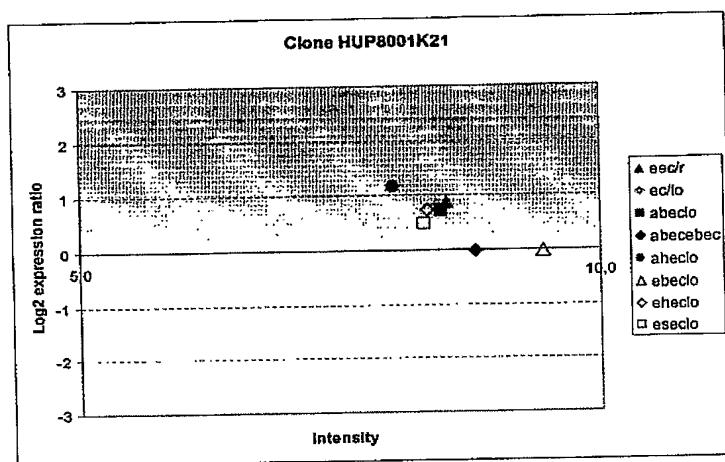


Figure 14

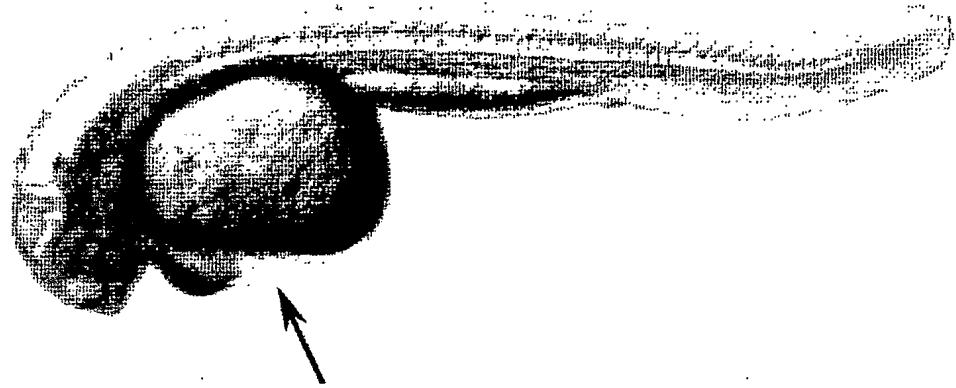


Figure 15

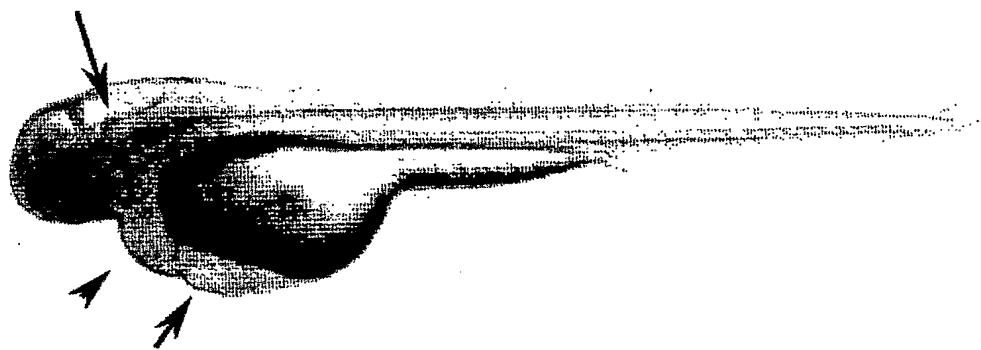
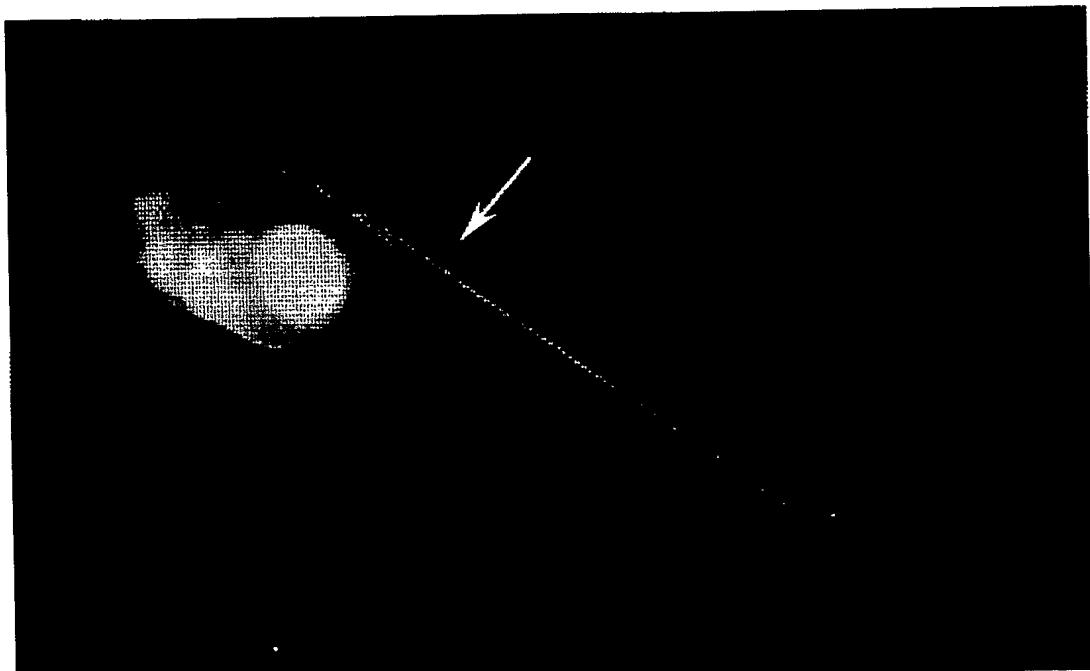


Figure 16



**Figure 17**

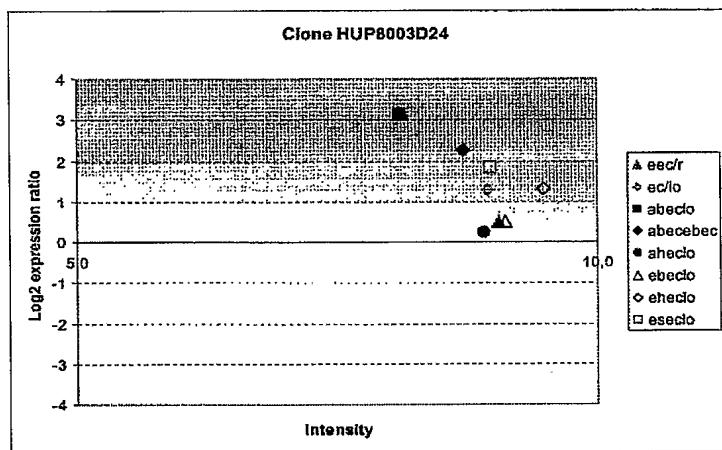


Figure 18

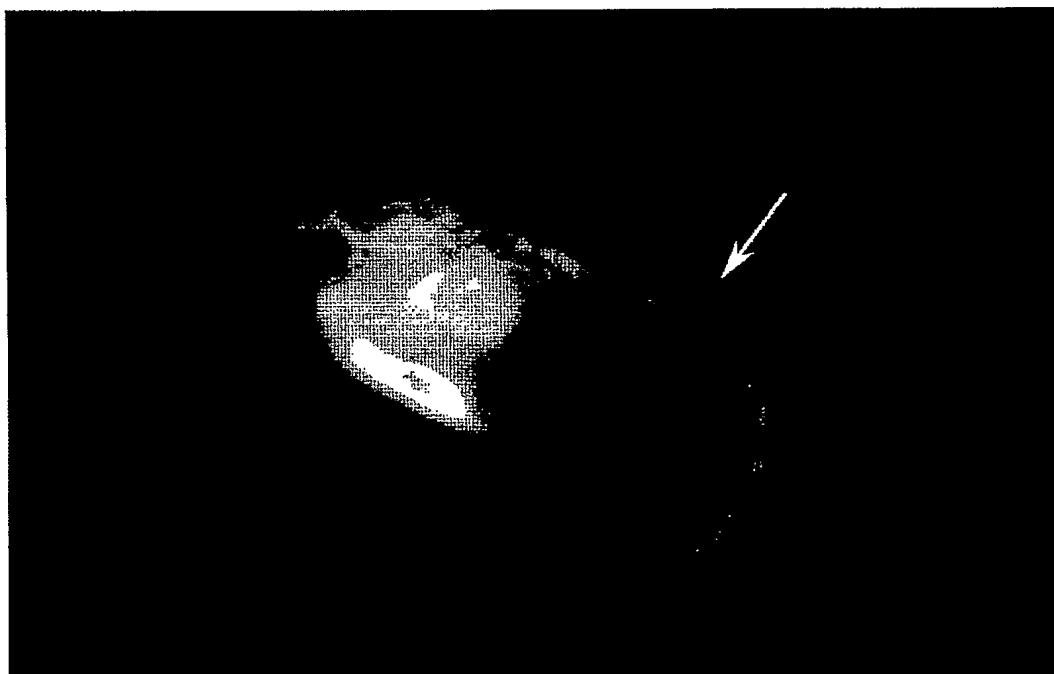


Figure 19

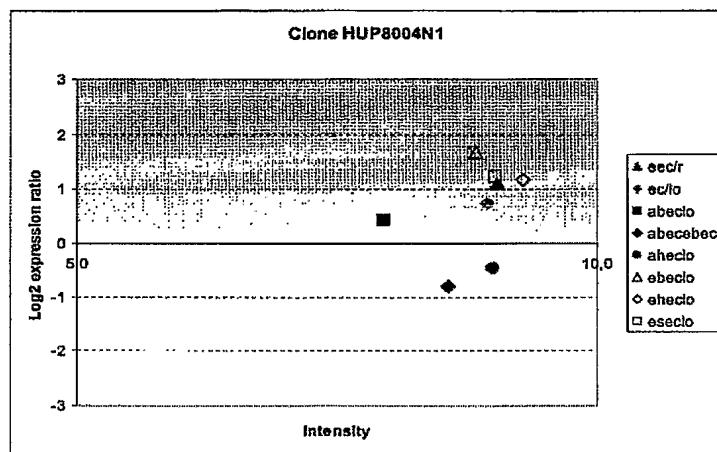


Figure 20



Figure 21

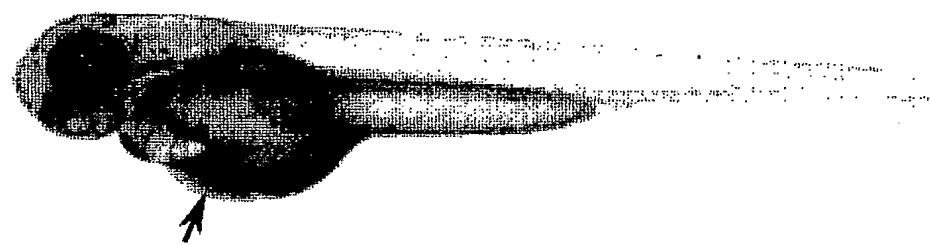


Figure 22

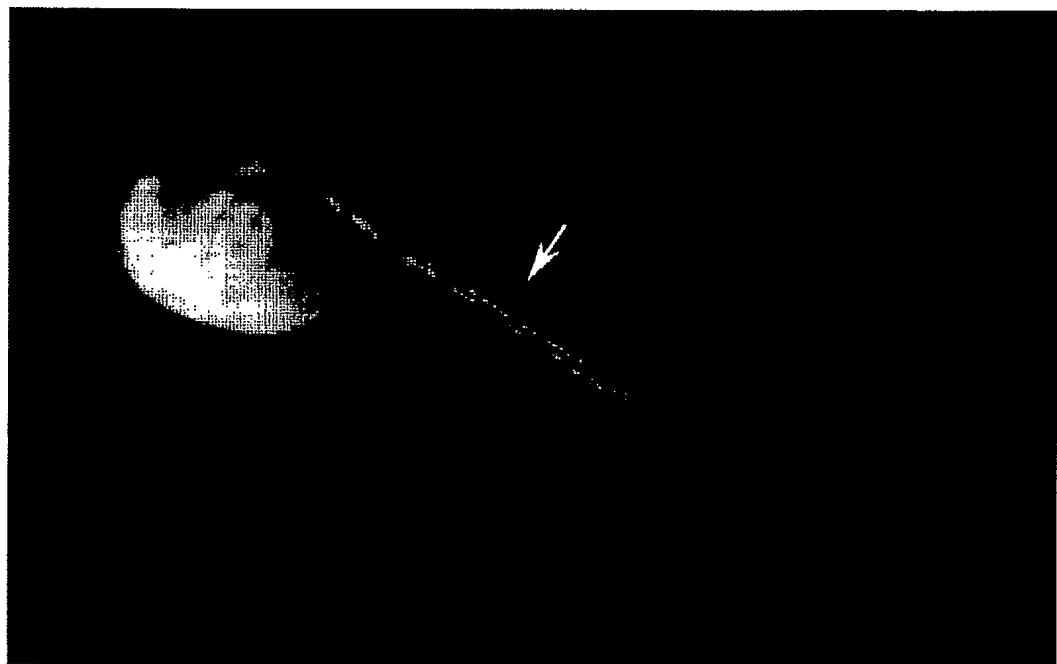


Figure 23

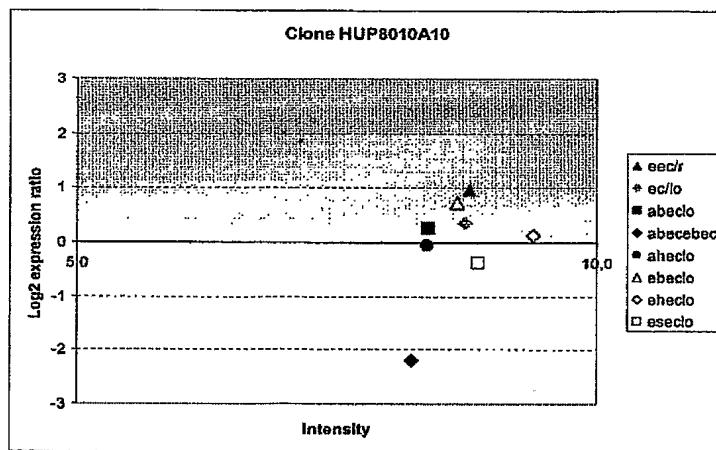


Figure 24

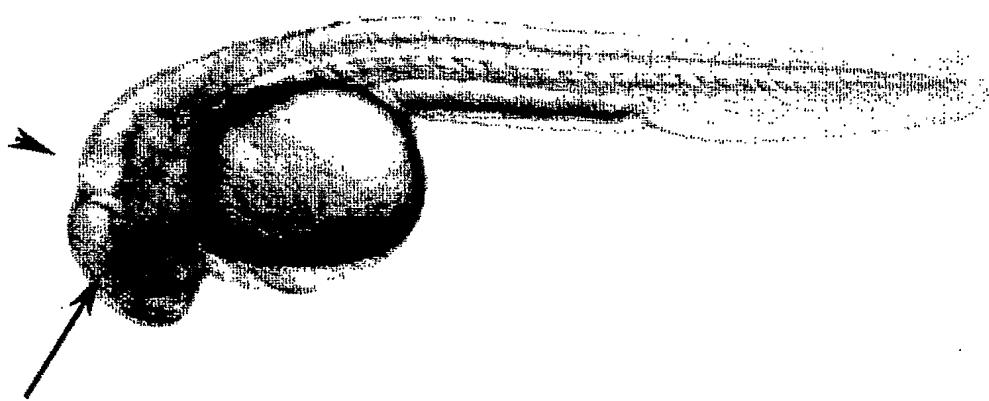


Figure 25

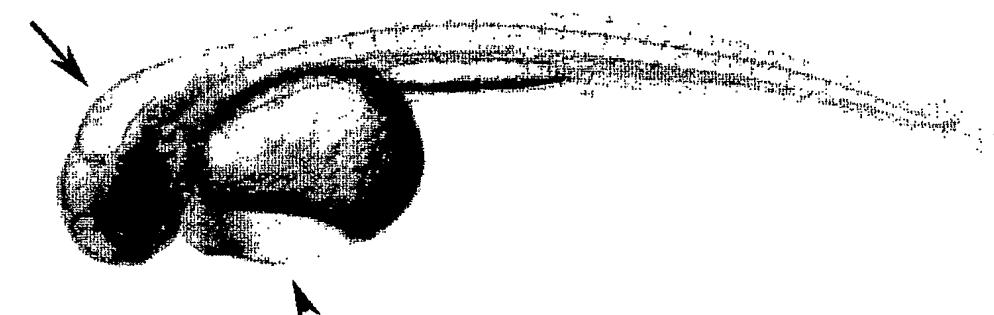


Figure 26

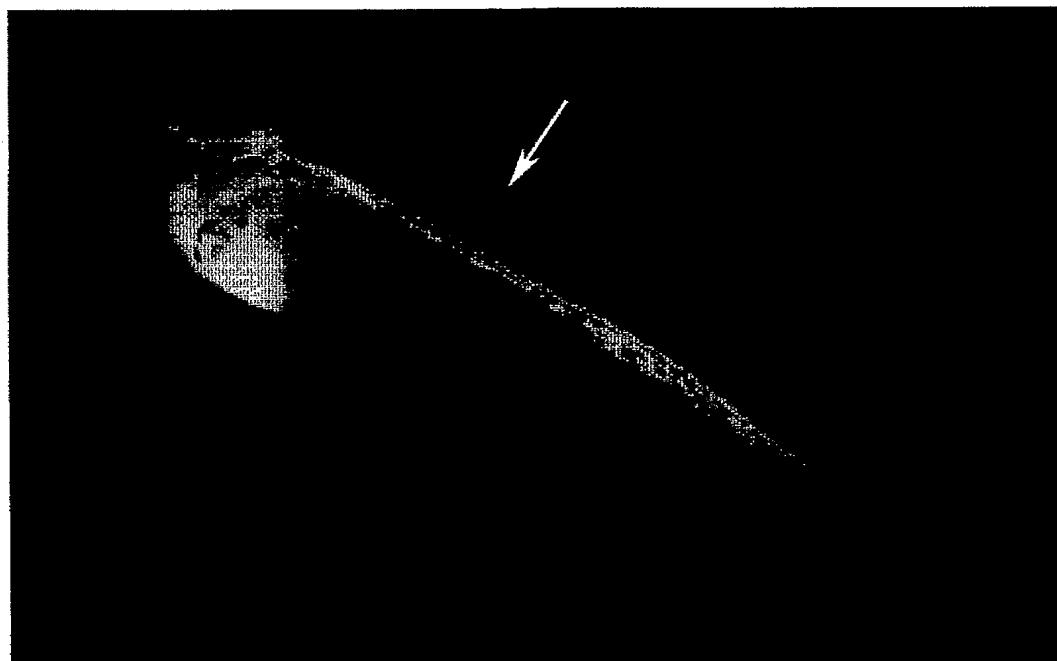


Figure 27

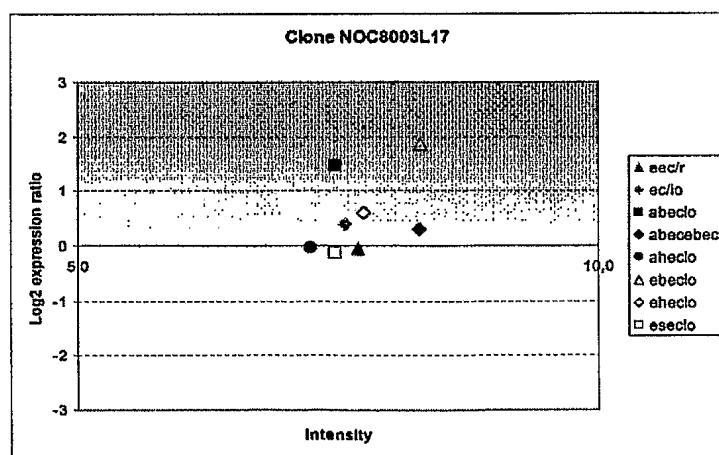


Figure 28

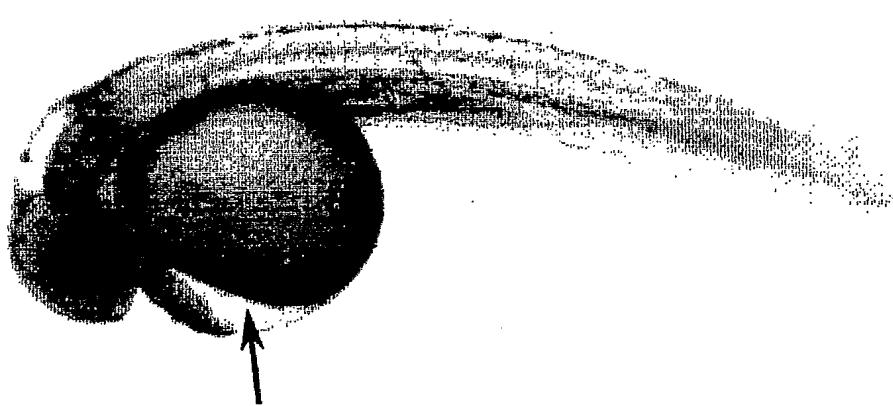


Figure 29

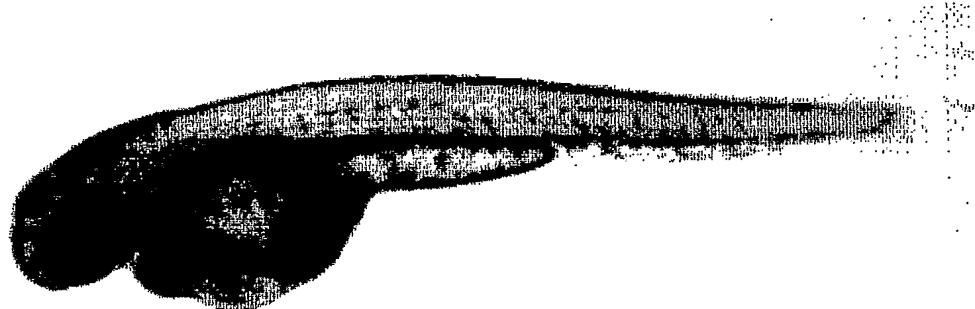


Figure 30

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Figure 31

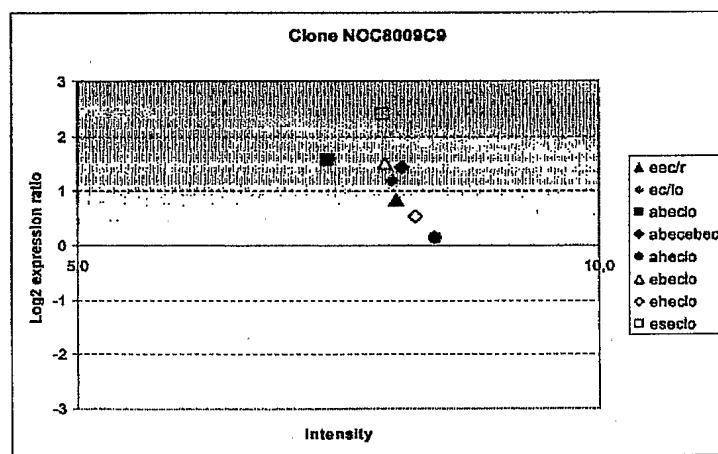


Figure 32

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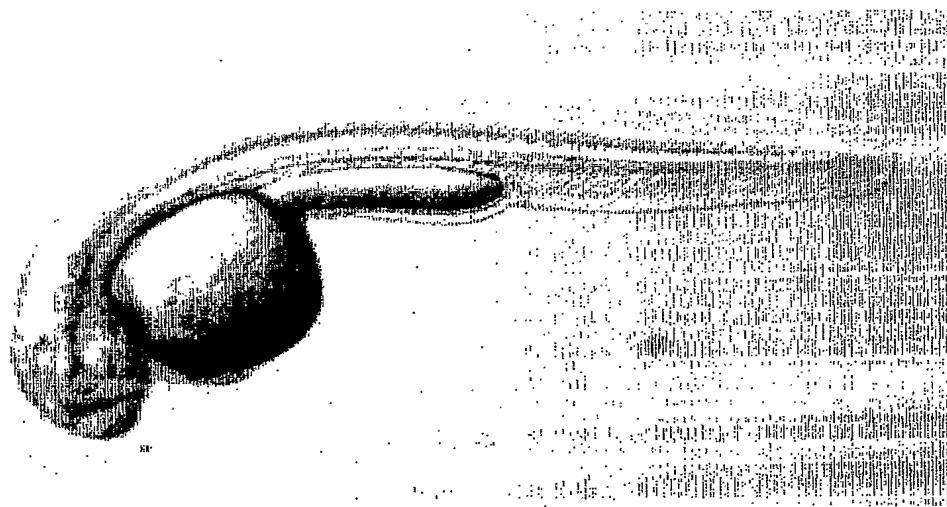


Figure 33

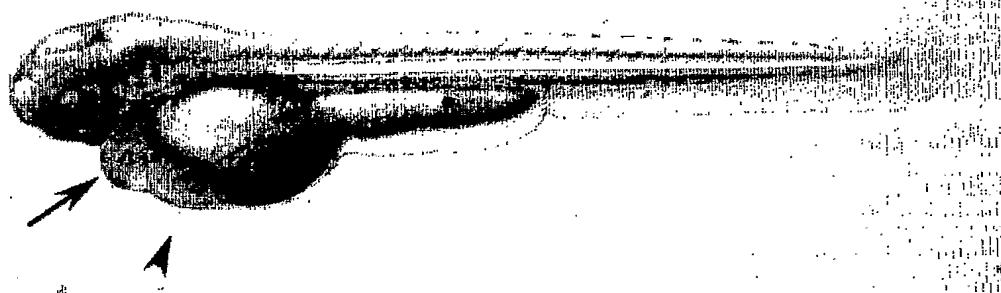


Figure 34

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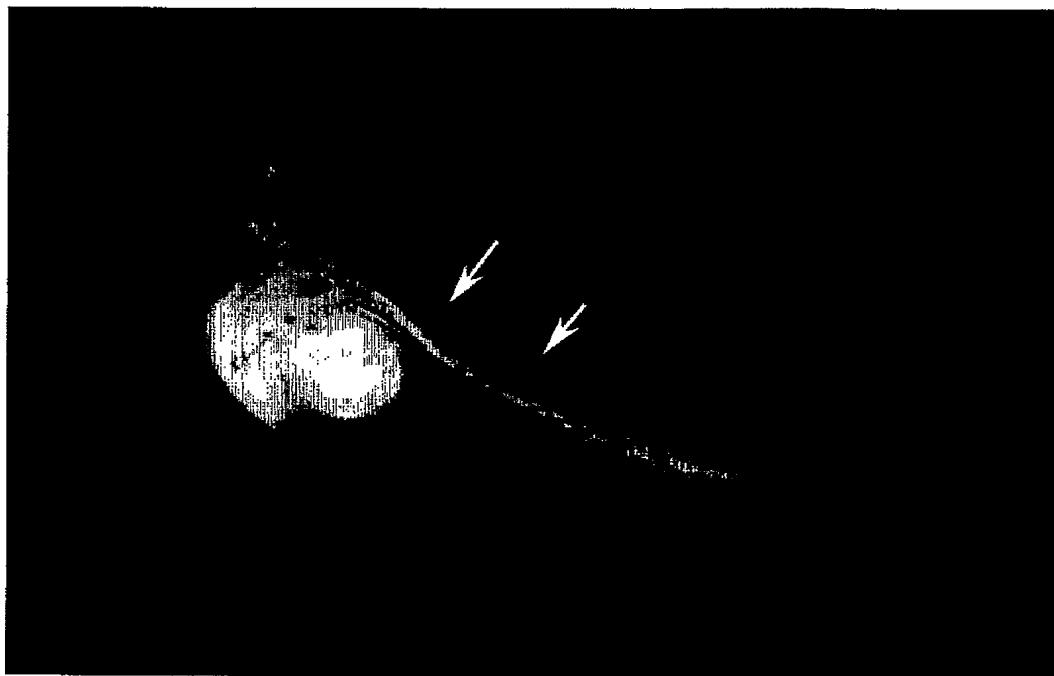


Figure 35

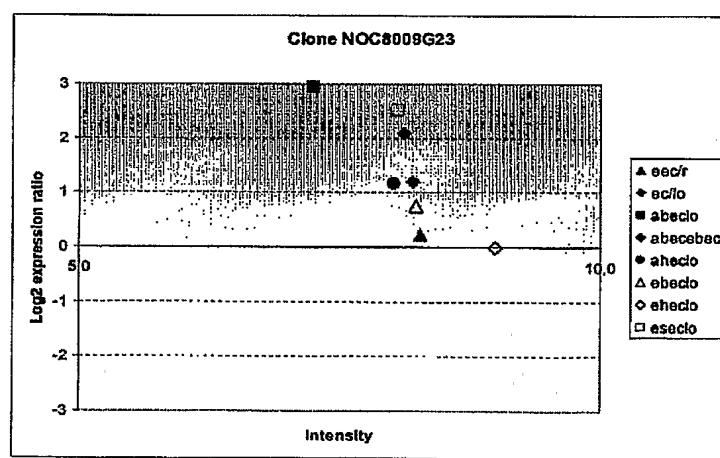


Figure 36

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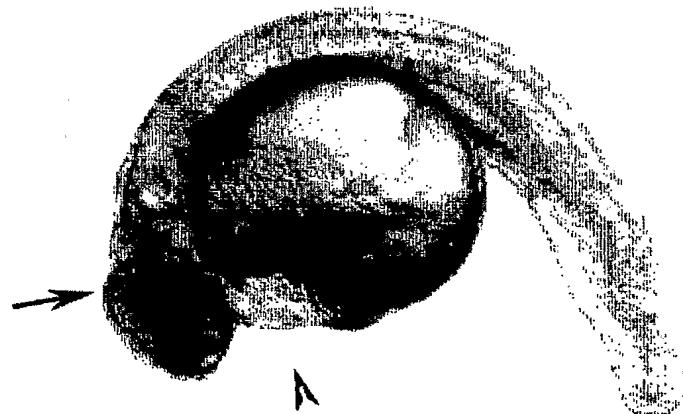


Figure 37

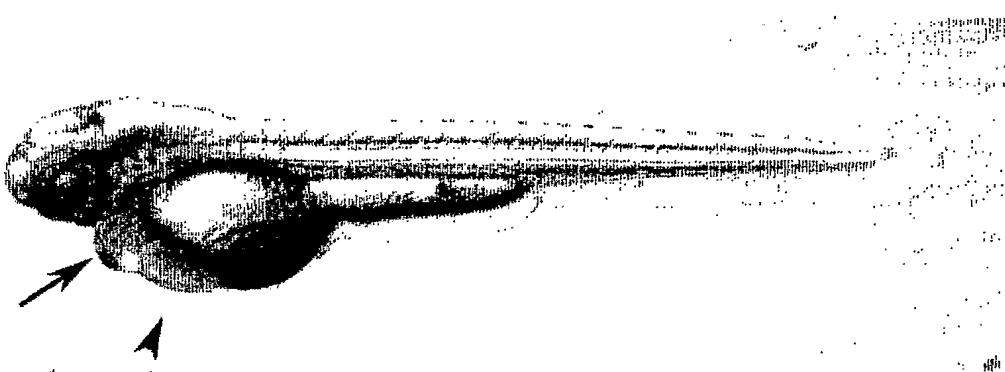


Figure 38

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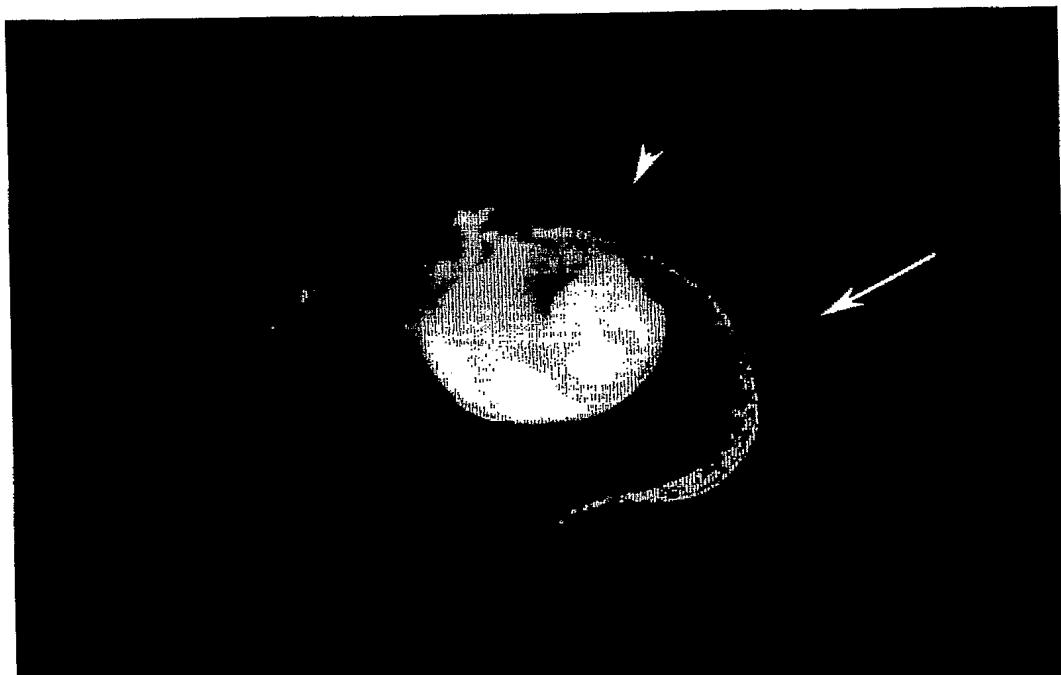


Figure 39

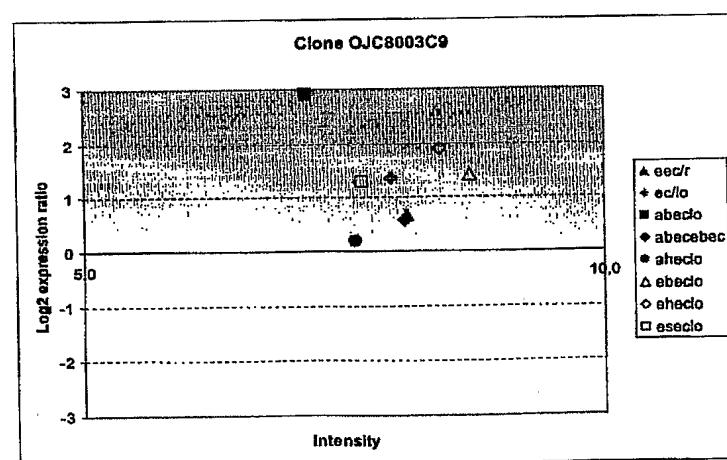


Figure 40



Figure 41

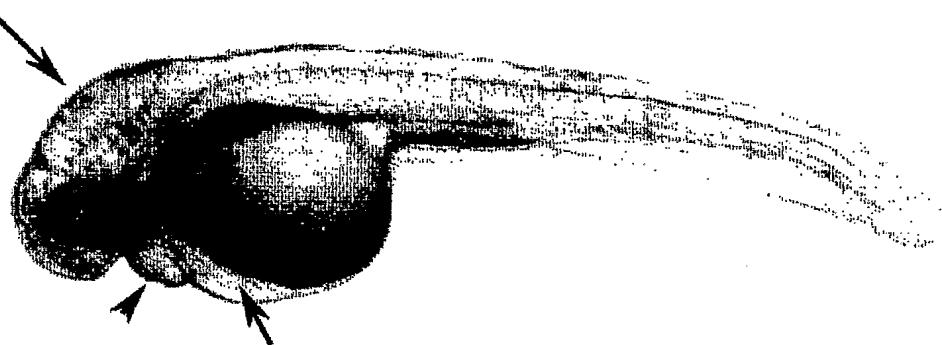


Figure 42

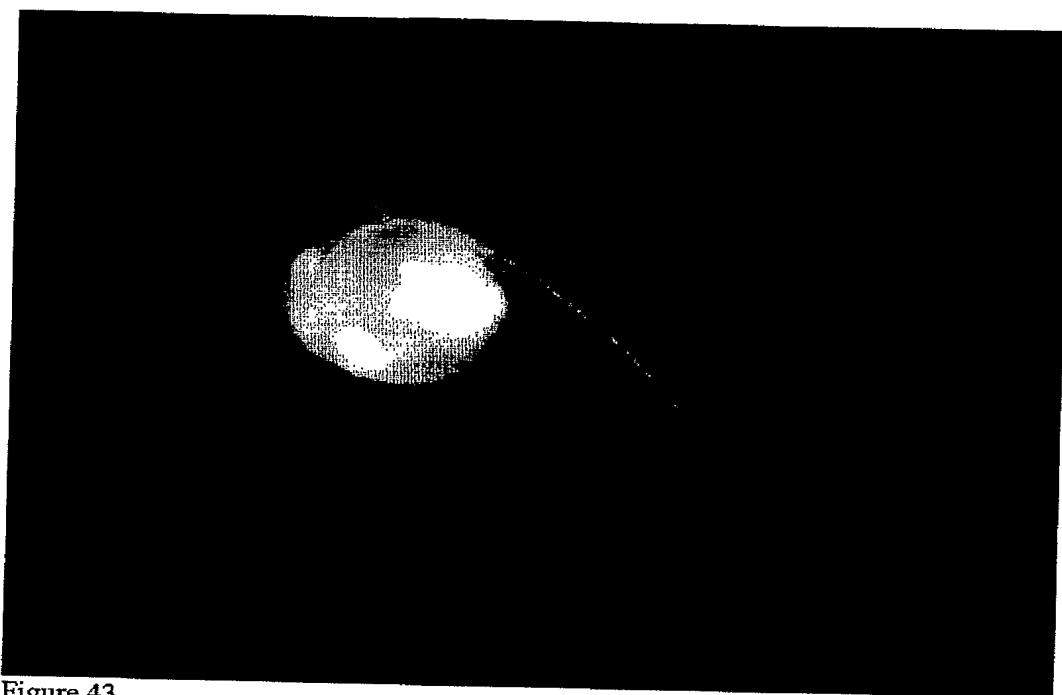


Figure 43

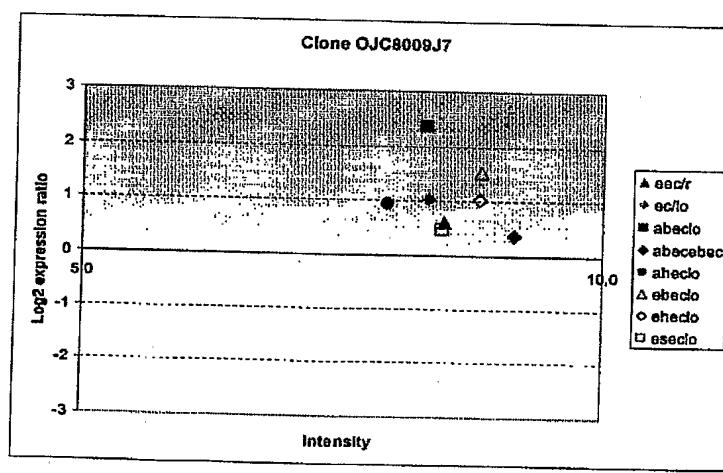


Figure 44

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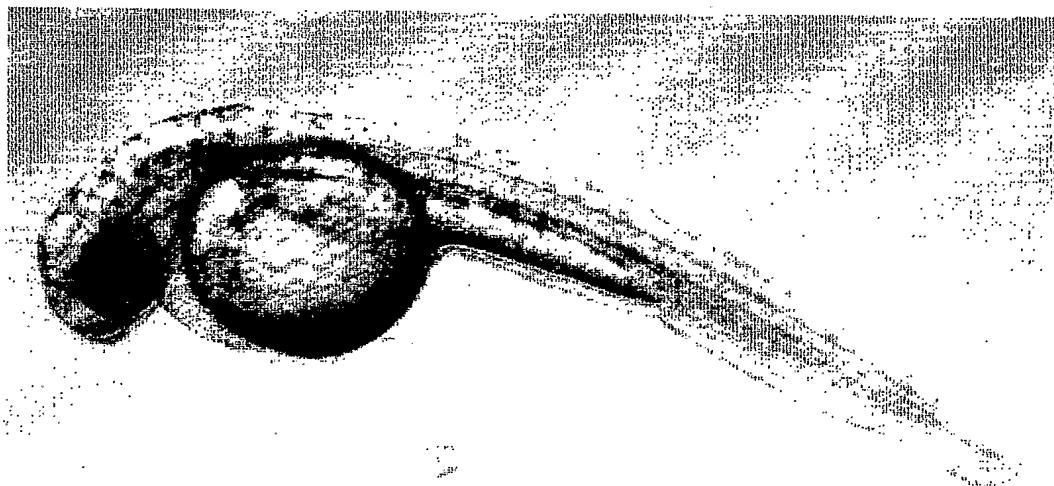


Figure 45

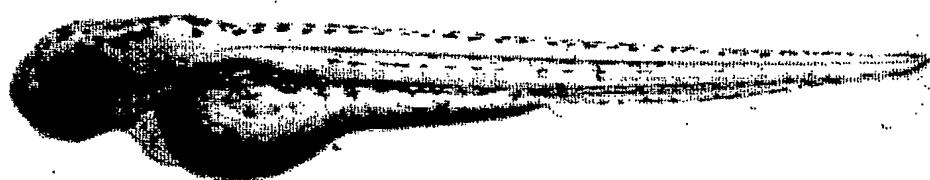


Figure 46

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<120> ANGIOGENESIS-AFFECTING POLYPEPTIDES, PROTEINS, AND  
COMPOSITIONS, AND METHODS OF USE THEREOF

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gatccctca tagaggaatt ttaattgata catctagaca cttcctgcgt gtgaagacaa  
240

ttttaaaaac tctggatgcc atggctttta ataagtttaa tggttttcac tggcacatag  
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420

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480

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<212> DNA  
<213> Murinae gen. sp.

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240

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atttcgagct gagccacagt tgcagaagct cctggtctcc attaccctcg agtcagagtg  
420

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540

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600

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900

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960

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caaagaaaatc agcagtgtgt ttccagatca gttcatccac ttgggaggag atgaagtaga  
1080

atttcaatgt tggcatcaa atccaaacat ccaagtttc atgaagagaa agggctttgg  
1140

PRIMED 1/19/93

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<211> 536  
<212> PRT  
<213> Murinae gen. sp.

<400> 8

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Pro Ala Leu Trp Pro Phe Pro Arg Ser Val Gln Met Phe Pro Arg Leu  
35 40 45

Leu Tyr Ile Ser Ala Glu Asp Phe Ser Ile Asp His Ser Pro Asn Ser  
50 55 60

Thr Ala Gly Pro Ser Cys Ser Leu Leu Gln Glu Ala Phe Arg Arg Tyr  
65 70 75 80

Tyr Asn Tyr Val Phe Gly Phe Tyr Lys Arg His His Gly Pro Ala Arg  
85 90 95

Phe Arg Ala Glu Pro Gln Leu Gln Lys Leu Leu Val Ser Ile Thr Leu  
100 105 110

Glu Ser Glu Cys Glu Ser Phe Pro Ser Leu Ser Ser Asp Glu Thr Tyr  
115 120 125

Ser Leu Leu Val Gln Glu Pro Val Ala Val Leu Lys Ala Asn Ser Val  
130 135 140

Trp Gly Ala Leu Arg Gly Leu Glu Thr Phe Ser Gln Leu Val Tyr Gln  
145 150 155 160

Asp Ser Phe Gly Thr Phe Thr Ile Asn Glu Ser Ser Ile Ala Asp Ser  
165 170 175

Pro Arg Phe Pro His Arg Gly Ile Leu Ile Asp Thr Ser Arg His Phe  
180 185 190

Leu Pro Val Lys Thr Ile Leu Lys Thr Leu Asp Ala Met Ala Phe Asn  
195 200 205

Lys Phe Asn Val Leu His Trp His Ile Val Asp Asp Gln Ser Phe Pro  
210 215 220

Tyr Gln Ser Thr Thr Phe Pro Glu Leu Ser Asn Lys Gly Ser Tyr Ser  
225 230 235 240

Leu Ser His Val Tyr Thr Pro Asn Asp Val Arg Met Val Leu Glu Tyr  
245 250 255

Ala Arg Leu Arg Gly Ile Arg Val Ile Pro Glu Phe Asp Thr Pro Gly  
260 265 270

His Thr Gln Ser Trp Gly Lys Gly Gln Lys Asn Leu Leu Thr Pro Cys  
275 280 285

Tyr Asn Gln Lys Thr Lys Thr Gln Val Phe Gly Pro Val Asp Pro Thr  
290 295 300

Val Asn Thr Thr Tyr Ala Phe Phe Asn Thr Phe Phe Lys Glu Ile Ser

305

310

315

320

Ser Val Phe Pro Asp Gln Phe Ile His Leu Gly Gly Asp Glu Val Glu  
325 330 335

Phe Gln Cys Trp Ala Ser Asn Pro Asn Ile Gln Gly Phe Met Lys Arg  
340 345 350

Lys Gly Phe Gly Ser Asp Phe Arg Arg Leu Glu Ser Phe Tyr Ile Lys  
355 360 365

Lys Ile Leu Glu Ile Ile Ser Ser Leu Lys Lys Asn Ser Ile Val Trp  
370 375 380

Gln Glu Val Phe Asp Asp Lys Val Glu Leu Gln Pro Gly Thr Val Val  
385 390 395 400

Glu Val Trp Lys Ser Glu His Tyr Ser Tyr Glu Leu Lys Gln Val Thr  
405 410 415

Gly Ser Gly Phe Pro Ala Ile Leu Ser Ala Pro Trp Tyr Leu Asp Leu  
420 425 430

Ile Ser Tyr Gly Gln Asp Trp Lys Asn Tyr Tyr Lys Val Glu Pro Leu  
435 440 445

Asn Phe Glu Gly Ser Glu Lys Gln Lys Gln Leu Val Ile Gly Gly Glu  
450 455 460

Ala Cys Leu Trp Gly Glu Phe Val Asp Ala Thr Asn Leu Thr Pro Arg  
465 470 475 480

Leu Trp Pro Arg Ala Ser Ala Val Gly Glu Arg Leu Trp Ser Pro Lys  
485 490 495

Thr Val Thr Asp Leu Glu Asn Ala Tyr Lys Arg Leu Ala Val His Arg  
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Cys Arg Met Val Ser Arg Gly Ile Ala Ala Gln Pro Leu Tyr Thr Gly  
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Tyr Cys Asn Tyr Glu Asn Lys Ile  
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<211> 1746

<212> DNA  
<213> Homo sapiens  
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1200

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1260

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1320

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1380

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1620

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1746

<210> 10  
<211> 556  
<212> PRT  
<213> Homo sapiens

<400> 10

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Leu Leu Leu Ala Thr Leu Leu Ala Ala Met Leu Ala Leu Leu Thr Gln  
20 25 30

Val Ala Leu Val Val Gln Val Ala Glu Ala Ala Arg Ala Pro Ser Val  
35 40 45

Ser Ala Lys Pro Gly Pro Ala Leu Trp Pro Leu Pro Leu Leu Val Lys  
50 55 60

Met Thr Pro Asn Leu Leu His Leu Ala Pro Glu Asn Phe Tyr Ile Ser  
65 70 75 80

His Ser Pro Asn Ser Thr Ala Gly Pro Ser Cys Thr Leu Leu Glu Glu  
85 90 95

Ala Phe Arg Arg Tyr His Gly Tyr Ile Phe Gly Phe Tyr Lys Trp His  
100 105 110

His Glu Pro Ala Glu Phe Gln Ala Lys Thr Gln Val Gln Gln Leu Leu  
115 120 125

Val Ser Ile Thr Leu Gln Ser Glu Cys Asp Ala Phe Pro Asn Ile Ser  
130 135 140

Ser Asp Glu Ser Tyr Thr Leu Leu Val Lys Glu Pro Val Ala Val Leu  
145 150 155 160

Lys Ala Asn Arg Val Trp Gly Ala Leu Arg Gly Leu Glu Thr Phe Ser  
165 170 175

Gln Leu Val Tyr Gln Asp Ser Tyr Gly Thr Phe Thr Ile Asn Glu Ser  
180 185 190

Thr Ile Ile Asp Ser Pro Arg Phe Ser His Arg Gly Ile Leu Ile Asp  
195 200 205

Thr Ser Arg His Tyr Leu Pro Val Lys Ile Ile Leu Lys Thr Leu Asp  
210 215 220

Ala Met Ala Phe Asn Lys Phe Asn Val Leu His Trp His Ile Val Asp  
225 230 235 240

Asp Gln Ser Phe Pro Tyr Gln Ser Ile Thr Phe Pro Glu Leu Ser Asn  
245 250 255

Lys Gly Ser Tyr Ser Leu Ser His Val Tyr Thr Pro Asn Asp Val Arg  
260 265 270

Met Val Ile Glu Tyr Ala Arg Leu Arg Gly Ile Arg Val Leu Pro Glu  
275 280 285

Phe Asp Thr Pro Gly His Thr Leu Ser Trp Gly Lys Gly Gln Lys Asp  
290 295 300

Leu Leu Thr Pro Cys Tyr Ser Arg Gln Asn Lys Leu Asp Ser Phe Gly  
305 310 315 320

Pro Ile Asn Pro Thr Leu Asn Thr Thr Tyr Ser Phe Leu Thr Thr Phe

325

330

335

Phe Lys Glu Ile Ser Glu Val Phe Pro Asp Gln Phe Ile His Leu Gly  
 340 345 350

Gly Asp Glu Val Glu Phe Lys Cys Trp Glu Ser Asn Pro Lys Ile Gln  
 355 360 365

Asp Phe Met Arg Gln Lys Gly Phe Gly Thr Asp Phe Lys Lys Leu Glu  
 370 375 380

Ser Phe Tyr Ile Gln Lys Val Leu Asp Ile Ile Ala Thr Ile Asn Lys  
 385 390 395 400

Gly Ser Ile Val Trp Gln Glu Val Phe Asp Asp Lys Ala Lys Leu Ala  
 405 410 415

Pro Gly Thr Ile Val Glu Val Trp Lys Asp Ser Ala Tyr Pro Glu Glu  
 420 425 430

Leu Ser Arg Val Thr Ala Ser Gly Phe Pro Val Ile Leu Ser Ala Pro  
 435 440 445

Trp Tyr Leu Asp Leu Ile Ser Tyr Gly Gln Asp Trp Arg Lys Tyr Tyr  
 450 455 460

Lys Val Glu Pro Leu Asp Phe Gly Thr Gln Lys Gln Lys Gln Leu  
 465 470 475 480

Phe Ile Gly Gly Glu Ala Cys Leu Trp Gly Glu Tyr Val Asp Ala Thr  
 485 490 495

Asn Leu Thr Pro Arg Leu Trp Pro Arg Ala Ser Ala Val Gly Glu Arg  
 500 505 510

Leu Trp Ser Ser Lys Asp Val Arg Asp Met Asp Asp Ala Tyr Asp Arg  
 515 520 525

Leu Thr Arg His Arg Cys Arg Met Val Glu Arg Gly Ile Ala Ala Gln  
 530 535 540

Pro Leu Tyr Ala Gly Tyr Cys Asn His Glu Asn Met  
 545 550 555

<210> 11  
 <211> 676

<212> DNA  
<213> Murinae gen. sp.

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<400> 11

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120

gacagagaac gagatccgtg gtctgtgcct caaatcccgg gagattttcc tgagccagcc  
180

cattcttctg gagcttgagg cgcgcctcaa gatctgtggt gacatccatg gccagtacta  
240

tgaccttcta cggctgtttg agtatggtgg cttccctcca gagagcaact acctcttctt  
300

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360

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420

caaccgcatt tatggcttct atgatgaatg caagagaaga tacaacatca aactgtggaa  
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<210> 12  
<211> 1369  
<212> DNA  
<213> Murinae gen. sp.

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120

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180

gattttcctg agccagccca ttcttctgga gcttgaggcg cccctcaaga tctgtggta  
240  
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ccatgagtgt gccagcatca accgcattta tggcttctat gatgaatgca agagaagata  
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780  
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<211> 330  
<212> PRT  
<213> Murinae gen. sp.

<400> 13

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Leu Glu Val Gln Gly Ser Arg Pro Gly Lys Asn Val Gln Leu Thr Glu  
20 25 30

Asn Glu Ile Arg Gly Leu Cys Leu Lys Ser Arg Glu Ile Phe Leu Ser  
35 40 45

Gln Pro Ile Leu Leu Glu Leu Glu Ala Pro Leu Lys Ile Cys Gly Asp  
50 55 60

Ile His Gly Gln Tyr Tyr Asp Leu Leu Arg Leu Phe Glu Tyr Gly Gly  
65 70 75 80

Phe Pro Pro Glu Ser Asn Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg  
85 90 95

Gly Lys Gln Ser Leu Glu Thr Ile Cys Leu Leu Leu Ala Tyr Lys Ile  
100 105 110

Arg Tyr Pro Glu Asn Phe Phe Leu Leu Arg Gly Asn His Glu Cys Ala  
115 120 125

Ser Ile Asn Arg Ile Tyr Gly Phe Tyr Asp Glu Cys Lys Arg Arg Tyr  
130 135 140

Asn Ile Lys Leu Trp Lys Thr Phe Thr Asp Cys Phe Asn Cys Leu Pro  
145 150 155 160

Ile Ala Ala Ile Val Asp Glu Lys Ile Phe Cys Cys His Gly Gly Leu  
165 170 175

Ser Pro Asp Leu Gln Ser Met Glu Gln Ile Arg Arg Ile Met Arg Pro  
180 185 190

Thr Asp Val Pro Asp Gln Gly Leu Leu Cys Asp Leu Leu Trp Ser Asp  
195 200 205

Pro Asp Lys Asp Val Gln Gly Trp Gly Glu Asn Asp Arg Gly Val Ser

PRIMER 1000

210

215

220

Phe Thr Phe Gly Ala Glu Val Val Ala Lys Phe Leu His Lys His Asp  
225 230 235 240

Leu Asp Leu Ile Cys Arg Ala His Gln Val Val Glu Asp Gly Tyr Glu  
245 250 255

Phe Phe Ala Lys Arg Gln Leu Val Thr Leu Phe Ser Ala Pro Asn Tyr  
260 265 270

Cys Gly Glu Phe Asp Asn Ala Gly Ala Met Met Ser Val Asp Glu Thr  
275 280 285

Leu Met Cys Ser Phe Gln Ile Leu Lys Pro Ala Asp Lys Asn Lys Gly  
290 295 300

Lys Tyr Gly Gln Phe Ser Gly Leu Asn Pro Gly Gly Arg Pro Ile Thr  
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Pro Pro Arg Asn Ser Ala Lys Ala Lys Lys  
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<210> 14

<211> 993

<212> DNA

<213> Homo sapiens

<400> 14

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120

aaatcccgaa agattttct gagccagccc attcttctgg agctggaggc acccctcaag  
180

atctgcggtg acatacaccg ccagtactac gaccttctgc gactatggta gtatggcggt  
240

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360

ctccgtggaa accacgagtg tgccagcatc aaccgcacatct atggttctta cgatgagtgc  
420

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480

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<210> 15  
<211> 330  
<212> PRT  
<213> Homo sapiens  
  
<400> 15

Met Ser Asp Ser Glu Lys Leu Asn Leu Asp Ser Ile Ile Gly Arg Leu  
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Leu Glu Val Gln Gly Ser Arg Pro Gly Lys Asn Val Gln Leu Thr Glu  
20 25 30

Asn Glu Ile Arg Gly Leu Cys Leu Lys Ser Arg Glu Ile Phe Leu Ser  
35 40 45

Gln Pro Ile Leu Leu Glu Leu Glu Ala Pro Leu Lys Ile Cys Gly Asp  
50 55 60

Ile His Gly Gln Tyr Tyr Asp Leu Leu Arg Leu Phe Glu Tyr Gly Gly  
65 70 75 80

Phe Pro Pro Glu Ser Asn Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg  
85 90 95

Gly Lys Gln Ser Leu Glu Thr Ile Cys Leu Leu Ala Tyr Lys Ile  
100 105 110

Lys Tyr Pro Glu Asn Phe Phe Leu Leu Arg Gly Asn His Glu Cys Ala  
115 120 125

Ser Ile Asn Arg Ile Tyr Gly Phe Tyr Asp Glu Cys Lys Arg Arg Tyr  
130 135 140

Asn Ile Lys Leu Trp Lys Thr Phe Thr Asp Cys Phe Asn Cys Leu Pro  
145 150 155 160

Ile Ala Ala Ile Val Asp Glu Lys Ile Phe Cys Cys His Gly Gly Leu  
165 170 175

Ser Pro Asp Leu Gln Ser Met Glu Gln Ile Arg Arg Ile Met Arg Pro  
180 185 190

Thr Asp Val Pro Asp Gln Gly Leu Leu Cys Asp Leu Leu Trp Ser Asp  
195 200 205

Pro Asp Lys Asp Val Gln Gly Trp Gly Glu Asn Asp Arg Gly Val Ser  
210 215 220

Phe Thr Phe Gly Ala Glu Val Val Ala Lys Phe Leu His Lys His Asp  
225 230 235 240

Leu Asp Leu Ile Cys Arg Ala His Gln Val Val Glu Asp Gly Tyr Glu  
245 250 255

Phe Phe Ala Lys Arg Gln Leu Val Thr Leu Phe Ser Ala Pro Asn Tyr  
260 265 270

Cys Gly Glu Phe Asp Asn Ala Gly Ala Met Met Ser Val Asp Glu Thr  
275 280 285

Leu Met Cys Ser Phe Gln Ile Leu Lys Pro Ala Asp Lys Asn Lys Gly  
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Lys Tyr Gly Gln Phe Ser Gly Leu Asn Pro Gly Gly Arg Pro Ile Thr  
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Pro Pro Arg Asn Ser Ala Lys Ala Lys Lys  
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<210> 16  
<211> 702  
<212> DNA

<213> Murinae gen. sp.

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420

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600

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<210> 17

<211> 1432

<212> DNA

<213> Murinae gen. sp.

<400> 17

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<211> 378  
<212> PRT

<213> Murinae gen. sp.

<400> 18

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Arg Pro Asn Pro Asn Cys Pro Gly Pro Cys Ser Arg Gln Ser Lys Arg  
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Leu Ala Val Ala Trp Gly Gly Arg His Pro Glu Gly Ala Leu Gly  
35 40 45

Ile Gly Tyr Leu Asp Arg Arg Gly Leu Phe Leu Pro Pro Leu Ala Pro  
50 55 60

Gly Gly Asp Thr Ile Gln Pro Val Thr Met Phe Asp Lys Thr Arg Leu  
65 70 75 80

Pro Tyr Val Ala Leu Asp Val Ile Cys Val Leu Leu Ala Gly Leu Pro  
85 90 95

Phe Ala Ile Leu Thr Ser Arg His Thr Pro Phe Gln Arg Gly Ile Phe  
100 105 110

Cys Asn Asp Asp Ser Ile Lys Tyr Pro Tyr Lys Glu Asp Thr Ile Pro  
115 120 125

Tyr Ala Leu Leu Gly Gly Ile Val Ile Pro Phe Cys Ile Ile Val Met  
130 135 140

Ser Ile Gly Glu Ser Leu Ser Val Tyr Phe Asn Val Leu His Ser Asn  
145 150 155 160

Ser Phe Val Gly Asn Pro Tyr Ile Ala Thr Ile Tyr Lys Ala Val Gly  
165 170 175

Ala Phe Leu Phe Gly Val Ser Ala Ser Gln Ser Leu Thr Asp Ile Ala  
180 185 190

Lys Tyr Thr Ile Gly Ser Leu Arg Pro His Phe Leu Ala Ile Cys Asn  
195 200 205

Pro Asp Trp Ser Lys Ile Asn Cys Ser Asp Gly Tyr Ile Glu Asp Tyr  
210 215 220

Ile Cys Gln Gly Asn Glu Glu Lys Val Lys Glu Gly Arg Leu Ser Phe

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Asp Phe Phe Lys Asp Thr His Ser Tyr Lys Glu Arg Lys Glu Glu Asp			
320		335	
Pro His Thr Thr Leu His Glu Thr Ala Ser Ser Arg Asn Tyr Trp Ala			
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1620

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1626

<210> 20  
<211> 378  
<212> PRT  
<213> Homo sapiens

<400> 20

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Pro Leu Leu Ala Val Gly Ala Pro Pro Gly Leu Ser Pro Pro Ser Ala  
35 40 45

Ala Leu Leu Leu Arg Leu Gly Gly Ala Val Ala Arg Gly Arg Arg Gln  
50 55 60

Pro Arg Pro Gly Leu Glu Asn Gln Gly Pro Arg Pro Pro Ser Arg Ser  
65 70 75 80

Ser Val His Arg Pro Cys Arg Ala Ala Arg Ala Glu Thr Met Phe Asp  
85 90 95

Lys Thr Arg Leu Pro Tyr Val Ala Leu Asp Val Leu Cys Val Leu Leu  
100 105 110

Ala Ser Met Pro Met Ala Val Leu Lys Leu Gly Gln Ile Tyr Pro Phe  
115 120 125

Gln Arg Gly Phe Phe Cys Lys Asp Asn Ser Ile Asn Tyr Pro Tyr His  
130 135 140

Asp Ser Thr Val Thr Ser Thr Val Leu Ile Leu Val Gly Val Gly Leu  
145 150 155 160

Pro Ile Ser Ser Ile Ile Leu Gly Glu Thr Leu Ser Val Tyr Cys Asn  
165 170 175

Leu Leu His Ser Asn Ser Phe Ile Arg Asn Asn Tyr Ile Ala Thr Ile  
 180 185 190

Tyr Lys Ala Ile Gly Thr Phe Leu Phe Gly Ala Ala Ser Gln Ser  
 195 200 205

Leu Thr Asp Ile Ala Lys Tyr Ser Ile Gly Arg Leu Arg Pro His Phe  
 210 215 220

Leu Asp Val Cys Asp Pro Asp Trp Ser Lys Ile Asn Cys Ser Asp Gly  
 225 230 235 240

Tyr Ile Glu Tyr Tyr Ile Cys Arg Gly Asn Ala Glu Arg Val Lys Glu  
 245 250 255

Gly Arg Leu Ser Phe Tyr Ser Gly His Ser Ser Phe Ser Met Tyr Cys  
 260 265 270

Met Leu Phe Val Ala Leu Tyr Leu Gln Ala Arg Met Lys Gly Asp Trp  
 275 280 285

Ala Arg Leu Leu Arg Pro Thr Leu Gln Phe Gly Leu Val Ala Val Ser  
 290 295 300

Ile Tyr Val Gly Leu Ser Arg Val Ser Asp Tyr Lys His His Trp Ser  
 305 310 315 320

Asp Val Leu Thr Gly Leu Ile Gln Gly Ala Leu Val Ala Ile Leu Val  
 325 330 335

Ala Val Tyr Val Ser Asp Phe Phe Lys Glu Arg Thr Ser Phe Lys Glu  
 340 345 350

Arg Lys Glu Glu Asp Ser His Thr Thr Leu His Glu Thr Pro Thr Thr  
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Gly Asn His Tyr Pro Ser Asn His Gln Pro  
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<210> 21

<211> 816

<212> DNA

<213> Homo sapiens

<400> 21

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<210> 22  
<211> 271  
<212> PRT  
<213> Homo sapiens

<400> 22

Ile Tyr Ser Leu Leu Ala Gly Leu Pro Phe Ala Ile Leu Thr Ser  
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Arg His Thr Pro Phe Gln Arg Gly Val Phe Cys Asn Asp Glu Ser Ile  
20 25 30

Lys Tyr Pro Tyr Lys Glu Asp Thr Ile Pro Tyr Ala Leu Leu Gly Gly  
35 40 45

Ile Ile Ile Pro Phe Ser Ile Ile Val Ile Ile Leu Gly Glu Thr Leu  
50 55 60

Ser Val Tyr Cys Asn Leu Leu His Ser Asn Ser Phe Ile Arg Asn Asn  
65 70 75 80

Tyr Ile Ala Thr Ile Tyr Lys Ala Ile Gly Thr Phe Leu Phe Gly Ala  
85 90 95

Ala Ala Ser Gln Ser Leu Thr Asp Ile Ala Lys Tyr Ser Ile Gly Arg  
100 105 110

Leu Arg Pro His Phe Leu Asp Val Cys Asp Pro Asp Trp Ser Lys Ile  
115 120 125

Asn Cys Ser Asp Gly Tyr Ile Glu Tyr Tyr Ile Cys Arg Gly Asn Ala  
130 135 140

Glu Arg Val Lys Glu Gly Arg Leu Ser Phe Tyr Ser Gly His Ser Ser  
145 150 155 160

Phe Ser Met Tyr Cys Met Leu Phe Val Ala Leu Tyr Leu Gln Ala Arg  
165 170 175

Met Lys Gly Asp Trp Ala Arg Leu Leu Arg Pro Thr Leu Gln Phe Gly  
180 185 190

Leu Val Ala Val Ser Ile Tyr Val Gly Leu Ser Arg Val Ser Asp Tyr  
195 200 205

Lys His His Trp Ser Asp Val Leu Thr Gly Leu Ile Gln Gly Ala Leu  
210 215 220

Val Ala Ile Leu Val Ala Val Tyr Val Ser Asp Phe Phe Lys Glu Arg  
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Thr Ser Phe Lys Glu Arg Lys Glu Glu Asp Ser His Thr Thr Leu His  
245 250 255

Glu Thr Pro Thr Thr Gly Asn His Tyr Pro Ser Asn His Gln Pro  
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<210> 23  
<211> 840  
<212> DNA  
<213> Murinae gen. sp.

<220>



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<210> 25  
<211> 171  
<212> PRT  
<213> Murinae gen. sp.

<400> 25

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20

25

30

Gln Val Ile Leu Thr Glu Lys Asp Leu Gln Glu Asp Gly Phe Gly Glu  
 35 40 45

His Pro Phe Tyr His Cys Leu Val Ala Glu Val Pro Lys Glu His Trp  
 50 55 60

Thr Pro Glu Gly His Ser Ile Val Gly Phe Ala Met Tyr Tyr Phe Thr  
 65 70 75 80

Tyr Asp Pro Trp Ile Gly Lys Leu Leu Tyr Leu Glu Asp Phe Phe Val  
 85 90 95

Met Ser Asp Tyr Arg Gly Phe Gly Ile Gly Ser Glu Ile Leu Lys Asn  
 100 105 110

Leu Ser Gln Val Ala Met Lys Cys Arg Cys Ser Ser Met His Phe Leu  
 115 120 125

Val Ala Glu Trp Asn Glu Pro Ser Ile Asn Phe Tyr Lys Arg Arg Gly  
 130 135 140

Ala Ser Asp Leu Ser Ser Glu Glu Gly Trp Arg Leu Phe Lys Ile Asp  
 145 150 155 160

Lys Glu Tyr Leu Leu Lys Met Ala Ala Glu Glu  
 165 170

<210> 26

<211> 1111

<212> DNA

<213> Homo sapiens

<400> 26

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 120

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 180

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<212> PRT  
<213> Homo sapiens  
  
<400> 27

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20 25 30

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35 40 45

Lys Tyr Glu Tyr Met Glu Glu Gln Val Ile Leu Thr Glu Lys Asp Leu  
 50 55 60

Leu Glu Asp Gly Phe Gly Glu His Pro Phe Tyr His Cys Leu Val Ala  
 65 70 75 80

Glu Val Pro Lys Glu His Trp Thr Pro Glu Gly Asn Pro Ser Pro Phe  
 85 90 95

Pro Glu Ala Arg Glu Thr Asn Ile Val Gly Phe Ala Met Tyr Tyr Phe  
 100 105 110

Thr Tyr Asp Pro Trp Ile Gly Lys Leu Leu Tyr Leu Glu Asp Phe Phe  
 115 120 125

Val Met Ser Asp Tyr Arg Gly Thr Ile Glu Leu Trp His Arg Ile Arg  
 130 135 140

Asn Ser Glu Glu Ser Lys Pro Gly Cys Asn Glu Val Ser Leu Ala Ala  
 145 150 155 160

Cys Thr Ser Trp Ala Glu Trp Asn Glu Pro Ser Ile Asn Phe Tyr Lys  
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Arg Arg Gly Ala Ser Asp Leu Ser Ser Glu Glu Gly Trp Arg  
 180 185 190

<210> 28  
 <211> 745  
 <212> DNA  
 <213> Murinae gen. sp.

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360  
  
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<211> 323  
<212> PRT  
<213> Murinae gen. sp.

<400> 30

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Asn Glu Ile Arg Gly Leu Cys Leu Lys Ser Arg Glu Ile Phe Leu Ser  
35 40 45

Gln Pro Ile Leu Leu Glu Leu Glu Ala Pro Leu Lys Ile Cys Gly Asp  
50 55 60

Ile His Gly Gln Tyr Tyr Asp Leu Leu Arg Leu Phe Glu Tyr Gly Gly  
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Phe Pro Pro Glu Ser Asn Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg  
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Gly Lys Gln Ser Leu Glu Thr Ile Cys Leu Leu Leu Ala Tyr Lys Ile  
100 105 110

Lys Tyr Pro Glu Asn Phe Phe Leu Leu Arg Gly Asn His Glu Cys Ala  
115 120 125

Ser Ile Asn Arg Ile Tyr Gly Phe Tyr Asp Glu Cys Lys Arg Arg Tyr  
130 135 140

Asn Ile Lys Leu Trp Lys Thr Phe Thr Asp Cys Phe Asn Cys Leu Pro  
145 150 155 160

Ile Ala Ala Ile Val Asp Glu Lys Ile Phe Cys Cys His Gly Gly Leu  
 165 170 175

Ser Pro Asp Leu Gln Ser Met Glu Gln Ile Arg Arg Ile Met Arg Pro  
 180 185 190

Thr Asp Val Pro Asp Gln Gly Leu Leu Cys Asp Leu Leu Trp Ser Asp  
 195 200 205

Pro Asp Lys Asp Val Leu Gly Trp Gly Glu Asn Asp Arg Gly Val Ser  
 210 215 220

Phe Thr Phe Gly Ala Glu Val Val Ala Lys Phe Leu His Lys His Asp  
 225 230 235 240

Leu Asp Leu Ile Cys Arg Ala His Gln Val Val Glu Asp Gly Tyr Glu  
 245 250 255

Phe Phe Ala Lys Arg Gln Leu Val Thr Leu Phe Ser Ala Pro Asn Tyr  
 260 265 270

Cys Gly Glu Phe Asp Asn Ala Gly Ala Met Met Ser Val Asp Glu Thr  
 275 280 285

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Asn Ala Thr Arg Pro Val Thr Pro Pro Arg Gly Met Ile Thr Lys Gln  
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 <212> DNA  
 <213> Homo sapiens

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<210> 32  
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Asn Glu Ile Arg Gly Leu Cys Leu Lys Ser Arg Glu Ile Phe Leu Ser  
 35 40 45

Gln Pro Ile Leu Leu Glu Leu Glu Ala Pro Leu Lys Ile Cys Gly Asp  
50 55 60

Ile His Gly Gln Tyr Tyr Asp Leu Leu Arg Leu Phe Glu Tyr Gly Gly  
65 70 75 80

Phe Pro Pro Glu Ser Asn Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg  
85 90 95

Gly Lys Gln Ser Leu Glu Thr Ile Cys Leu Leu Leu Ala Tyr Lys Ile  
100 105 110

Lys Tyr Pro Glu Asn Phe Phe Leu Leu Arg Gly Asn His Glu Cys Ala  
115 120 125

Ser Ile Asn Arg Ile Tyr Gly Phe Tyr Asp Glu Cys Lys Arg Arg Tyr  
130 135 140

Asn Ile Lys Leu Trp Lys Thr Phe Thr Asp Cys Phe Asn Cys Leu Pro  
145 150 155 160

Ile Ala Ala Ile Val Asp Glu Lys Ile Phe Cys Cys His Gly Gly Leu  
165 170 175

Ser Pro Asp Leu Gln Ser Met Glu Gln Ile Arg Arg Ile Met Arg Pro  
180 185 190

Thr Asp Val Pro Asp Gln Gly Leu Leu Cys Asp Leu Leu Trp Ser Asp  
195 200 205

Pro Asp Lys Asp Val Gln Gly Trp Gly Glu Asn Asp Arg Gly Val Ser  
210 215 220

Phe Thr Phe Gly Ala Glu Val Val Ala Lys Phe Leu His Lys His Asp  
225 230 235 240

Leu Asp Leu Ile Cys Arg Ala His Gln Val Val Glu Asp Gly Tyr Glu  
245 250 255

Phe Phe Ala Lys Arg Gln Leu Val Thr Leu Phe Ser Ala Pro Asn Tyr  
260 265 270

Cys Gly Glu Phe Asp Asn Ala Gly Ala Met Met Ser Val Asp Glu Thr  
275 280 285

Leu Met Cys Ser Phe Gln Ile Leu Lys Pro Ala Asp Lys Asn Lys Gly  
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Lys Tyr Gly Gln Phe Ser Gly Leu Asn Pro Gly Gly Arg Pro Ile Thr  
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<212> DNA  
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<210> 35  
<211> 709  
<212> PRT  
<213> Murinae gen. sp.

<400> 35

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Met Lys Met Thr Tyr Asn Met Thr Phe Phe Pro Asn Leu Met Gly His  
35 40 45

Tyr Asp Gln Gly Ile Ala Ala Val Glu Met Gly His Phe Leu His Leu  
50 55 60

Ala Asn Leu Glu Cys Ser Pro Asn Ile Glu Met Phe Leu Cys Gln Ala  
65 70 75 80

Phe Ile Pro Thr Cys Thr Glu Gln Ile His Val Val Leu Pro Cys Arg  
85 90 95

Lys Leu Cys Glu Lys Ile Val Ser Asp Cys Lys Lys Leu Met Asp Thr  
100 105 110

Phe Gly Ile Arg Trp Pro Glu Glu Leu Glu Cys Asn Arg Leu Pro His  
115 120 125

Cys Asp Asp Thr Val Pro Val Thr Ser His Pro His Thr Glu Leu Ser  
130 135 140

Gly Pro Gln Lys Lys Ser Asp Gln Val Pro Arg Asp Ile Gly Phe Trp  
145 150 155 160

Cys Pro Lys His Leu Arg Thr Ser Gly Asp Gln Gly Tyr Arg Phe Leu  
165 170 175

Gly Ile Glu Gln Cys Ala Pro Pro Cys Pro Asn Met Tyr Phe Lys Ser  
180 185 190

Asp Glu Leu Asp Phe Ala Lys Ser Phe Ile Gly Ile Val Ser Ile Phe  
195 200 205

Cys Leu Cys Ala Thr Leu Phe Thr Phe Leu Thr Phe Leu Ile Asp Val  
210 215 220

Arg Arg Phe Arg Tyr Pro Glu Arg Pro Ile Ile Tyr Tyr Ser Val Cys  
225 230 235 240

Tyr Ser Ile Val Ser Leu Met Tyr Phe Val Gly Phe Leu Leu Gly Asn  
245 250 255

Ser Thr Ala Cys Asn Lys Ala Asp Glu Lys Leu Glu Leu Gly Asp Thr  
260 265 270

Val Val Leu Gly Ser Lys Asn Lys Ala Cys Ser Val Val Phe Met Phe  
275 280 285

Leu Tyr Phe Phe Thr Met Ala Gly Thr Val Trp Trp Val Ile Leu Thr  
290 295 300

Ile Thr Trp Phe Leu Ala Ala Gly Arg Lys Trp Ser Cys Glu Ala Ile  
305 310 315 320

Glu Gln Lys Ala Val Trp Phe His Ala Val Ala Trp Gly Ala Pro Gly  
325 330 335

Phe Leu Thr Val Met Leu Leu Ala Met Asn Lys Val Glu Gly Asp Asn  
340 345 350

Ile Ser Gly Val Cys Phe Val Gly Leu Tyr Asp Leu Asp Ala Ser Arg  
355 360 365

Tyr Phe Val Leu Leu Pro Leu Cys Leu Cys Val Phe Val Gly Leu Ser  
370 375 380

Leu Leu Leu Ala Gly Ile Ile Ser Leu Asn His Val Arg Gln Val Ile  
385 390 395 400

Gln His Asp Gly Arg Asn Gln Glu Lys Leu Lys Lys Phe Met Ile Arg  
405 410 415

Ile Gly Val Phe Ser Gly Leu Tyr Leu Val Pro Leu Val Thr Leu Leu  
420 425 430

Gly Cys Tyr Val Tyr Glu Leu Val Asn Arg Ile Thr Trp Glu Met Thr  
435 440 445

Trp Phe Ser Asp His Cys His Gln Tyr Arg Ile Pro Cys Pro Tyr Gln  
450 455 460

Ala Asn Pro Lys Ala Arg Pro Glu Leu Ala Leu Phe Met Ile Lys Tyr  
465 470 475 480

Leu Met Thr Leu Ile Val Gly Ile Ser Ala Val Phe Trp Val Gly Ser  
485 490 495

Lys Lys Thr Cys Thr Glu Trp Ala Gly Phe Phe Lys Arg Asn Arg Lys  
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Arg Asp Pro Ile Ser Glu Ser Arg Arg Val Leu Gln Glu Ser Cys Glu  
 515 520 525

Phe Phe Leu Lys His Asn Ser Lys Val Lys His Lys Lys Lys His Gly  
 530 535 540

Ala Pro Gly Pro His Arg Leu Lys Val Ile Ser Lys Ser Met Gly Thr  
 545 550 555 560

Ser Thr Gly Ala Thr Thr Asn His Gly Thr Ser Ala Met Ala Ile Ala  
 565 570 575

Asp His Asp Tyr Leu Gly Gln Glu Thr Ser Thr Glu Val His Thr Ser  
 580 585 590

Pro Glu Ala Ser Val Lys Glu Gly Arg Ala Asp Arg Ala Asn Thr Pro  
 595 600 605

Ser Ala Lys Asp Arg Asp Cys Gly Glu Ser Ala Gly Pro Ser Ser Lys  
 610 615 620

Leu Ser Gly Asn Arg Asn Gly Arg Glu Ser Arg Ala Gly Gly Leu Lys  
 625 630 635 640

Glu Arg Ser Asn Gly Ser Glu Gly Ala Pro Ser Glu Gly Arg Val Ser  
 645 650 655

Pro Lys Ser Ser Val Pro Glu Thr Gly Leu Ile Asp Cys Ser Thr Ser  
 660 665 670

Gln Ala Ala Ser Ser Pro Glu Pro Thr Ser Leu Lys Gly Ser Thr Ser  
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 <211> 2039  
 <212> DNA  
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 <211> 706  
 <212> PRT  
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<400> 37

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Met Lys Met Ala Tyr Asn Met Thr Phe Phe Pro Asn Leu Met Gly His  
 35 40 45

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Tyr Asp Gln Ser Ile Ala Ala Val Glu Met Glu His Phe Leu Pro Leu  
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Ala Asn Leu Glu Cys Ser Pro Asn Ile Glu Thr Phe Leu Cys Lys Ala  
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Phe Val Pro Thr Cys Ile Glu Gln Ile His Val Val Pro Pro Cys Arg  
85 90 95

Lys Leu Cys Glu Lys Val Tyr Ser Asp Cys Lys Lys Leu Ile Asp Thr  
100 105 110

Phe Gly Ile Arg Trp Pro Glu Glu Leu Glu Cys Asp Arg Leu Gln Tyr  
115 120 125

Cys Asp Glu Thr Val Pro Val Thr Phe Asp Pro His Thr Glu Phe Leu  
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Gly Pro Gln Lys Lys Thr Glu Gln Val Gln Arg Asp Ile Gly Phe Trp  
145 150 155 160

Cys Pro Arg His Leu Lys Thr Ser Gly Gly Gln Gly Tyr Lys Phe Leu  
165 170 175

Gly Ile Asp Gln Cys Ala Pro Pro Cys Pro Asn Met Tyr Phe Lys Ser  
180 185 190

Asp Glu Leu Glu Phe Ala Lys Ser Phe Ile Gly Thr Val Ser Ile Phe  
195 200 205

Cys Leu Cys Ala Thr Leu Phe Thr Phe Leu Thr Phe Leu Ile Asp Val  
210 215 220

Arg Arg Phe Arg Tyr Pro Glu Arg Pro Ile Ile Tyr Tyr Ser Val Cys  
225 230 235 240

Tyr Ser Ile Val Ser Leu Met Tyr Phe Ile Gly Phe Leu Leu Gly Asp  
245 250 255

Ser Thr Ala Cys Asn Lys Ala Asp Glu Lys Leu Glu Leu Gly Asp Thr  
260 265 270

Val Val Leu Gly Ser Gln Asn Lys Ala Cys Thr Val Leu Phe Met Leu  
275 280 285

Leu Tyr Phe Phe Thr Met Ala Gly Thr Val Trp Trp Val Ile Leu Thr  
290 295 300

Ile Thr Trp Phe Leu Ala Ala Gly Arg Lys Trp Ser Cys Glu Ala Ile  
305 310 315 320

Glu Gln Lys Ala Val Trp Phe His Ala Val Ala Trp Gly Thr Pro Gly  
325 330 335

Phe Leu Thr Val Met Leu Leu Ala Met Asn Lys Val Glu Gly Asp Asn  
340 345 350

Ile Ser Gly Val Cys Phe Val Gly Leu Tyr Asp Leu Asp Ala Ser Arg  
355 360 365

Tyr Phe Val Leu Leu Pro Leu Cys Leu Cys Val Phe Val Gly Leu Ser  
370 375 380

Leu Leu Leu Ala Gly Ile Ile Ser Leu Asn His Val Arg Gln Val Ile  
385 390 395 400

Gln His Asp Gly Arg Asn Gln Glu Lys Leu Lys Lys Phe Met Ile Arg  
405 410 415

Ile Gly Val Phe Ser Gly Leu Tyr Leu Val Pro Leu Val Thr Leu Leu  
420 425 430

Gly Cys Tyr Val Tyr Glu Gln Val Asn Arg Ile Thr Trp Glu Ile Thr  
435 440 445

Trp Val Ser Asp His Cys Arg Gln Tyr His Ile Pro Cys Pro Tyr Gln  
450 455 460

Ala Lys Ala Lys Ala Arg Pro Glu Leu Ala Leu Phe Met Ile Lys Tyr  
465 470 475 480

Leu Met Thr Leu Ile Val Gly Ile Ser Ala Val Phe Trp Val Gly Ser  
485 490 495

Lys Lys Thr Cys Thr Glu Trp Ala Gly Phe Phe Lys Arg Asn Arg Lys  
500 505 510

Arg Asp Pro Ile Ser Glu Ser Arg Arg Val Leu Gln Glu Ser Cys Glu  
515 520 525

Phe Phe Leu Lys His Asn Ser Lys Val Lys His Lys Lys His Tyr

530 535 540

Lys Pro Ser Ser His Lys Leu Lys Val Ile Ser Lys Ser Met Gly Thr  
545 550 555 560

Ser Thr Gly Ala Thr Ala Asn His Gly Thr Ser Ala Val Ala Ile Thr  
565 570 575

Ser His Asp Tyr Leu Gly Gln Glu Thr Leu Thr Glu Ile Gln Thr Ser  
580 585 590

Pro Glu Thr Ser Met Arg Glu Val Lys Ala Asp Gly Ala Ser Thr Pro  
595 600 605

Arg Leu Arg Glu Gln Asp Cys Gly Glu Pro Ala Ser Pro Ala Ala Ser  
610 615 620

Ile Ser Arg Leu Ser Gly Glu Gln Val Asp Gly Lys Gly Gln Ala Gly  
625 630 635 640

Ser Val Ser Glu Ser Ala Arg Ser Glu Gly Arg Ile Ser Pro Lys Ser  
645 650 655

Asp Ile Thr Asp Thr Gly Leu Ala Gln Ser Asn Asn Leu Gln Val Pro  
660 665 670

Ser Ser Ser Glu Pro Ser Ser Leu Lys Gly Ser Thr Ser Leu Leu Val  
675 680 685

His Pro Val Ser Gly Val Arg Lys Glu Gln Gly Gly Cys His Ser  
690 695 700

Asp Thr  
705

<210> 38  
<211> 773  
<212> DNA  
<213> Murinae gen. sp.

<400> 38  
ctgaggtgct agcaccagcc tggttgtctc tggcgggcct gaagcaagca tggatcaaga  
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ggctgtggc aacgttgtgc tcctggccct tgcacccctc atcagcgtgg tccagaatgc  
120

gttctttgcc cacaagggtgg agcatgaaag caaggcgcatt aatgggagaa gcttccagag  
180

gaccgggact cttgcctttg agcgggtcta cactgccaaac cagaactgcg tagatgcgt  
240

cccaacttc cttgtggta ctcggactgc aggactactt tgcagccaag tccctgcagc  
300

cttcggcggga ctgatgtacc tggttgtag gcaaaaatac tttgtcggt atctggaga  
360

gagaactcag agcacccctg gctacatctt cggcaagcgg atcatcctgt tcctgttcct  
420

catgtccttc gccggatac tcaaccatta cctcatcttc ttcttcggaa gcgactttga  
480

gaactacatc agaacggtaa gcacgacgat ctccccctg ctctcatcc cctgattgt  
540

ggagacagag aaggacgctc accagatcaa tagagacgca tcataacgca acgcccgcga  
600

ggcttctgtct cctttcaag ctgttagatgc tgtcaatctt gctgccctcg gggctctgt  
660

gcatccgtta actttgttt tccgggaaga aaaatgtttt gtgctaagct ccaccctcg  
720

aatgcggcgg tgggccagga tttatgtcta catccagcct atacttctcc tgg  
773

<210> 39

<211> 852

<212> DNA

<213> Murinae gen. sp.

<400> 39

gaaaggctga ggtgctagca ccagcctgggt tgtctctggc gggcctgaag caagcatgga  
60

tcaagaggct gtgggcaacg ttgtgctcct ggcccttgct accctcatca gcgtggtcca  
120

gaatgtgttt tttgcccact atgtggagca tgaaagcaat gcgcataatg ggagaagctt  
180

ccagaggacc gggactcttg ccttgagcg ggtctacact gccaaccaga actgcgtaga  
240

tgcgtaaaaa actttcttg tggtaactctg gactgcagga ctactttgca gccaagtccc  
300

tgccgccttc gccggactga tgtacctgtt tgtgaggcaa aaatactttg tcggctatct  
360

ggagagagaga actcagagca cccctggcta catttcggc aagcggatca tcctgttcct  
420

gttcctcatg tccttcggcg ggatactcaa ccattacctc atcttcttct tcggaagcga  
480

ctttgagaac tacatcagaa cggttaagcac gacgatctcc ccgctgcttc tcataccctg  
540

attgctggag acagagaagg acgctcacca gatcaataga gacgcacat aacgcaacgc  
600

cgcgaaggct tctgctcctc ttcaagctgt agatgctgac aatcttgcgt ccctcgcccc  
660

tctgtggcat ccgttaactt tgctttccg ggaagaaaaa tgtcttgac tagctccacc  
720

cctcgaatgc ggcgggtggcc caggattat tgtctacatc cagcctatac ttctcctggc  
780

ttatcctgct ttctgaagat gtcttgaaat cagacacgtg tttcctaaa ataaaggta  
840

tagacaaaaat tt  
852

<210> 40

<211> 161

<212> PRT

<213> Murinae gen. sp.

<400> 40

Met Asp Gln Glu Ala Val Gly Asn Val Val Leu Leu Ala Leu Val Thr  
1 5 10 15

Leu Ile Ser Val Val Gln Asn Val Phe Phe Ala His Tyr Val Glu His  
20 25 30

Glu Ser Asn Ala His Asn Gly Arg Ser Phe Gln Arg Thr Gly Thr Leu  
35 40 45

Ala Phe Glu Arg Val Tyr Thr Ala Asn Gln Asn Cys Val Asp Ala Tyr  
50 55 60

Pro Thr Phe Leu Val Val Leu Trp Thr Ala Gly Leu Leu Cys Ser Gln  
65 70 75 80

Val Pro Ala Ala Phe Ala Gly Leu Met Tyr Leu Phe Val Arg Gln Lys  
85 90 95

Tyr Phe Val Gly Tyr Leu Gly Glu Arg Thr Gln Ser Thr Pro Gly Tyr  
100 105 110

Ile Phe Gly Lys Arg Ile Ile Leu Phe Leu Phe Leu Met Ser Phe Ala  
115 120 125

Gly Ile Leu Asn His Tyr Leu Ile Phe Phe Phe Gly Ser Asp Phe Glu  
130 135 140

Asn Tyr Ile Arg Thr Val Ser Thr Thr Ile Ser Pro Leu Leu Leu Ile  
145 150 155 160

Pro

<210> 41

<211> 873

<212> DNA

<213> Homo sapiens

<400> 41

acttccctt cctgtacagg gcaggttgtg cagctggagg cagagcagtc ctctctgggg  
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agcctgaagc aaacatggat caagaaaactg taggcaatgt tgcctgttg gccatcgta  
120

ccctcatcag cgtggtccag aatggattct ttgcccataa agtggagcac gaaagcagga  
180

cccagaatgg gaggagcttc cagaggaccg gaacacttgc cttttagcgg gtctacactg  
240

ccaaccagaa ctgtgttagat gcgtacccca ctttcctcgc tgcctctgg tctgcggggc  
300

tactttgcag ccaagtcct gctgcgttg ctggactgat gtacttgaaa gtgaggcaaa  
360

agactttgt cggttaccta ggagagagaa cgcagagcac ccctggctac atattttggaa  
420

aacgcacatcat acttttcctg ttccatgt ccgttgcgg catattcaac tattaccta  
480

tcttcctttt cgaaagtgc tttgaaaact acataaagac gatctccacc accatctccc  
540

ctctacttct cattccctaa ctctctgcgtg aatatgggt tgggtttctc atctaata  
600

tacctacaag tcatcataat tcagcttttgc agagcattct gctttttttt agatggctgt  
660

aaatctatttgc gcatctggg cttcacagct tgagttaacc ttgtttttcc gggaaacaaaa  
720

tgatgtcatg tcagctccgc cccttgaaca tgaccgtggc cccaaatttg ctattccat  
780

gcattttgtt tgtttcttca cttatcctgt tctctgaaga tgttttgtga ccaggtttgt  
840

gttttcttaa aataaaatgc agagacatgt ttt  
873

<210> 42  
<211> 161  
<212> PRT  
<213> Homo sapiens  
  
<400> 42

Met Asp Gln Glu Thr Val Gly Asn Val Val Leu Leu Ala Ile Val Thr  
1 5 10 15

Leu Ile Ser Val Val Gln Asn Gly Phe Phe Ala His Lys Val Glu His  
20 25 30

Glu Ser Arg Thr Gln Asn Gly Arg Ser Phe Gln Arg Thr Gly Thr Leu  
35 40 45

Ala Phe Glu Arg Val Tyr Thr Ala Asn Gln Asn Cys Val Asp Ala Tyr  
50 55 60

Pro Thr Phe Leu Ala Val Leu Trp Ser Ala Gly Leu Leu Cys Ser Gln  
65 70 75 80

Val Pro Ala Ala Phe Ala Gly Leu Met Tyr Leu Phe Val Arg Gln Lys  
85 90 95

Tyr Phe Val Gly Tyr Leu Gly Glu Arg Thr Gln Ser Thr Pro Gly Tyr  
100 105 110

Ile Phe Gly Lys Arg Ile Ile Leu Phe Leu Phe Leu Met Ser Val Ala  
115 120 125

Gly Ile Phe Asn Tyr Tyr Leu Ile Phe Phe Phe Gly Ser Asp Phe Glu  
130 135 140

Asn Tyr Ile Lys Thr Ile Ser Thr Thr Ile Ser Pro Leu Leu Ile  
145 150 155 160

Pro

<210> 43  
<211> 803  
<212> DNA  
<213> Murinae gen. sp.  
  
<400> 43

PRJU03-12-05

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tcatggattt tctcgaccag aaaatcagac tattttcctg aataatctac tagaaacttt  
120  
tacggaacac atttcatgtt tccttgaag agttaagaga agaaaagtatt tgtaagaaca  
180  
ggaaaagaaa caaatacttt gcaaataaac tggctgctgc tgtgaccaca tctgaatgc  
240  
aaaggcgatc gatcaagcgc tgccggacaaa aggccctcctg taagctgcac tgccctgacaa  
300  
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360  
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420  
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480  
tgtcagattt acttttcgtc ttactttgc catttcggat ttttacttt gcaacacgga  
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600  
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780  
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803

<210> 44  
<211> 1849  
<212> DNA  
<213> Murinae gen. sp.

<400> 44  
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cgcgaaacatgccttagaatt tatctggat cccttaaacg actgcctatc gccgtccgga  
120  
atcaatgttag aaatacaaag tttgagaata aaaagaagga agaagtaccc gaggacgacg  
180  
ggcggacgga cgcacggcga gtgtttgtga ctgaagtaaa gctggtttgg accctggcgg  
240

ctgaaggcaca agtttccacg cggaactggc tggccgact tggAACAGTT tttccttaca  
300  
ctttcagctt tatgggttgg ctcccttgac tgcattttct gtcagttAAC taaaactccag  
360  
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420  
tttacggAAC acatttcatg ttcccttga agagttaaga gaagaaagta tttgtaaagaa  
480  
caggAAAAGA aacaaataact ttgcaaataa actggctgct gctgtgacca catctgaata  
540  
gcaaaggcga tcgatcaAGC gctgcggaca aaaggcctcc tggAAgctgc actgcctgac  
600  
aatggtaAGC tccaatggct cccagtgcCC ttatgacgac tccttaagt acactctgta  
660  
cggtgcATG ttcagcatgg tcttcgtgct tggctgata tccaaCTgtg ttgcgatata  
720  
catttcatc tggccctca aagtgagaaa tgaaactaca acgtacatga ttaacctggc  
780  
aatgtcAGAT ttactttcg tctttacttt gccatttcgg atttttact ttgcaacacg  
840  
gaattggcca tttggagatc tactctgtaa gatttcagta atgctgttt acaccaata  
900  
gtatggAAAGC attctgttct taacctgtat cagtgttagat cgatttctgg caattgtcta  
960  
cccatTTAAG tcaaAGACTT taagaacgaa acgaaatgca aagatcgTTT gcattgtgt  
1020  
gtggttcaca gtgatggag gaagtgcGCC tgcagTTTC ttcaGtcga cccactctca  
1080  
ggggAACAAcT acctcagaag cctgcttga gaactttcca gggccacat ggAAAactta  
1140  
tctctccagg attgtgattt tcattgaaat agtgggcttt ttatccctc tcattttgaa  
1200  
cgtaacttgc tctagtagatgg tgctaaAGAAC ttAAATAAA CCTGTTACAT taagttagaa  
1260  
caaaatgaac aaaactaagg tttaaaaaat gatTTTgtc cacttggtca tcttctgttt  
1320  
ctgttttgc ccctacaaca tcaacctcat tttgtactcg ctcatgagga cacagacTT  
1380  
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1440

tgttccaac tgctgcttg accctattgt ttactacttc acctcagaca caattcagaa  
1500

ctcaataaaa atgaaaaact ggtcggttag aagaagtgc tccaggttct ctgaaggta  
1560

ggcactgag aatttatcc aacacaacct acagaccta aaaaataaga tatttgataa  
1620

tgaatctgca atataagctg cctgactaag ccactggac tgctccgtgt tcaactgtga  
1680

aaactgtgtt ctggaaact atctctccgg ctccaacaga aaatatttt aaaggaagtt  
1740

tgtgtctgat gtgttaaaca ttaaaatata ttctattttt gtatgcacgc cattttactt  
1800

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1849

<210> 45

<211> 316

<212> PRT

<213> Murinae gen. sp.

<400> 45

Asp Asp Ser Phe Lys Tyr Thr Leu Tyr Gly Cys Met Phe Ser Met Val  
1 5 10 15

Phe Val Leu Gly Leu Ile Ser Asn Cys Val Ala Ile Tyr Ile Phe Ile  
20 25 30

Cys Ala Leu Lys Val Arg Asn Glu Thr Thr Tyr Met Ile Asn Leu  
35 40 45

Ala Met Ser Asp Leu Leu Phe Val Phe Thr Leu Pro Phe Arg Ile Phe  
50 55 60

Tyr Phe Ala Thr Arg Asn Trp Pro Phe Gly Asp Leu Leu Cys Lys Ile  
65 70 75 80

Ser Val Met Leu Phe Tyr Thr Asn Met Tyr Gly Ser Ile Leu Phe Leu  
85 90 95

Thr Cys Ile Ser Val Asp Arg Phe Leu Ala Ile Val Tyr Pro Phe Lys  
100 105 110

Ser Lys Thr Leu Arg Thr Lys Arg Asn Ala Lys Ile Val Cys Ile Ala  
115 120 125

Val Trp Phe Thr Val Met Gly Gly Ser Ala Pro Ala Val Phe Phe Gln  
 130 135 140

Ser Thr His Ser Gln Gly Asn Asn Thr Ser Glu Ala Cys Phe Glu Asn  
 145 150 155 160

Phe Pro Ala Ala Thr Trp Lys Thr Tyr Leu Ser Arg Ile Val Ile Phe  
 165 170 175

Ile Glu Ile Val Gly Phe Phe Ile Pro Leu Ile Leu Asn Val Thr Cys  
 180 185 190

Ser Ser Met Val Leu Arg Thr Leu Asn Lys Pro Val Thr Leu Ser Arg  
 195 200 205

Ser Lys Met Asn Lys Thr Lys Val Leu Lys Met Ile Phe Val His Leu  
 210 215 220

Val Ile Phe Cys Phe Cys Phe Val Pro Tyr Asn Ile Asn Leu Ile Leu  
 225 230 235 240

Tyr Ser Leu Met Arg Thr Gln Thr Phe Val Asn Cys Ser Val Val Ala  
 245 250 255

Ala Val Arg Thr Met Tyr Pro Ile Thr Leu Cys Ile Ala Val Ser Asn  
 260 265 270

Cys Cys Phe Asp Pro Ile Val Tyr Tyr Phe Thr Ser Asp Thr Ile Gln  
 275 280 285

Asn Ser Ile Lys Met Lys Asn Trp Ser Val Arg Arg Ser Asp Ser Arg  
 290 295 300

Phe Ser Glu Val Gln Gly Thr Glu Asn Phe Ile Gln  
 305 310 315

<210> 46  
 <211> 1035  
 <212> DNA  
 <213> Homo sapiens

<400> 46  
 atggtaagcg ttaacagctc ccactgcttc tataatgact cctttaagta cactttgtat  
 60

gggtgcatgt tcagcatggc gtttgcgtt gggtaataat ccaattgtgt tgccatatac  
 120

atttcatct gcgtcccaa agtccgaaat gaaactacaa ottacatgtat taacttggca  
180

atgtcagact tgcttttgtt ttttacttta cccttcagga ttttttactt cacaacacgg  
240

aattggccat ttggagatTTT actttgtaaG atttctgtGA tgctgtttta taccaacatg  
300

tacggaagca ttctgttctt aacctgtatt agttagatc gatttctggc aattgtctac  
360

ccatTTAAGT caaAGACTCT aagaaccaaa agaaatgcaa agattgtttg cactggcgtg  
420

tggTTaactg tgatcggagg aagtgcaccc gccgTTTTG ttcaGtctac ccactctcag  
480

ggtAACAAATg cctcagaAGC ctgctttgaa aattttccag aagccacatg gaaaacatAT  
540

ctctcaagga ttgtAAtttt catcgaaata gTgggatttt ttatTCCTt aattttAAat  
600

gtaacttGtt ctatGatGgt gctaaaaact ttaaccaac ctgttacatt aagttagaAGC  
660

aaaataAAaca aaactaaggT tttaaaaATg atttttgtac atttgatcat attctgtttc  
720

tgttttgttC cttacaatAT caatcttatt ttatattctc ttgtgagaac acaaAcattt  
780

gttaattgct cagtagtggc agcagtaagg acaatgtacc caatcactct ctgtattgct  
840

gtttccaact gttgtttga ccctatagtt tactacttta catggacac aattcagaat  
900

tcaataaaaaa tgaaaaactg gtGtGtcagg agaagtgact tcagattctc tgaagttcat  
960

ggtgcagaga attttattca gcataaccta cagaccttaa aaagtaagat atttgacaat  
1020

gaatctgctg cctga  
1035

<210> 47  
<211> 344  
<212> PRT  
<213> Homo sapiens

<400> 47

Met Val Ser Val Asn Ser Ser His Cys Phe Tyr Asn Asp Ser Phe Lys  
1 5 10 15

Tyr Thr Leu Tyr Gly Cys Met Phe Ser Met Val Phe Val Leu Gly Leu

20

25

30

Ile Ser Asn Cys Val Ala Ile Tyr Ile Phe Ile Cys Val Leu Lys Val  
35 40 45

Arg Asn Glu Thr Thr Tyr Met Ile Asn Leu Ala Met Ser Asp Leu  
50 55 60

Leu Phe Val Phe Thr Leu Pro Phe Arg Ile Phe Tyr Phe Thr Thr Arg  
65 70 75 80

Asn Trp Pro Phe Gly Asp Leu Leu Cys Lys Ile Ser Val Met Leu Phe  
85 90 95

Tyr Thr Asn Met Tyr Gly Ser Ile Leu Phe Leu Thr Cys Ile Ser Val  
100 105 110

Asp Arg Phe Leu Ala Ile Val Tyr Pro Phe Lys Ser Lys Thr Leu Arg  
115 120 125

Thr Lys Arg Asn Ala Lys Ile Val Cys Thr Gly Val Trp Leu Thr Val  
130 135 140

Ile Gly Gly Ser Ala Pro Ala Val Phe Val Gln Ser Thr His Ser Gln  
145 150 155 160

Gly Asn Asn Ala Ser Glu Ala Cys Phe Glu Asn Phe Pro Glu Ala Thr  
165 170 175

Trp Lys Thr Tyr Leu Ser Arg Ile Val Ile Phe Ile Glu Ile Val Gly  
180 185 190

Phe Phe Ile Pro Leu Ile Leu Asn Val Thr Cys Ser Ser Met Val Leu  
195 200 205

Lys Thr Leu Thr Lys Pro Val Thr Leu Ser Arg Ser Lys Ile Asn Lys  
210 215 220

Thr Lys Val Leu Lys Met Ile Phe Val His Leu Ile Ile Phe Cys Phe  
225 230 235 240

Cys Phe Val Pro Tyr Asn Ile Asn Leu Ile Leu Tyr Ser Leu Val Arg  
245 250 255

Thr Gln Thr Phe Val Asn Cys Ser Val Val Ala Ala Val Arg Thr Met  
260 265 270

Tyr Pro Ile Thr Leu Cys Ile Ala Val Ser Asn Cys Cys Phe Asp Pro  
 275 280 285

Ile Val Tyr Tyr Phe Thr Ser Asp Thr Ile Gln Asn Ser Ile Lys Met  
 290 295 300

Lys Asn Trp Ser Val Arg Arg Ser Asp Phe Arg Phe Ser Glu Val His  
 305 310 315 320

Gly Ala Glu Asn Phe Ile Gln His Asn Leu Gln Thr Leu Lys Ser Lys  
 325 330 335

Ile Phe Asp Asn Glu Ser Ala Ala  
 340

<210> 48  
 <211> 814  
 <212> DNA  
 <213> Murinae gen. sp.

<400> 48  
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ggccaggcct agcacatgta cctcacagac caactggcaa gcagcttca gggagctcga  
 120

tccccaaaca gccagtcacc acctctgtcc cctcttcaact gttggtcgtc agactgcctg  
 180

agtggacagc aggctggcgt cgttgtatcc tcacttcctt cctctgactg gcttgctctt  
 240

gtctctcagt ctttcatccc aggcagctgc ctgaggtagg tgaggaggat ggtgagccag  
 300

gcaggtctac aataaaggca gctctgtccg gtccttctg gtcgtgagt gtcacccggcc  
 360

tgaaagactg agggaaatggc tccccctctt cctccccgtc tttccccagt tccttcctta  
 420

tgtggccca tgtgcccagg gagttggaag catcagggag accctcttag tgggggaag  
 480

gaagtcagag accattgaca cagtgaagag gcaggatcat gtgttggaaag cctgttagca  
 540

ggaccaaggt gactcttggg agagactctt gtggacacag gccgtggtg cttgtcagac  
 600

cttaaagggt ccaggcccac ccctgccagg atccctggtc tgctttctcc aggacacact  
 660

gggacactgc ttagtaatga gcagcttatt acacacaatg ggaagagggg cagagaggc  
720

tgtgtcggtt gagtctcggc tggactgaa gtttgcata agtagtggtt gtacatccag  
780

gagcctggct acctgtctt accccttgaa ggac  
814

<210> 49  
<211> 1164  
<212> DNA  
<213> Murinae gen. sp.

<400> 49  
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agctgtgggg ctcccaactt ccggcaggtg cggggaggcc tccctgtgtt tggcatggg  
120

cagcccagcc tcttggggtt caggagggtc ctgcagaaac tccagacgga cggactcaag  
180

gagtgcatta tcttctgcgt gcgggaggag cctgtggtgt tcttgcgcgc tgaggaggac  
240

tttgtgtctt acacacctcg agacaaggag agcttcatg agaacctcg ggaccctagt  
300

ccaggggtca aggctgagaa tctggagctg gccatccaga aagagatcca tgactttgcc  
360

caattgagag ataatgtgtt ccacgtatac cacaacacag aggacctgcg cggggagccg  
420

cacaccgtgg ccattccgagg tgaggatggc gtgtgcgtga ccgaggaggt gtttaagccg  
480

ccgctcttcc tgcagccac ctacagatac caccgcctcc cttgccaga gcaagggcc  
540

cccttggaaag cccagtttga tgcctttgtc agcgtttttc gggagacccc cagccttctg  
600

ccactcagag ataaccacgg gcctctgcct gcctcctgt tcagctgcc atcaggtgt  
660

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APRIL 2008 - 12 - 008

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<400> 50

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Gln Lys Glu Ile His Asp Phe Ala Gln Leu Arg Asp Asn Val Tyr His  
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Ser Phe Arg Pro Glu Ser Arg Gly Gln Glu Cys Gly Ser Gln Gln Ala  
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Val Gln Gln Arg Ala Leu Trp Ser Leu Glu Leu Tyr Phe Tyr Leu Leu  
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Ser Phe Ser Arg Trp Leu Cys Thr His Pro Glu Leu Tyr Arg Leu Leu  
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PPU103-12-05

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 <213> Homo sapiens  
  
 <400> 52

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Asp Ser Arg His Ser Val Ser Ile His Ser Phe Gln Ser Thr Ser Leu  
 35 40 45

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His Asn Ser Lys Ala Lys Ser Ile Ile Pro Asn Lys Val Ala Pro Val  
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Val Ile Thr Tyr Asn Cys Lys Glu Glu Phe Gln Ile His Asp Glu Leu  
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Leu Lys Ala His Tyr Thr Leu Gly Arg Leu Ser Asp Asn Thr Pro Glu  
85 90 95

His Tyr Leu Val Gln Gly Arg Tyr Phe Leu Val Arg Asp Val Thr Glu  
100 105 110

Lys Met Asp Val Leu Gly Thr Val Gly Ser Cys Gly Ala Pro Asn Phe  
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Arg Gln Val Gln Gly Gly Leu Thr Val Phe Gly Met Gly Gln Pro Ser  
130 135 140

Leu Ser Gly Phe Arg Arg Val Leu Gln Lys Leu Gln Lys Asp Gly His  
145 150 155 160

Arg Glu Cys Val Ile Phe Cys Val Arg Glu Glu Pro Val Leu Phe Leu  
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Arg Ala Asp Glu Asp Phe Val Ser Tyr Thr Pro Arg Asp Lys Gln Asn  
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195 200 205

Leu Glu Leu Ala Ile Arg Lys Glu Ile His Asp Phe Ala Gln Leu Ser  
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Glu Asn Thr Tyr His Val Tyr His Asn Thr Glu Asp Leu Trp Gly Glu  
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Pro His Ala Val Ala Ile His Gly Glu Asp Asp Leu His Val Thr Glu  
245 250 255

Glu Val Tyr Lys Arg Pro Leu Phe Leu Gln Pro Thr Tyr Arg Tyr His  
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Arg Leu Pro Leu Pro Glu Gln Gly Ser Pro Leu Glu Ala Gln Leu Asp  
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Ala Phe Val Ser Val Leu Arg Glu Thr Pro Ser Leu Leu Gln Leu Arg  
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Asp Ala His Gly Pro Pro Pro Ala Leu Val Phe Ser Cys Gln Met Gly  
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Val Gly Arg Thr Asn Leu Gly Met Val Leu Gly Thr Leu Ile Leu Leu  
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His Arg Ser Gly Thr Thr Ser Gln Pro Glu Ala Ala Pro Thr Gln Ala  
340 345 350

Lys Pro Leu Pro Met Glu Gln Phe Gln Val Ile Gln Ser Phe Leu Arg  
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Met Val Pro Gln Gly Arg Arg Met Val Glu Glu Val Asp Arg Ala Ile  
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Thr Ala Cys Ala Glu Leu His Asp Leu Lys Glu Val Val Leu Glu Asn  
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Gln Lys Lys Leu Glu Gly Ile Arg Pro Glu Ser Pro Ala Gln Gly Ser  
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Gly Ser Arg His Ser Val Trp Gln Arg Ala Leu Trp Ser Leu Glu Arg  
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Tyr Phe Tyr Leu Ile Leu Phe Asn Tyr Tyr Leu His Glu Gln Tyr Pro  
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Leu Ala Phe Ala Leu Ser Phe Ser Arg Trp Leu Cys Ala His Pro Glu  
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Leu Tyr Arg Leu Pro Val Thr Leu Ser Ser Ala Gly Pro Val Ala Pro  
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Arg Asp Leu Ile Ala Arg Gly Ser Leu Arg Glu Asp Asp Leu Val Ser  
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Pro Asp Ala Leu Ser Thr Val Arg Glu Met Asp Val Ala Asn Phe Arg  
500 505 510

Arg Val Pro Arg Met Pro Ile Tyr Gly Thr Ala Gln Pro Ser Ala Lys  
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Ala Leu Gly Ser Ile Leu Ala Tyr Leu Thr Asp Ala Lys Arg Arg Leu

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535

540

Arg Lys Val Val Trp Val Ser Leu Arg Glu Glu Ala Val Leu Glu Cys  
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Asp Gly His Thr Tyr Ser Leu Arg Trp Pro Gly Pro Pro Val Ala Pro  
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Asp Gln Leu Glu Thr Leu Glu Ala Gln Leu Lys Ala His Leu Ser Glu  
580 585 590

Pro Pro Pro Gly Lys Glu Gly Pro Leu Thr Tyr Arg Phe Gln Thr Cys  
595 600 605

Leu Thr Met Gln Glu Val Phe Ser Gln His Arg Arg Ala Cys Pro Gly  
610 615 620

Leu Thr Tyr His Arg Ile Pro Met Pro Asp Phe Cys Ala Pro Arg Glu  
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Glu Asp Phe Asp Gln Leu Leu Glu Ala Leu Arg Ala Ala Leu Ser Lys  
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Asp Pro Gly Thr Gly Phe Val Phe Ser Cys Leu Ser Gly Gln Gly Arg  
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Thr Thr Thr Ala Met Val Val Ala Val Leu Ala Phe Trp His Ile Gln  
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Gly Phe Pro Glu Val Gly Glu Glu Leu Val Ser Val Pro Asp Ala  
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Lys Phe Thr Lys Gly Glu Phe Gln Val Val Met Lys Val Val Gln Leu  
705 710 715 720

Leu Pro Asp Gly His Arg Val Lys Lys Glu Val Asp Ala Ala Leu Asp  
725 730 735

Thr Val Ser Glu Thr Met Thr Pro Met His Tyr His Leu Arg Glu Ile  
740 745 750

Ile Ile Cys Thr Tyr Arg Gln Ala Lys Ala Ala Lys Glu Ala Gln Glu  
755 760 765

Met Arg Arg Leu Gln Leu Arg Ser Leu Gln Tyr Leu Glu Arg Tyr Val  
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Cys Leu Ile Leu Phe Asn Ala Tyr Leu His Leu Glu Lys Ala Asp Ser  
785 790 795 800

Trp Gln Arg Pro Phe Ser Thr Trp Met Gln Glu Val Ala Ser Lys Ala  
805 810 815

Gly Ile Tyr Glu Ile Leu Asn Glu Leu Gly Phe Pro Glu Leu Glu Ser  
820 825 830

Gly Glu Asp Gln Pro Phe Ser Arg Leu Arg Tyr Arg Trp Gln Glu Gln  
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<212> DNA

<213> Murinae gen. sp.

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<210> 55

<211> 306

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<213> Murinae gen. sp.

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Gln Thr Cys Tyr His Pro Ile Arg Gly Asp Gln Leu Ala Leu Leu Gly  
35 40 45

Arg Arg Thr Tyr Pro Arg Pro His Glu Tyr Leu Ser Pro Ala Asp Leu  
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Pro Lys Asn Trp Asp Trp Arg Asn Val Asn Gly Val Asn Tyr Ala Ser  
65 70 75 80

Val Thr Arg Asn Gln His Ile Pro Gln Tyr Cys Gly Ser Cys Trp Ala  
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His Gly Ser Thr Ser Ala Met Ala Asp Arg Ile Asn Ile Lys Arg Lys  
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Gly Ala Trp Pro Ser Ile Leu Leu Ser Val Gln Asn Val Ile Asp Cys

PDB ID: 1Q2F

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Ala Lys Asp Gln Asp Cys Asp Lys Phe Asn Gln Cys Gly Thr Cys Thr		
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Glu Phe Lys Glu Cys His Thr Ile Gln Asn Tyr Thr Leu Trp Arg Val		
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Gly Asp Tyr Gly Ser Leu Ser Gly Arg Glu Lys Met Met Ala Glu Ile		
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Tyr Ala Asn Gly Pro Ile Ser Cys Gly Ile Met Ala Thr Glu Met Met		
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<212> DNA		
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Cys Tyr Arg Pro Leu Arg Gly Asp Gly Leu Ala Pro Leu Gly Arg Ser  
35 40 45

Thr Tyr Pro Arg Pro His Glu Tyr Leu Ser Pro Ala Asp Leu Pro Lys  
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Ser Trp Asp Trp Arg Asn Val Asp Gly Val Asn Tyr Ala Ser Ile Thr  
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Arg Asn Gln His Ile Pro Gln Tyr Cys Gly Ser Cys Trp Ala His Ala  
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Ser Thr Ser Ala Met Ala Asp Arg Ile Asn Ile Lys Arg Lys Gly Ala  
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Trp Pro Ser Thr Leu Leu Ser Val Gln Asn Val Ile Asp Cys Gly Asn  
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Ala Gly Ser Cys Glu Gly Gly Asn Asp Leu Ser Val Trp Asp Tyr Ala  
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Asp Gln Glu Cys Asp Lys Phe Asn Gln Cys Gly Thr Cys Asn Glu Phe

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Lys Glu Cys His Ala Ile Arg Asn Tyr Thr Leu Trp Arg Val Gly Asp  
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Tyr Gly Ser Leu Ser Gly Arg Glu Lys Met Met Ala Glu Ile Tyr Ala  
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Asn Gly Pro Ile Ser Cys Gly Ile Met Ala Thr Glu Arg Leu Ala Asn  
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Tyr Thr Gly Gly Ile Tyr Ala Glu Tyr Gln Asp Thr Thr Tyr Ile Asn  
225 230 235 240

His Val Val Ser Val Ala Gly Trp Gly Ile Ser Asp Gly Thr Glu Tyr  
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Trp Ile Val Arg Asn Ser Trp Gly Glu Pro Trp Gly Glu Arg Gly Trp  
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Leu Arg Ile Val Thr Ser Thr Tyr Lys Asp Gly Lys Gly Ala Arg Tyr  
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Asn Leu Ala Ile Glu Glu His Cys Thr Phe Gly Asp Pro Ile Val  
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